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Micro-magnetic resonance imaging and embryological analysis of wild-type and pma mutant mice with clubfoot

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Abstract
Gross similarities between the external appearance of the hind limbs of the peroneal muscle atrophy (pma) mouse mutant and congenital talipes equinovarus (CTEV), a human disorder historically referred to as ‘clubfoot’, suggested that this mutant could be a useful model. We used micro-magnetic resonance imaging to visualize the detailed anatomy of the hind limb defect in mutant pma mice and performed 3D comparisons between mutant and wild-type hind limbs. We found that the pma foot demonstrates supination (i.e. adduction and inversion of the mid foot and fore foot together with plantar flexion of the ankle and toes) and that the tibiale and distal tarsals display 3D abnormalities in positioning. The size and shape of the tibia, fibula, tarsal and metatarsal bones are similar to the wild-type. Hypoplasia of the muscles in the antero-lateral (peroneal) compartment was also demonstrated. The resemblance of these features to those seen in CTEV suggests that the pma mouse is a possibly useful model for the human condition. To understand how the observed deformities in the pma mouse hind foot arise during embryonic development, we followed the process of foot rotation in both wild-type and pma mutant mice. Rotation of the hind foot in mouse embryos of wild-type strains (CD-1 and C57/Black) occurs from embryonic day 14.5 onwards with rotation in C57/Black taking longer. In embryos from both strains, rotation of the right hind foot more commonly precedes rotation of the left. In pma mutants, the initiation of rotation is often delayed and rotation is slower and does not reach completion. If the usefulness of the pma mutant as a model is confirmed, then these findings on pma mouse embryos, when extrapolated to humans, would support a long-standing hypothesis that CTEV is due to the failure of completion of the normal process of rotation and angulation, historically known as the ‘arrested development hypothesis’.

Key words anatomy; clubfoot; congenital talipes equinovarus; embryo; hind limb; limb development; magnetic resonance imaging; micro-magnetic resonance imaging; mouse; musculoskeletal; peroneal muscle atrophy.

Introduction
Congenital talipes equinovarus (CTEV), classically referred to as ‘clubfoot’, is a 3D malformation that is immediately recognisable at birth, in which the ankle is in the plantar flexed (equinus) position, the heel is inverted (varus) and the mid foot and fore foot are inverted and adducted (varus). Several recent studies have described the detailed anatomy of both skeletal and soft tissues in clubfoot in human fetuses between the 25th and 37th week of gestation (Fritsch & Eggers, 1999; Gruber et al., 2001; Windisch et al., 2007a,b) and a 3D model for clubfoot has been made.
using micro-computed tomography to visualize the shape of the individual bones of the foot (Windisch et al., 2007c). The prevalence of CTEV varies around the world from 0.3 to 7 per 1000 live births and it may be bilateral or unilateral (reviewed Miedzybrodzka, 2003; Cardy et al., 2007). Where unilateral, CTEV affects the right side slightly more often than the left (Miedzybrodzka, 2003).

Various aetiopathogenic mechanisms for CTEV have been proposed, such as uterine restriction, malformation of bones, joints, connective tissue or limb vasculature, and impairment of myogenesis and neurogenesis in the limbs (Miedzybrodzka, 2003). Genetic predisposition is clearly implicated by twin studies and there is increased prevalence in close relatives (reviewed by Miedzybrodzka, 2003). Kawashima & Ulthoff (1990) (see also England, 1996 for illustrations of lower limb development) give a detailed description of the rotation of the human embryonic lower limb. At the beginning of the 8th week the footplate is aligned with the long axis of the lower leg in a slightly supinated position. The foot then goes through a CTEV position and achieves, by the end of the 11th week, a plantigrade position, which is the normal adult foot position. As early as 1929, Böhm proposed that CTEV arises as an ‘arrest’ of normal embryonic development of the foot but there is as yet no evidence for a single specific pathogenic mechanism.

