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Identification and characterisation of Helicobacter pylori O-acetylserine-dependent cystathionine β-synthase, a distinct member of the PLP-II family

Running title: Structural insights into H. pylori OCBS

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Summary

O-acetylserine sulphydrylase (OASS) and cystathionine β-synthase (CBS) are members of the PLP-II family, and involved in L-cysteine production. OASS produces L-cysteine via a de novo pathway while CBS participates in the reverse transsulfuration pathway. O-acetylserine-dependent CBS (OCBS) was previously identified as a new member of the PLP-II family, which are predominantly seen in bacteria. The bacterium *Helicobacter pylori* possesses only one OASS (hp0107) gene and we showed that the protein coded by this gene actually functions as an OCBS and utilizes L-homocysteine and O-acetylserine (OAS) to produce cystathionine. HpOCBS did not show CBS activity with the substrate L-serine and required OAS exclusively. The HpOCBS structure in complex with methionine showed a closed-cleft state, explaining the initial mode of substrate binding. Sequence and structural analyses showed differences between the active sites of OCBS and CBS, and explain their different substrate preferences. We identified three hydrophobic residues near the active site of OCBS, corresponding to one serine and two tyrosine residues in CBSs. Mutational studies were performed on HpOCBS and *Saccharomyces cerevisiae* CBSs. A ScCBS double mutant (Y158F/Y226V) did not display activity with L-serine, indicating indispensability of these polar residues for selecting substrate L-serine, however did show activity with OAS.

**Keywords:** Cystathionine β-synthase, *Helicobacter pylori*, Hydrogen sulfide, O-acetylserine sulphydrylase, pyridoxal 5′-phosphate, reverse transsulfuration pathway

Introduction

L-cysteine can be produced via the transsulfuration pathway or a de novo pathway depending on the presence of enzymes in the organism. In the transsulfuration pathway, L-methionine is converted into L-homocysteine in multiple steps, with this conversion requiring the action of different enzymes. The L-homocysteine is either converted back to L-methionine via the action of the enzyme L-homocysteine S-methyltransferase or is converted to L-cysteine through an intermediate L-cystathionine via the reverse transsulfuration pathway (Scriver & Kaufman, 2001, Stipanuk, 2004) (Fig. 1). In the reverse transsulfuration pathway, homocysteine reacts with L-serine and is converted to cystathionine by the action of cystathionine β-synthase (CBS), and cystathionine is further acted upon by cystathionine γ-lyase (CGL) to produce L-cysteine. The de novo pathway of L-cysteine production from L-serine with O-acetylserine as an intermediate has been shown in bacteria and plants, where serine O-acetyl transferase (SAT) and O-acetylserine sulphydrylase (OASS) function in tandem (Rabe & Cook, 2004, Wirtz & Droux, 2005, Kopriva, 2006) (Fig. 1). Recently a few O-acetylserine-dependent CBSs (OCBs) have been identified and characterised in certain bacteria, have been shown to function in the reverse transsulfuration pathway, specifically being involved in methionine-to-cysteine conversion that takes O-acetylserine from the de novo pathway (Devi et al., 2017, Hullo et al., 2007, Matoba et al., 2017).
CBS belongs to the type II fold of PLP-dependent enzymes, and OASS also adopts a similar three-dimensional fold (Alexander et al., 1994, Grishin et al., 1995, Meier et al., 2001, Burkhard et al., 1998). Extensive research has been carried out on Homo sapiens CBS (HsCBS), Drosophila melanogaster CBS (DmCBS) (Koutmos et al., 2010) and Saccharomyces cerevisiae CBS (ScCBS) (Tu et al., 2018). HsCBS and DmCBS each have an extended N-terminal heme-binding region, which probably serves as a redox sensor (Taoka et al., 2002). But neither ScCBS (Tu et al., 2018) nor protozoan CBSs have N-terminal heme-binding region (Nozaki et al., 2001). HsCBS, DmCBS and ScCBS each have a regulatory C-terminal Bateman domain, which is absent in protozoan CBS (Nozaki et al., 2001), but only HsCBS is activated by the binding of the allosteric regulator S-adenosylmethionine (AdoMet) (Finkelstein et al., 1975). All CBSs catalyze the generation of cystathionine via a β-replacement reaction of L-serine with L-homocysteine, along with other alternative β-replacement and β-elimination reactions in which L-cysteine and L-homocysteine are utilized to release H₂S. CBS, especially in the brain, serves as a major source of H₂S, which is involved in modulating the cell function (Chen et al., 2004). Bacterial OCBS also belongs to the PLP type II fold and catalyzes the synthesis of cystathionine via the β-replacement reaction of O-acetylserine and L-homocysteine. OCBS has neither an N-terminal heme-binding region nor a C-terminal regulatory domain (Devi et al., 2017, Matoba et al., 2017). These OCBSs are also capable of catalyzing the production of H₂S efficiently via β-replacement with substrate L-cysteine and L-homocysteine (Devi et al., 2017, Matoba et al., 2017).