In order to increase our knowledge of the origins of CTEV, we sought an animal model. Peroneal muscle atrophy (pma) is the only mouse mutant, of which we are aware, that has ‘clubfoot’ in hind limbs and not fore limbs, and no other developmental abnormalities in either homoyogotes or heterozygotes and it therefore seems to be the best available mouse model. Homozygous pma mice have bilateral ‘clubfoot’, whereas heterozygotes may have a unilateral defect. The characteristic ‘clubfoot’ phenotype in pma hind limbs is associated with the degeneration of lateral (peroneal) and anterior compartment muscle groups (Esaki et al., 1981; Nonaka et al., 1986; Katoh et al., 2003). These myopathic changes are reported to be due to the absence of the peroneal division of the sciatic nerve, which renders the lateral and anterior compartment muscles of the lower limb aneural throughout development (Ashby et al., 1993). Sangha et al. (2003) described another ‘clubfoot’ mouse mutant, the radiation-induced model gammy, in which the Rho-GTPase encoding gene GRIT is deleted. Gammy heterozygotes have unilateral hind limb ‘clubfoot’ but the mutation is lethal in homozygotes and associated with severe brain abnormalities.

Here we have compared the 3D musculoskeletal anatomy of the hind limb in pma and wild-type mice using micro-magnetic resonance imaging (µMRI) in order to assess whether pma mice could serve as a possible model for human CTEV. High-resolution µMRI is increasingly being used to investigate mouse anatomy giving a remarkable opportunity to examine both bone and soft tissue structures and their relationships to each other (Smith et al., 1994; Jacobs et al., 1999; Dhenain et al., 2001; Nieman et al., 2005). We also studied embryonic foot rotation in the hind limbs of normal mice and pma mutants to gain insights into the developmental basis of the ‘clubfoot’ phenotype in these mice.

**Materials and methods**

**Animal husbandry**

Wild-type mouse strains (CD-1 and CS7/Black) and the mutant strain pma were used. The pma mice were bred as homoyogotes. Heterozygous pma mice, when required, were the offspring of CD-1-pma matings. All mice were housed in Thoren IV C cages maintained at positive pressure with a light cycle of 14 h light, including a dusk period, and 10 h dark. All were fed autoclavable RM3 diet (Special Diets Services, Witham, Essex, UK) with ad-libitum autoclaved water. The work was carried out in accordance with the Animal Scientific Procedures Act (1986) and after local ethical review by the Dundee University Ethical Review Committee.

**Preparation of specimens for hind limb anatomy**

Hind limbs were removed from 3-week-old wild-type (CS7/Black) and mutant pma animals at the level of the proximal femur (at the hip joint) and skin was removed down to the ankle. Dissected limbs were then fixed by immersion in 4% paraformaldehyde in 0.1 M phosphate-buffered saline and left overnight at 4 °C. On day 2, specimens were washed three times with phosphate-buffered saline for 15 min per wash.

For µMRI, most specimens were mounted in 25-mm glass tubes containing Fomblin LC808 (Solvay Solexis, Milan, Italy). Specimens were dried of any excess fluid and held in place with plastic tubing. Some specimens were embedded in 1% agar containing 60 μL mL⁻¹ (0.5 mmol mL⁻¹) of the gadolinium (III) contrast agent Gd-diethylenetriamine-penta-acetate-bis(methyylamide) (Omniscan®).

**MicroMRI instrumentation**

MicroMRI data were acquired on a Bruker AVANCE FT NMR spectrometer (Bruker Biospin GmbH, Rheinstetten, Germany) with a wide-bore 7.1-Tesla magnet resonating at 300.15 MHz for ¹H. The spectrometer was fitted with Bruker micro-imaging magnetic field gradients. A birdcage radio-frequency resonator with an internal diameter of 25 mm was used. Spin-echo and gradient-echo (FLASH) magnetic resonance imaging (MRI) experiments, from the Bruker Paravision® library, were performed. The radio-frequency resonator was tuned and the magnet shimmed for each sample. Typically, four to 12 acquisition sequences were collected and averaged to improve the signal-to-noise ratio and reduce artefacts. 3D 128 × 128 × 128 and 256 × 256 × 256 data sets were acquired. Short and long echo time (T₁, spin-echo and gradient-echo imaging experiments were carried out to observe the entire hind limb of the 3-week-old wild-type (CS7/Black) and pma mutant mice. Both left and right limbs were studied. All acquisitions were made at 19 °C. The MRI pulse sequences used, size of data set, recycle time (T₁)
and $T_2$ are reported in the figure legends along with fields of view and spatial resolution.

To acquire high-resolution μMRI of murine feet, in which the bones and much of the surrounding soft tissues have short 1H $T_2$ relaxation time, a FLASH gradient-echo imaging pulse sequence with $T_2\text{r}$ of 2.3 ms was used and 256 × 256 × 256 data sets were acquired. A spatial resolution of 98 μm pixel$^{-1}$ was achievable even with a field of view of 2.5 cm.