Helicobacter pylori is a pathogenic Gram-negative bacterium known to colonize the stomachs of about half of the global human population, largely in developing and resource-poor countries. It affects the majority of people with subclinical to progressive clinical gastritis and peptic ulcers (Nesic et al., 2014, Singh et al., 2002). Thus, considering the global etiological magnitude and prominence of the invasiveness of H. pylori, this pathogen has been listed as a group 1 carcinogen by the World Health Organisation (WHO) and is considered as the strongest risk factor associated with duodenal and gastric cancers (Peek & Blaser, 2002).

In the H. pylori genome, both the SAT and OASS genes have been annotated to belong to the de novo pathway, but an overview of the genome of H. pylori showed a gene cluster containing the annotated OASS gene (hp0107) along with the genes of the cystathionine γ-synthase (hp0106) and S-ribosylhomocysteinase (hp0105) enzymes of the transsulfuration pathway. This analysis suggested the product of the hp0107 gene to be also involved in the transsulfuration pathway, where the products of all of these genes are required and thus expressed simultaneously as reported earlier (Doherty et al., 2010). Some recent findings also suggested (Devi et al., 2017, Matoba et al., 2017) that the product of hp0107 gene could actually be an OAS-dependent CBS (HpOCBS) in H. pylori. It should be noted that the transsulfuration pathway has been indicated to be crucial for the viability of H. pylori since it grows only in L-methionine-rich media (Mendz & Hazell, 1995, Nedenskov, 1994, Reynolds...
and mutations in any one of these genes have been shown to impede the growth of the organism and to result in it becoming a cysteine auxotroph (Doherty et al., 2010).

In the current work, we first carried out an in silico study to identify the sub-family to which HP0107 belongs. A phylogenetic tree analysis showed the presence of an OCBS clad separate from OASS and CBS, with the OCBS branch including all annotated OCBSs along with HP0107. This observation indicated HP0107 to be an OCBS (HpOCBS). The HpOCBS protein showed OAS-dependent CBS activity and also catalysed the production of H₂S in the presence of L-cysteine and L-homocysteine. Furthermore, we determined the crystal structure of HpOCBS with bound L-methionine (substrate analog); this analog was observed to be bound near PLP at the active site in one of the protomers of the dimeric structure, explaining the pre-reactive substrate binding state. A comparison of the active sites of OCBS and CBS revealed three key residues playing an important role in substrate selectivity for β-replacement activity to produce cystathionine. Mutation of two of these residues in Saccharomyces cerevisiae CBS (ScCBS) resulted in a loss of activity with substrate L-serine and a gain of activity with OAS. Earlier reports on OCBS (Devi et al., 2017, Hullo et al., 2007, Matoba et al., 2017) suggested OCBS to be present only in a few bacteria, but our study revealed the presence of OCBS in a greater variety of bacteria.

Results

In silico study indicates HP0107 is an OCBS

We retrieved many protein sequences annotated either as cysteine synthase or OASS from the NCBI database using PSI-BLAST with Bacillus anthracis OCBS as the search template (due to it having previously been shown to be an OCBS (Devi et al., 2017)). Most of these retrieved sequences probably corresponded to OCBS, with the product of the gene hp0107 of H. pylori being one of them. These probable OCBSs derived from the phyla Firmicutes, Chlamydiae, Acidobacteria, Fibrobacter, Actinobacteria and Proteobacteria of Bacteria and Eurychoaeota of Archaea (Table S1). A separate list of CBS and OASS proteins from various organisms were also created using PSI-BLAST, with the OASS family containing sequences of OASSs from both bacteria and plants, and the CBS family including sequences from the phyla Arthropoda, Chordata and Nematoda.

Furthermore, a phylogenetic tree was constructed using sequences considered likely to be OCBS, CBS and OASS. A total of 55 probable OCBS sequences (most of them annotated as cysteine synthase in NCBI) clubbed neither with OASS nor with CBS and were instead included in a separate OCBS branch. This separate OCBS branch also contained the product of the gene hp0107 along with other annotated OCBS sequences as well. This analysis suggested OCBS to differ from OASS and CBS in the PLP-II family of enzymes (Fig. 2) and indicated that the product of the gene hp0107 belongs to the OCBS subfamily.
Sequence conservation near the active site in OCBS, CBS and OASS