**MicroMRS data analysis**

MicroMRS data were Fourier transformed and then visualized using *Amira* imaging software (Mercury Computer Systems Inc., Chelmsford, MA, USA), which allowed 2D slices to be viewed from any angle within the 3D data set and regions of interest to be segmented and data reconstructed to produce 3D surfaces. The 3D finite element meshes were generated from segmented image data, surface rendered and volumetric measurements determined. The software was used to register and overlay the 3D surface-rendered data acquired using different μMRI parameters. This allowed the musculoskeletal anatomy to be digitally reconstructed and also enabled features of the skeleton of normal and mutant mouse hind limbs to be compared.

**Embryology of late fetal mouse hind limbs**

According to the atlas of mouse development (Kaufman, 1992), hind limbs in mouse embryos begin to rotate at embryonic day (E)15.5. Therefore, we collected embryos at daily intervals from E14.5 to E18.5. CD-1, C57/Black and pma mouse embryos were obtained from females mated in the evening. By convention, the morning on which the vaginal plug was found is referred to as 0.5 days post-conception (Kaufman, 1992). Embryos were fixed in 4% paraformaldehyde as described above and washed in phosphate-buffered saline. Selected specimens were dehydrated with methanol and stored at −20 °C. Embryos were placed on their backs, observed under a stereomicroscope for recording orientation of the hind limbs/feet and then photographed.

**Results**

**MicroMRS anatomical study of mouse hind limbs**

In order to visualize the internal anatomy of pma hind limbs, we carried out various gradient-echo and spin-echo 3D μMRI experiments and compared mutant limbs with wild-type limbs from 3-week-old pma and C57/Black mice. Figure 1A, an image from a 3D μMRI data set of a wild-type hind limb, demonstrates the quality of the anatomical information obtained. The images of the bones appear heterogeneous because the signal arises from a combination of tissues including bone, bone marrow, growth plates, blood vessels and cartilage. Mineralized bone has a very short transverse relaxation time ($T_2$) and so appears black in the image, whereas the cartilaginous tissue of the epiphyseal growth plate gives rise to a high signal intensity and appears white. The major tendons in the hind limb including the calcaneal tendon and flexor digitorum longus tendon can be observed and appear black.

The *Amira* software package was used to surface render the whole leg and different anatomical structures in the magnetic resonance images, and 3D surface reconstructions were produced. To facilitate comparisons, all images presented here are of left hind limbs. Surface reconstructions of the whole leg highlight a number of significant anatomical differences between wild-type and pma legs. The most striking differences occur in the foot. The pma foot is supinated with the toes adducted and curled medially (compare Fig. 1B,C with Fig. D,E). In addition, the anterior lateral side of the leg is much thinner in pma (Fig. 1E, see dashed line) than in wild-type (Fig. 1, see also Table 1) mice. Surface reconstructions of the skeletal elements showed that the fibula and tibia of the pma and wild-type hind limbs (Fig. 1F–I) are very similar in size, shape and orientation. The distal ends of the tibia and fibula in the pma leg do not appear to be distorted and thus are unlikely to be implicated in the pathogenesis of the ‘clubfoot’. The size and shape of the calcaneus (green) and talus (yellow) bones are also very similar in pma and wild-type feet (Figs 1 and 4). This suggests that bone growth is not affected in pma. However, there are major differences in the position and orientation of these two bones in the feet of pma mice with the calcaneus lying behind the talus (Fig. 1I) rather than lying alongside it as in the normal foot (Fig. 1G). Associated with this plantar flexion of the talus and calcaneus, there is also plantar flexion and adduction of the metatarsals and phalanges in the mutant foot (Fig. 1H,I).

The positional abnormalities in the pma foot are very obvious when 3D surface-rendered images of pma hind limb bones (blue) are overlaid onto wild-type leg bones (white) aligned relative to the tibial mechanical axis in Fig. 2. The views in the sagittal plane (Fig. 2A,C) show that the pma foot exhibits plantar flexion, whereas the frontal views (Fig. 2B,D) emphasize the inversion and adduction of the mid and fore foot in pma.

Most of the anatomical changes seen in pma mutant hind limbs occur in the foot and we therefore carried out a high-resolution μMRI study of the mouse hind foot (Fig. 3). The bony structures such as the tibia, calcaneus, talus, distal tarsals, metatarsals, phalanges and claws are clearly resolved (Fig. 3A,B). The flexor digitorum longus tendon, which, like other tendons, has a short 1H $T_2$ relaxation time, appears black and can be seen running along the base of the foot. In the individual metatarsals seen in the frontal sections (Fig. 3C,D), the mineralized bone appears black and the four blood vessels in the centre of the bone and the bone marrow appear grey. The extensor digitorum longus, flexor hallucis longus and flexor digitorum longus tendons are all visible.

Surface reconstructions were produced of the distal ends of the tibia and fibula, and the calcaneus, talus, centrale, distal tarsal, metatarsal bones and tendons in wild-type and pma left hind feet (Fig. 4A–F). The pma foot has marked plantar flexion (compare Fig. 4B with Fig. 4A). The
Flexor digitorum longus tendons (brown) were seen in a similar position in both wild-type and pma feet but pma tendons appeared to be shorter. The wild-type and pma feet shown in Fig. 4C–J are aligned relative to the talus bone and the tibial mechanical axis. This alignment was chosen in order to focus on changes in the orientation of the bones of the foot distal to the ankle in pma. Figure 4C–J shows the proximal bones, calcaneus (green), talus

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(yellow), centrale (red), first (pink), second (purple), third (light blue) and fourth/fifth (dark blue) distal tarsals and all other bones (white), with pma bones being assigned a darker shade than the wild-type. The interface between the centrale bone and third distal tarsal was unclear, making accurate digital segmentation of this interface a slight problem.

The µMRI surface reconstruction of the bones in the wild-type mouse foot (Fig. 4C) compares very well with Margaret Cook’s diagram of the bones in a mouse foot (http://www.informatics.jax.org/cookbook/figures/figure44.shtml).

In addition, the comparison in Fig. 4C–F demonstrates the inversion that twists the pma foot inwards resulting in the fourth/fifth metatarsal lying below the third (compare Fig. 4D with Fig. 4C) and also the plantar flexion of the pma foot downwards (compare Fig. 4F with Fig. 4E). Note that, in this detailed examination, no differences in the distal ends of the tibia and fibula were detected.

The positional abnormalities of the bones in the pma foot became more obvious when the surface-rendered images were overlaid onto the wild-type data set (Fig. 4G–J). The red arrows in Fig. 4G show that the tibiale and first distal tarsal pma bones are superior to the equivalent wild-type bones and this change in orientation of the first distal tarsal bone is shown in a different view in Fig. 4H. Figure 4H also shows that the third and fourth/fifth distal tarsals of the pma foot are inferior to the equivalent distal tarsals of the wild-type foot, whereas the second distal tarsal of the pma foot is at the same height as the second distal tarsal of the wild-type foot but is displaced laterally. These changes are consistent with inversion of the pma foot. The same gross displacements were not seen when comparing the pma talus, calcaneus and centrale bones with their equivalent wild-type bones (Fig. 4I,J). This is due to the fact that the feet are aligned relative to the talus bone and the tibial mechanical axis. However, it is worth noting that the surface-rendered images show that the relative positions of the calcaneus, talus and centrale pma bones are the same as those of the wild-type bones as shown by red lines in Fig. 4I; in addition the bones are of similar size and shape in the mutant and wild-type foot.

The surface reconstructions of the whole leg in Fig. 1B–E highlighted a difference in the shape of the pma shank compared with the wild-type. A µMRI study was therefore undertaken to determine the volumes of the various muscle groups in the shank region in hind limbs of wild-type and pma mice. MRI experiments can be designed to differentiate between the various muscle groups. The striations in the muscles generate local magnetic field inhomogeneities (Heemskerk et al., 2005) and these perturbations affect the image contrast especially in long $T_2$ images. A slice from a 3D spin-echo data set of a wild-type mouse leg is shown in Fig. 5A; the image is taken across the shank of the lower hind limb, which had been embedded in agarose gel containing gadolinium contrast agent. Several muscle groups can be identified. The shank muscles were digitally segmented into three muscle groups to determine whether there are any significant changes in shank muscle volume between the wild-type and pma mice. The anterior lateral muscles were assigned as muscle Group A comprising the tibialis anterior, extensor hallucis longus, extensor digitorum longus and fibularis (peroneus) tertius; the anterior medial muscles were assigned as muscle Group B comprising the flexor digitorum longus; and the posterior shank muscles were assigned as muscle Group C comprising the