Three multiple sequence alignments were carried out using OCBS, OASS and CBS sequences, respectively, to determine the level of conservation within each of these three types of proteins. The Multalin server was used, which gave a consensus sequence and printed a residue at any particular position when the level of conservation was at least 50% (Fig. S1). The consensus sequence near the active site of the enzyme was divided into five conserved blocks, which showed differences among the sub-families. Inspection of the sequence alignment of conserved block 1, consisting of residues TAGNTG in OCBS sequences (residue numbers according to HpOCBS), indicated TSGNTG to be the corresponding conserved residues in CBS and OASS (Table S2, see also Fig. 3). This sequence in OASS and CBS has been shown to interact using multiple contacts with, respectively, substrate OAS (Schnell et al., 2007) and L-serine (Koutmos et al., 2010) and to also stabilize the α-aminoacrylate intermediate. Inspection of crystal structures indicated this conserved block, also known as the ‘asparagine loop’, to move upon substrate binding and to close the active site cleft. Conserved block 2, consisting of GSGGT in OCBS, has been indicated to interact with the phosphate moiety of PLP and to anchor the moiety throughout the β-replacement reaction in OCBS (Matoba et al., 2017), OASS (Schnell et al., 2007) and CBS (Koutmos et al., 2010). However, GTGGT was found to be the corresponding sequence conserved in both CBS and OASS. Conserved block 3, has been predicted to interact with the second substrate (L-homocysteine or HS’ in CBS (Lodha et al., 2009) or OASS, respectively) and to allow this second substrate to participate in a nucleophilic attack on α-aminoacrylate and then release product and regenerate enzyme with an internal aldime. In the third block, the Met was found to be mostly conserved, but with a Val instead present in HpOCBS. Moreover, these conserved residues were observed to be located near the lip of the active site. Previous analyses of CBS and OCBS crystal structures showed Glu222 of this conserved stretch to form a network of hydrogen bond interactions and to close the active site cleft upon substrate binding (Devi et al., 2017). Sequence alignment revealed IQGIGAG and VEGIGYD to be the block 3 sequences conserved in OASS and CBS, respectively. Tyr in conserved block 3 of CBS has been reported to interact with the substrate L-serine and to play an important role during the β-replacement reaction (Koutmos et al., 2010, Tu et al., 2018). Ser267 of conserved block 4 of OCBS was observed to interact with the nitrogen of the pyridine ring of PLP; thus, this conserved block may be stabilizing the PLP ring and also holding it during the interconversion between the internal and external aldimines (Schnell et al., 2007, Koutmos et al., 2010). Ser267 and Ser268 were found to be conserved in CBS and OASS as well. Inspection of crystal structures showed conserved block 5 to be located at the C-terminal tail and Lys302 of this block to interact with other residues at the lip of active site in OCBS and CBS. And inspection of the sequence alignment indicated RYLSK and NYMTK to be the sequences of this block conserved in OASS and CBS, respectively, with the lysine at the fifth position of this block notably conserved in OCBS and CBS. These analyses taken together revealed differences at the sequence level in the conserved blocks in different members of the PLP-II family (Table S2 and Fig. 3).
Kinetics analysis revealed that HP0107 is an OCBS

To confirm that HP0107 is an OCBS, we carried out various biochemical assays. Enzyme was purified to homogeneity to carry out all the enzymatic assays. First, we performed a CBS assay in the presence of substrate L-serine and homocysteine, but it did not show any production of cystathionine. Then OCBS activity was measured to determine whether HP0107 produces L-cystathionine via the β-replacement reaction with OAS and L-homocysteine. HP0107 showed OCBS activity with a $k_{\text{cat}}/K_m$ of 12.6 mM$^{-1}$·s$^{-1}$ for OAS (Table 1 and Fig. 4a), similar to that previously reported from B. anthracis (Devi et al., 2017) and L. plantarum (Matoba et al., 2017). The HP0107 $K_m$ value for OAS was 2.92 mM, comparable to 1.16 mM and 4.2 mM of OCBS activity of B. anthracis (Devi et al., 2017) and L. plantarum (Matoba et al., 2017) respectively. Various reported $K_m$ values for L-serine of Homo sapiens, Leishmania major and Trypanosoma cruzi CBS were 1.74 mM, 1.1 mM and 1.0 mM respectively (Chen et al., 2004, Nozaki et al., 2001, Williams et al., 2009), similar to $K_m$ of OCBS activity of HP0107. Initially, the OCBS assay was performed using various buffers at different pH values and the highest activity was observed when using HEPES [pH 7.5] buffer. Thus, this buffer was used to carry out all of the reactions. Absence of CBS activity with the substrate L-serine was also observed in the case of other OCBSs, i.e., from B. anthracis (Devi et al., 2017), B. subtilis (Hullo et al., 2007) and L. plantarum (Matoba et al., 2017) and showed production of cystathionine with the substrate O-acetylserine. This result thus indicated HP0107 to be exclusively an OCBS enzyme and to require O-acetylserine instead of L-serine as its first substrate for the reaction (Fig. 5).

HpOCBS releases hydrogen sulfide (H$_2$S)

CBSs and OCBS produce H$_2$S in the presence of L-cysteine and L-homocysteine via the β-replacement reaction (Chiku et al., 2009, Devi et al., 2017, Matoba et al., 2017). HpOCBS was also found to produce H$_2$S with $K_m$ and $k_{\text{cat}}$ values for L-homocysteine of 86 µM and 6.05 s$^{-1}$, respectively (Table 1 and Fig. 4b). The specificity constant of HpOCBS for H$_2$S generation was observed to be relatively high, when compared with other enzymatic activities of this enzyme. Similarly high efficiency for H$_2$S production has also been observed in B. anthracis (Devi et al., 2017) and L. plantarum (Matoba et al., 2017). HpOCBS was also able to produce H$_2$S using only substrate L-cysteine, which might be producing another product L-lanthionine as suggested by (Matoba et al., 2017) but with very low efficiency (data not shown), compared to the production of H$_2$S with the β-replacement reaction of cysteine and homocysteine that might be producing another product L-cystathionine (Fig. 5b) as reported (Matoba et al., 2017). The presence of L-homocysteine as a second substrate was found in the current work as well as in previous investigations with B. anthracis and L. plantarum to greatly enhance the production of H$_2$S. Inhibition of H$_2$S production has been shown to make pathogenic bacteria more susceptible to antibiotics (Shatalin et al., 2011). Thus H$_2$S was concluded to protect the bacteria from oxidative stress, and the higher $k_{\text{cat}}/K_m$ indicated its importance for survival of the bacteria.
**HpOCBS also shows O-acetylserine sulphydrylase (OASS) activity**

OCBS, like OASS, can consume H$_2$S to produce L-cysteine (Matoba et al., 2017, Devi et al., 2017). We found HP0107 showing OASS activity, but with a low specificity constant compared to its OCBS and H$_2$S generation activities. Its k$_{\text{cat}}$ was measured to be approximately 0.108 s$^{-1}$ (Table 1 and Fig. 4c), considerably less than the values for OASSs from other organisms (Bonner et al., 2005, Mizuno et al., 2002). OCBS and CBS from other organisms also displayed OASS activity, attributed to their three-dimensional structures and active site residues being similar to those of the OASS proteins.

**Three-dimensional structure of HpOCBS reveals its dimeric conformation**

The crystal structure of native HpOCBS was determined to a resolution of 1.9 Å (Fig. 6), and refined to an R and R$_{\text{free}}$ of 18% and 21%, respectively. Figure 6c shows the quality of the electron density of the PLP molecule and one of the representative regions of the protein. Over 96.4% of the 506 total number of amino acid residues refined in the most favourable region of the Ramachandran plot, and the remaining 3.6% of the residues in the additionally allowed region, i.e., with no residues in the outlier region (Table 2).

The HpOCBS protein crystallized as a dimer with extensive interactions between two protomers (Fig. 6a). The dimeric interface was observed to include both the N- and C-terminal domains. The association between the two monomers was observed to primarily take place through hydrophobic interactions, salt bridges, sulfur-aromatic amino acid interactions (of Phe17 of chain A with Met7 of chain B and chain B Phe17 with chain A Met7) and hydrogen bonds. Together this association resulted in a buried surface area of 3533 Å$^2$, equivalent to 13.4% of the total surface area of the monomer.

Each protomer was observed to consist of two non-contiguous $\alpha/\beta$ motifs, each having a topology similar to those of other type-II PLP enzymes (Fig. 6b). Inspection of the structure of the N-terminal domain indicated it to consist of residues 1-12 and 40-149, and to include $\alpha$-helices H1, H2, H3 and H4, $\beta$-strands B1, B4, B5, B6 and B7, and $\beta_{10}$ helices G1, G3 and G4, with the $\beta$-strands mostly parallel to each other and surrounded by helices H2 and H3 from one side and H4 from the other. Inspection of the structure of the C-terminal domain indicated it to consist of residues 13-39 and 150-306, and to include $\alpha$-helices H5, H6, H7, H8 and H9, $\beta$-strands B2, B3, B8, B9, B10 and B11, and $\beta_{10}$ helices G2, G5 and G6, with these strands also parallel to each other, and surrounded by helices H6 and H7 from one side and H8 and H9 from the other.

In the HpOCBS crystal structure, PLP cofactor was observed buried in a cleft between the two domains. The cofactor linked to the $\varepsilon$-amino group of Lys44 via a Schiff base linkage to form a lysine-pyridoxal-5'-phosphate (LLP) internal aldimine structure (Fig. 6c). The nitrogen of the pyridine ring (N1) formed a hydrogen bond with the oxygen of Ser267, and the OH (O3) of PLP formed an interaction with Asn76. The phosphate-binding loop or ‘G/T loop’ was observed to connect $\beta$8 and $\alpha$7 and to be composed of residues $^{180}$GSGG$^{184}$. These amino acid residues were found to anchor the phosphate moiety by forming a hydrogen bond.
bond network holding the PLP coenzyme firmly (Fig. S2). These residues and the associated hydrogen bonds and hydrophobic interactions near PLP were found to be mostly conserved or similar in other homologous CBS, OCBS and OASS structures.