<table>
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<th>Anatomical feature</th>
<th>pma mouse hind limb anatomy</th>
<th>Human CTEV leg anatomy</th>
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<tr>
<td>Tibia and fibula</td>
<td>pma bones same size and shape as wild-type. Distal end of the pma bones not distorted</td>
<td>Not affected</td>
</tr>
<tr>
<td>Shank muscle</td>
<td>Reduced</td>
<td>Overall shank muscle reduced</td>
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<tr>
<td>Anterior lateral shank muscles</td>
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<td>Relatively unaffected</td>
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<td>Anterior medial shank muscles</td>
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<td>Relatively unaffected</td>
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<td>Orientation of the foot in sagittal plane</td>
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<td>Equinus deviation</td>
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<td>pma foot shows inversion and adduction of the mid foot and fore foot</td>
<td>Inversion and adduction</td>
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<tr>
<td>Flexor digitorum longus tendons</td>
<td>Appear shorter in pma</td>
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</tr>
<tr>
<td>Calcaneus, talus, centrale</td>
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<td>Variable evidence for changed shape/size</td>
</tr>
<tr>
<td>Tarsals</td>
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<tr>
<td>Metatarsal</td>
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</tr>
<tr>
<td>Phalanges</td>
<td>Curled and inverted in pma</td>
<td>May demonstrate latent curling during correction due to shortened flexors</td>
</tr>
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lateral and medial gastrocnemius, soleus, tibialis posterior and flexor hallucis longus.

Surface reconstructions of the muscle groups in the shank of a wild-type and pma hind limb are shown in Fig. 5B,C, respectively. Mean muscle volumes were measured for three wild-type and three pma legs (Fig. 5D). There were no marked differences in the total muscle sizes of Group B (anterior medial shank muscles) and Group C (posterior...
shank muscles) between wild-type and pma. However, the anterior lateral shank muscles (Group C) in the pma hind limb were on average one-third of the size of those of the wild-type hind limbs. A reduction in mass of these muscles was expected as pma causes atrophy of the peroneal muscles; however, it might be, given the marked difference in volume, that more than just the fibularis (peroneal) muscles have atrophied.

**Positional changes of the hind limb during late development of wild-type and pma mouse embryos**

To gain insights into how the anatomical defects in pma mice described above might arise, we studied hind foot rotation during embryonic development. We devised a four-stage classification system based on the position of the feet of wild-type embryos seen when the embryo was placed on its back. The pre-rotation position was termed ‘inclined’ (i) with the foot inclined slightly outwards with the lateral border lying closer to the midline than the medial border (Fig. 6A); the next stage, ‘straight’ (s), was reached when the footplate rotated to the sagittal plane and the ventral (plantar) surfaces of the footplates lie parallel to each other (Fig. 6B); and the third stage, ‘tilted’ (t), described an intermediate rotational position in which the foot had commenced rotation towards the midline and the medial border of the footplate was nearer the midline than the lateral border so that the ventral (plantar) surface of the foot was just becoming visible (Fig. 6C,D, left foot); and the final fourth stage was when the foot had rotated completely to the plantigrade position, termed ‘rotated’ (r) (Fig. 6C,D, right foot). In this final position, the ‘rotated’ hind feet resemble the neonatal and adult position. Left and right feet were individually scored; this was indicated by the prefix ‘L’ and ‘R’, respectively.

The hind foot position was scored in embryos of litters of the wild-type strains CD-1 and C57/Black (the latter strain had been used as controls for examining foot anatomy) and pma mutants. Figure 7 shows histograms of the data collected on the position of both left and right feet of embryos between E14.5 and E18.5 of the three strains of mice. In CD-1 embryos, turning of the hind feet, from inclined (i) to straight (s), began around E15.5 and rotation was completed in some embryos by E16.5 (i.e. both feet fully rotated, RrLr) although in other embryos some feet had not fully rotated at E18.5. In another wild-type strain, C57/Black, foot rotation also started around E14.5 but in none of the embryos had the feet completely turned (RrLr) by E17.5, although in some embryos this had occurred by E18.5. Therefore, foot rotation was slightly slower in C57/Black compared with CD-1 mice. Final rotation of the feet in some mice is completed postnatally.