**Methionine complexed with one of the protomers of HpOCBS shows a pre-reactive state**

In the HpOCBS crystal structure, methionine (substrate analog) was observed to be bound to one of the protomers of the HpOCBS dimer, at the active site near the PLP. This was, to the best of our knowledge, the first-ever OCBS structure reported with bound methionine along with an internal Schiff base (with PLP covalently linked to the ε-amino group of Lys). In most of the reported substrate-bound structures of this family, PLP is present as an external Schiff base, i.e., with PLP covalently linked to the α-amino group of the substrate or present as an aminoacylate intermediate (Koutmos et al., 2010, Schnell et al., 2007, Matoba et al., 2017). In two experiments involving OASS, one from *Salmonella typhimurium* (PDB ID: 1D6S) (Burkhard et al., 1999) and the other from *Arabidopsis thaliana* (PDB ID: 1Z7Y) (Bonner et al., 2005), the lysine normally bound to PLP (PLP-linked lysine-LLP) was mutated by the experimenters to Ala, and in these structures PLP makes a covalent bond with methionine (external aldime). In both of these structures, methionine was not added in the crystallization condition but OASS acquired it during the protein production, indicative of its affinity for methionine. Of all the available OASS structures, only that from *Entamoeba histolytica* has been reported to have methionine at the active site along with the internal aldime, and this structure was obtained using co-crystallization with the methionine (Raj et al., 2013). Binding constant (K_d) values of methionine were reported to be 78 ± 9 µM and 0.70 ± 0.06 mM, respectively, for the OASSs of *S. typhimurium* and *A. thaliana*, with the latter value comparable to the 0.252 ± 0.055 mM value measured here for HpOCBS (Fig. S3). Based in part on this result, HpOCBS may be predicted to have a high affinity for methionine and trapped methionine in the pre-reactive state from the media during protein production. The presence of methionine in the pre-reactive state in only one of the protomers (of the dimeric HpOCBS) and without it covalently bound to the PLP to form an external aldime were attributed to the suboptimal concentration of the methionine in our experiments. However, a high K_d value of 1.5 ± 0.4 mM was reported for methionine with *E. histolytica* OASS, indicative of the inability of this protein to easily take up methionine from media, and thus the need to have added it externally during the crystallization (Chinthalapudi et al., 2008).

Supersetion of the two protomers of the HpOCBS dimer revealed an approximately 0.78 Å root-mean-square deviation (RMSD) of backbone α-carbon atoms, similar to that found for L-serine-bound *D. melanogaster* CBS (Koutmos et al., 2010). Methionine binding led to a closing of the active site as a result of the movement of mobile loop 222-230 of the C-terminal domain and a nearly 10° bending of the N-terminal domain (Fig. 7c), making the two domains approach each other, and specifically decreasing the distance between the domains by 3.2 Å (Fig. 7c). Here, in the presence of methionine, Glu222 (conserved in OCBSs (Devi et al., 2017) and CBSs) formed interactions with Lys302 of the C-terminal tail and with the backbone amide of Ser101 of the N-terminal domain (Fig. 7d).
Methionine in HpOCBS was observed to form a hydrogen bond with the side chain of Lys104. A mutation of the corresponding lysine in yCBS (K112) was carried out previously by Lodha et al., 2009 to determine its importance in L-homocysteine binding, but surprisingly it yielded an increase in the $K_m$ of L-serine instead of L-homocysteine, indicating that this particular Lys binds to L-serine instead of L-homocysteine (Lodha et al., 2009). This conclusion was also consistent with the above-indicated observation of an interaction between methionine and Lys104 in the present methionine-bound HpOCBS structure. The backbone nitrogen atom of Asn76 of the TAGNT$^{77}$ ‘asparagine loop’ also made a hydrogen bond with the carboxyl group of methionine, while other residues of this loop formed hydrophobic interactions with methionine (Fig. 7a and b) and apparently promoted closing of the active site (Burkhard et al., 1999). Ser has been found to be conserved instead of Ala74 in the asparagine loops of CBSs, and mutation of Ala to Ser in OCBS yielded an increase in $H_2S$ generation (Devi et al., 2017, Matoba et al., 2017), indicating the involvement of the Ser residue in anchoring the substrate.

Gln146, conserved across OCBS, CBS and OASS sequences, was observed to interact with methionine in HpOCBS to form an internal Schiff base. A Gln157Ala mutation in yeast CBS abolished β-replacement reaction and this result confirmed the role of this residue in substrate positioning and in the subsequent reaction mechanism (Aitken & Kirsch, 2004). In the current HpOCBS crystal structure, Phe147 and Gly223 also interacts with the methionine (Fig. 7a and b). Also note that the residue corresponding to HpOCBS Phe147 is a Tyr in CBS, and a Tyr158Phe mutation in yeast CBS was previously shown to yield a 3-fold decrease in $k_{cat}/K_m$ for L-serine in the β-replacement reaction (Aitken & Kirsch, 2004). The hydroxyl group of this Tyr was reported to form a hydrogen bond network with the β-OH group of the substrate L-serine and other residues at the active site of CBS and hence to facilitate the β-replacement reaction (Koutmos et al., 2010).

**Structural comparison of HpOCBS with other homologous enzymes of the PLP-II family**

We compared the HpOCBS structure with the available structures of CBS, OCBS and OASS of the PLP-II family. CBS was found to have N- and C-terminal domains with three-dimensional structures similar to those of HpOCBS. In addition to these domains, CBSs from higher organisms also have C-terminal extended regulatory and N-terminal heme-binding domains (Omura et al., 1984, Finkelstein et al., 1975, Koracevic & Djordjevic, 1977). In mammals, CBS is activated by the binding of AdoMet to the regulatory domain (Prudova et al., 2006, Finkelstein et al., 1975, Koracevic & Djordjevic, 1977), while CBS proteins from different classes remain constitutively active and do not require AdoMet for the activation (Koutmos et al., 2010, Gimenez-Mascarell et al., 2017). Four CBS structures have been reported, from *H. sapiens* (Meier et al., 2001), *D. melanogaster* (Koutmos et al., 2010), *Apis mellifera* (Gimenez-Mascarell et al., 2017) and *S. cerevisiae* (Tu et al., 2018), and 3-dimensional structural superposition of these CBSs with HpOCBS showed RMSD values of approximately 2.0 Å for about 300 aligned Ca atoms of the core catalytic domain (i.e., N- and C-terminal domains) (Table 3 and Fig. 8b). Sequence comparison of CBSs with HpOCBS showed only 23 to 28 % sequence identity values (Table 3), with these low values due to the presence of extra residues at both the N- and C-termini.

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To date, only two OCBS structures have been published, from *B. anthracis* (PDB ID: 5XW3) (Devi et al., 2017) and *L. plantarum* (PDB ID: 5B1H ) (Matoba et al., 2017), and rigid body superpositions of these structures with the *HpOCBS* structure using the FATCAT server gave RMSD values of 3.0 Å and 1.15 Å, respectively. The very high RMSD between *HpOCBS* and *B. anthracis* OCBS was due to the large angle between the N- and C-terminal domains in the latter. On comparing the methionine-bound *H. pylori* OCBS structure (PDB ID: 6AHI) with the methionine-bound *L. plantarum* OCBS structure (PDB ID: 5B1I), we found the methionine in the *H. pylori* structure to be present as a pre-reactive state and not linked to PLP (Fig. 9b). This methionine is expected to adopt a conformation similar to the methionine of *L. plantarum* OCBS after making a Schiff base linkage with PLP.

Structural superposition of *HpOCBS* with known OASSs yielded RMSD values of 1.8 to 2.0 Å, indicating their structures to be highly similar; these superpositions also yielded very good structural alignments of their active site residues (Fig. 8a). Sequence comparisons of *HpOCBS* with OASSs showed only 33 to 40 % identities but with the OCBSs showed nearly 53 to 58 % identities (Table 3). These results clearly justified the placement of *HpOCBS* in a separate OCBS clad in the phylogenetic tree.

**Mode of OAS and L-homocysteine binding at active site**

To date, no CBS or OCBS structure with bound L-homocysteine has been published. Therefore, to gain an understanding of the modes of interactions of OAS and L-homocysteine at the active site for the β-replacement reaction, L-homocysteine was docked in *silico* in the closed conformation of the methionine-bound *HpOCBS* structure to identify the residues involved in the binding of L-homocysteine. Here, L-homocysteine bound between the asparagine loop and the loop containing residues 222 to 227 of the C-terminal domain (Fig. S4a). The docking produced hydrogen-bond interactions of the backbone of Gly223 and Gly225 with the α-carboxylate and α-amino group of L-homocysteine (Fig. S4b and c). These same residues formed interactions with a distal portion of L-cystathionine docked in yeast CBS, with this distal portion of L-cystathionine corresponding to the L-homocysteine moiety (Lodha et al., 2009). Residues Glu222, Ile224, Glu227 and His219 from the C-terminal domain and Phe100 and Ile124 from the N-terminal domain formed hydrophobic contacts with L-homocysteine (Fig. S4b and c). Methionine was observed to be located in between PLP and L-homocysteine, more towards the N-terminal domain below the asparagine loop (Fig. S4a).