Foot rotation in pma mutant mice is very different. The initial phase of foot rotation from inclined (i) to straight (s) can be delayed by several days. Although in some embryos, turning from inclined to straight did occur at E14.5, in several embryos, rotation had not even started by E16.5. Furthermore, the ‘straight’ position in pma embryos appears to persist longer than in CD-1 and C57/Black. The most striking feature is, however, that hind foot rotation was not completed in pma embryos (no embryos were scored RrLr). Thus, at E18.5 (Fig. 7E), 77% of pma (10/13) had either one or two legs tilted and foot rotation had not advanced...
beyond this stage, whereas 91% of CD-1 (30/33) and 88% of C57/Black (7/8) mouse embryos had one or both feet completely rotated. Examination of these data also revealed, unexpectedly, that the timing of foot rotation was asymmetrical in many wild-type embryos. In wild-type embryos in which one leg was more advanced than the other, going from inclined to straight, from straight to tilted or from tilted to rotated, the right leg was more advanced than the left in 73% (37/51) of CD-1 and in 78% (14/18) of C57/Black mouse embryos. In contrast in pma mice, in only 38% (3/8) of the cases in which one leg was more advanced than the other was the right leg involved.

As part of the breeding programme associated with this work, pma mice were mated with CD-1 mice; the resulting heterozygous offspring were brother–sister mated. From eight original heterozygote matings, 75% of the offspring had unilateral ‘clubfoot’ affecting the right
foot; in 6 litters from the brother–sister matings between 100 and 66% of the affected animals had right ‘clubfoot’, whereas one litter contained five animals in which four mice had left ‘clubfoot’. This shows that unilateral clubfoot in pma heterozygotes predominantly affects the right foot.


Discussion

We have used μMRI to analyse and compare the detailed 3D anatomy of the hind limbs in pma and wild-type mice. The gross anatomical appearance of the pma mouse foot shows prima facie similarities with human CTEV. This study has demonstrated (Table 1) that many of the underlying skeletal changes seen in pma mouse legs are similar to those of human clubfoot (Ponseti, 1996; Pirani et al., 2001). Careful comparisons showed that the size, shape and orientation of the tibia and fibula are not affected in pma mice. This corresponds to the usual pattern seen in human CTEV, in which tibial torsion is uncommon, although it is occasionally seen. Although the fact that the mouse is quadripedal must be borne in mind, the gross positional anatomical abnormalities of the foot observed in pma mice resemble those typically seen in human CTEV. The plantar flexion of the ankle in the sagittal plane and the inversion and adduction of the mid foot and fore foot in the frontal plane seen in the pma mouse (Fig. 2) are equivalent to the equinus and varus deviations, respectively, which are used to classify the severity of human clubfoot (Windisch et al., 2007a). In addition, more distal elements, the tibiale and tarsals, in the feet of pma mice are clearly displaced from their normal positions (Fig. 4). More detailed analysis will be needed to examine the detailed relationships between the joints in the ankle in the normal and mutant mice and to compare these with normal and CTEV joints. This would confirm how closely pma mirrors the human condition.

In contrast to these changes in the orientation of the distal bones, their size and shape are similar in pma and wild-type mice. The potential for comparative anatomical study in this respect is hampered by the diversity of reported bony anatomy in human CTEV. Thus, MRI studies of human patients with surgically treated CTEV demonstrate smaller articulations and a different shape in the treated ‘clubfoot’ compared with the contralateral normal foot (Roche et al., 2006). Conversely, the appearances of the tarsal bones, following Ponseti (non-surgical) treatment, more closely resemble those of unaffected feet. Ongoing MRI studies of the human CTEV model should clarify this point (Pirani S, personal communication). Prominent curling of the phalanges in pma may be due to phalanges and claws (nails) in the quadripedal mouse foot being proportionately longer than in the bipedal human foot.
Our observations on the distal tendons of the foot in pma mutant mice suggested that they were unaffected, apart from a slight shortening. This is similar to human clubfoot, in which soft tissues on the plantar, medial and posterior aspects of the foot, ankle and distal leg are present but consistently shortened (Ippolito & Ponseti, 1980; Windisch et al., 2007). Our analysis of shank muscle volumes showed that the volume of the antero-lateral muscle group, which includes the extensor hallucis longus, extensor digitorum longus and fibularis (peroneus) tertius muscles, was reduced by around 70% in pma. These MRI data support dissection data describing the significantly reduced volume of the anterior lateral shank compartment muscles (peroneal muscles) in pma legs (Ashby et al., 1993). In human CTEV, shank muscle is generally reduced on the affected side (Gutstein, 2006; Chesney et al., 2007) and a recent study showed that it is the posterior compartment muscles that are most affected in human patients rather than the anterior lateral compartment muscles as in pma (Ippolito et al., 2009). This study also detected muscle atrophy associated with clubfoot in 13- and 19-week-old human fetuses, suggesting that muscle changes are a primary feature of CTEV.