**Mutational studies confirm the critical active site residue differences between conventional CBS and OCBS**

Conventional CBS prefers L-serine as its first substrate to carry out β-replacement reaction, while OCBSs exclusively require OAS for the reaction and do not show any functionality in the presence of the substrate L-serine (Devi et al., 2017), with this different behaviour towards OAS and L-serine perhaps due to the difference between certain residues at the active site of CBS and those of OCBS (Table S2 and Fig. 9a). Inspection of the *HpOCBS* and CBS structures specifically showed three residues located near the active site, at (*HpOCBS*) positions 74, 147 and 226, to differ between them —identified, respectively, as Ala, Phe, and
Val in HpOCBS, but Ser, Tyr and Tyr in CBS. The hydroxyl groups of the Ser and Tyr residues may provide necessary hydrogen bonding to the leaving hydroxyl (β-OH) group of the L-serine substrate and facilitate its β-elimination (Koutmos et al., 2010), while absence of these residues in OCBS, may be making it incapable of utilizing L-serine for the reactions and thus requiring activated L-serine, i.e., OAS, for its activity.

To confirm the identity of the residues crucial for the reaction mechanism, a series of mutations of HpOCBS were made (listed in Table S3). The β-replacement activity of each mutation was checked with the substrate L-serine or OAS in the presence of 5 mM of homocysteine. Specifically three structure-guided mutations, A74S, F147Y, and V226Y, were made to HpOCBS. In addition to these single-residue-substitution mutants, one double-residue-substitution mutant (F147Y/V226Y) and one triple-residue-substitution mutant (A74S/F147Y/V226Y) were also generated. All five of these mutants were generated with the expectation that they would show activity with the substrate L-serine, with this expectation due to the interactions made between the β-OH of L-serine-bound CBS structures (Koutmos et al., 2010) (Tu et al., 2018) and the homologous residues. However, none of the mutants showed β-replacement activity with the L-serine (Fig. S5), even though there was no structural change upon mutation (Fig. S6). The F147Y mutant showed activity with OAS but with less catalytic efficiency than did wild type (WT) HpOCBS. Surprisingly, the V226Y variant did not show any β-replacement activity with substrate OAS. We checked OCBS activity with a higher enzyme concentration (1.48 μM) of the HpOCBS V226Y mutant together with 5mM each of OAS and L-homocysteine; only slight activity was observed in this case. Also, neither the double mutant (F147Y/V226Y) nor the triple mutant (A74S/F147Y/V226Y) of HpOCBS showed any activity with substrate OAS even at the high enzyme concentration, again despite the lack of any considerable structural differences between the mutant and WT HpOCBS structures (Fig. S6).

To verify our hypothesis on the importance of these residues for substrate binding occurring via the β-replacement reaction mechanism, mutation was made to S. cerevisiae CBS (ScCBS) at the corresponding homologous residues listed in Table S3. ScCBS was chosen due to its lack of an N-terminal iron-binding region and due to the constitutive activity of its catalytic core domain, which has been shown to not require AdoMet for activation. Also, ScCBS has been shown to be stable for nearly 6-7 days at 4°C. ScCBS showed kcat and Km values for CBS activity to be 55.6 s⁻¹ and 2.26 mM respectively. The discrepancy in the kcat and Km values of CBS activity of ScCBS with the earlier report may have been due to the difference in the method employed for detecting cystathionine (Table 4); however the kcat/Km of ScCBS was determined to be 24.68 mM⁻¹s⁻¹, comparable to the previously reported value (Lodha et al., 2010). The double mutant (Y158F/Y248V) generated in ScCBS was tested for cystathionine generation in the presence of substrate L-serine and OAS, individually. The ScCBS double mutant (Y158F/Y248V) did not show any CBS activity with L-serine but showed enhanced activity in the presence of substrate OAS, with a kcat/Km of about 18.2 mM⁻¹s⁻¹ (Table 4), compared to the 12.6 mM⁻¹s⁻¹ value for the WT ScCBS OCBS activity (Fig. S5). Based on these results, we hypothesized that Y248 and Y158 are indispensable for CBS activity with L-serine not only in yeast CBS but in all CBSs, since these tyrosine residues are conserved across this sub-family. However, the OCBS activity displayed by the ScCBS

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double mutant (Y158F/Y248V) showed a higher specificity constant than did that displayed by WT ScCBS (Table 4 and Fig. S5), indicating the favorability of the active site architecture for OAS, hence validating the importance of these residues for OAS binding.

The absence of CBS activity with substrate L-serine by the triple mutant (A74S/F147Y/V226Y) and double mutant (F147Y/V226Y) of HpOCBS could be due to the different orientation adopted by Val226 in HpOCBS than that adopted by the homologous Tyr248 in ScCBS. In the HpOCBS crystal structure, Val226 faces away from the active site towards the C-terminal domain and its movement seems to be restricted due to the presence of Glu227 which interacts with the residues of N-terminal domain in a closed conformation upon substrate binding (methionine-bound protomer of HpOCBS) (Matoba et al., 2017). Thus even upon mutation of Val226 to Tyr in HpOCBS, this residue would still presumably be facing away from the active site and hence unable to contact the substrate L-serine.