Our l MRI analysis of the arrangement of the tarsal and metatarsal bones in pma hind feet is consistent with the defect in these mice being the result of incomplete embryonic rotation of the foot. Indeed, our observations on foot orientation in pma mouse embryos at a series of stages in development show quite clearly that rotation is incomplete. In wild-type mice, although the detailed timing appears to differ in the two different strains that we investigated, rotation of the foot takes place towards the end of gestation over several days (from E14.5), with final rotation only being completed in some individuals post-natally. In contrast, in pma mice, rotation is delayed and never completed. These observations show that we can exclude the possibility that the foot rotates normally in pma mice and that later the foot is pulled into an abnormal position. If the usefulness of pma as a model for clubfoot is confirmed, our observations would provide support for the long-standing hypothesis of Böhm (1929) that CTEV is due to the ‘arrest’ of the normal process of rotation and angulation of the foot.

A striking feature to emerge from our detailed analysis of foot rotation in wild-type mouse embryos is the difference between right and left. In wild-type embryos in which one foot is more advanced than the other, this is more commonly the right. The basis for this difference in the timing of rotation between right and left is not clear. One of the signals that is involved in setting up left/right asymmetry of the body in early vertebrate embryos is retinoic acid (Levin et al., 2006) and it is intriguing that, in rats, a single intragastric dose of retinoic acid (120 mg kg⁻¹ body weight) on day 10 of pregnancy (Delgado-Baeza et al., 1999) invokes a ‘clubfoot’ phenotype. Analysis of heterozygous pma mice with unilateral ‘clubfoot’ also showed that the right foot

![Histograms showing position of hind feet in CD-1, C57/Black and pma embryos (A) E14.5; (B) E15.5; (C) E16.5; (D) E17.5; (E) E18.5. R, right foot; L, left foot; i, inclined; s, straight; t, tilted; r, rotated.](image-url)
was predominantly affected. Strikingly, in human patients, the ratio of right-sided to left-sided CTEV again shows a right bias.

The gross similarities between the anatomy of the pma hind limb and human CTEV suggest that the study of pma mice could open up new avenues of investigation into the potential origins and causes of CTEV in humans. The gene for pma has been mapped by genetic linkage to a large region on mouse chromosome 5 (Katoh et al., 2003) and further mapping of the genetic basis of pma may identify a gene and/or developmental mechanism causative of CTEV in humans.

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References

Supporting Information
Additional Supporting Information may be found in the online version of this article.

Fig. S1. Movie of 3D surface reconstructions of mineralized bones from µMRI of left legs of 3-week-old mice; wild-type leg is shown in white and pma leg is shown in blue. Images are aligned relative to the tibial mechanical axis to allow direct comparison of anatomy. Mineralized bones of wild-type and pma legs were reconstructed from T_E/TE = 500/6 ms and T_E/TE = 500/40 ms spin-echo image data sets and a
\( T_R/T_E = 250/2.3 \text{ ms gradient-echo image data set (matrix size, } \) 
\( 56 \times 256 \times 256; \text{ field of view, } 25 \times 25 \times 25 \text{ mm; voxel dimensions, } \)
\( 97 \times 97 \times 97 \text{ μm).} \)

**Fig. S2.** Movie of 3D surface reconstruction of μMRI data sets of left feet of 3-week-old wild-type and pma mice; both pma and wild-type data sets are opaque, the pma data set has a darker hue. Reconstructed from \( T_R/T_E = 250/2.3 \) ms gradient-echo image data sets (matrix size, 128 × 128 × 128; field of view, 10 × 10 × 10 mm; voxel dimensions, 78 × 78 × 78 μm).

\( T_R/T_E = 500/2.3 \text{ ms and } T_R/T_E = 1000/20 \text{ ms gradient-echo image data sets.} \)

**Fig. S3.** Movie of a 3D surface reconstruction of μMRI of the shank muscles of a leg of a 3-week-old wild-type mouse. Generated from \( T_R/T_E = 500/2.3 \) ms and \( T_R/T_E = 1000/20 \) ms gradient-echo image data sets.

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