**Discussion**

The reverse transsulfuration pathway is well characterised in higher organisms such as *H. sapiens* (Ereno-Orbea et al., 2013, Ereno-Orbea et al., 2014, McCorvie et al., 2014), *S. cerevisiae* (Jhee et al., 2000), *A. mellifera* (Gimenez-Mascarell et al., 2017) and *D. melanogaster* (Koutmos et al., 2010), where it is the sole source of L-cysteine production from methionine. In bacteria, the reverse transsulfuration pathway is also known as the ‘methionine-to-cysteine conversion pathway’. CBS and CGL have been named, respectively, as MccA and MccB enzymes of the methionine-to-cysteine conversion pathway. This pathway is not universally present in bacteria but study shows some bacteria possess enzymes to carry out methionine-to-cysteine conversion. In *Lactobacillus casei*, the expression levels of *cbs* and *cgl* genes decrease in cysteine-containing media; these genes are present in a reading frame along with the gene for serine O-acetyl transferase (Bogicevic et al., 2012). Various *Lactobacillus paracasei* strains possess the highly mobile gene cluster *cysK-ctl-cysE* responsible for the conversion of methionine to cysteine in dairy products, which are highly rich in methionine compared to cysteine (Wuthrich et al., 2018). Similarly, *Klebsiella pneumoniae* uses its mtbBC operon to produce cystathionine β-synthase and cystathionine γ-lyase (Seiflein & Lawrence, 2006) for the production of cysteine from methionine. *Mycobacterium tuberculosis* has a bifunctional gene (rv1079), which allows this organism to carry out both cystathionine γ-lyase and cystathionine γ-synthase activities and use methionine as a sole sulfur source, and hence demonstrate a functional reverse transsulfuration pathway (Wheeler et al., 2005). *Bacillus subtilis* possesses CBS, which shows activity with OAS and not with serine and can use methionine as a sole source of sulfur since it also has the *cgl* gene (Hullo et al., 2007). Recent investigations have also shown the importance of CBS for the survivability and pathogenicity of bacterial pathogens under oxidative stress (Shatalin et al., 2011, Devi et al., 2017, Matoba et al., 2017).

The present study along with earlier studies showed the presence of the OCBS gene in the genomes of some bacteria, and showed these bacterial OCBSs to differ from the CBSs found in higher organisms in their domain architecture, length, regulation and most importantly substrate requirements. These studies showed bacterial OCBSs to function in the presence of...
OAS instead of L-serine to produce L-cystathionine, to not require AdoMet for its activation, and to also be devoid of a heme-binding domain and C-terminal regulatory domain (Devi et al., 2017, Matoba et al., 2017). OCBS was shown to be widely found in bacteria, and to share a high similarity with OASS, and in fact has been misannotated as either cysK or cysteine synthase (OASS). OASS has been shown to be involved in different activities in addition to its role in cysteine production in the de novo pathway, such as an antibacterial nuclease toxin produced by Escherichia coli (Campanini et al., 2015) and in transcription repression in B. subtilis (Tanous et al., 2008). OCBS has apparently maintained its integrity at the sequence level, showing no mixing either with OASS and CBS in the phylogenetic tree (Fig. 2). The H. pylori hp0107 gene is annotated as an OASS in the NCBI database but the protein encoded by this gene was found in the current work to show OCBS activity and to also display O-acetylserine sulfhydrylase activity, albeit at a relatively low level, a feature observed for other OCBSs as well (Matoba et al., 2017, Devi et al., 2017). Based on the phylogenetic analysis and analysis of the HpOCBS structure with bound methionine (substrate analog) at the active site, we concluded the present hp0107 gene to actually code for an OCBS.

An in depth structural comparison of OCBS with conventional CBS, which uses L-serine, revealed possible key residues that determine the respective substrate preferences of these two proteins. Analysis of the active sites was performed by using L-serine-bound S. cerevisiae CBS (PDB ID: 6C2Q) (Tu et al., 2018) and methionine-bound HpOCBS. We identified three residues that may play important roles in selecting L-serine as the substrate for the reaction performed by CBS. Inspection of the ScCBS structure showed Ser82 and Tyr248 forming hydrogen bonds with the hydroxyl group of the substrate L-serine, and this substrate covalently linked to PLP (forming an external aldimine). A network of polar interactions of Tyr158 and Tyr248 would appear to also stabilize the reaction intermediate (Fig. 9a). Inspection of conventional CBS also showed its active site cavity to be polar, due to the presence of two tyrosine residues and one serine residue, with these three residues ultimately making polar interactions with the hydroxyl group of the L-serine substrate and participating in its activation and dehydroxylation. Analysis of the OCBS structures showed them to contain a conserved Ala74 (residue number according to H. pylori OCBS) instead of Ser82 in the ‘TSGNT’ loop, while showing Tyr158 replaced by Phe147 and Val226 (or Met or any other hydrophobic amino acid) at the homologous position of Tyr248 in the ‘EGIGY’ loop (Fig. 9a). These residues make the active site of OCBS hydrophobic and unable to participate in the activation of L-serine. Interestingly, the ScCBS double mutant containing Phe and Val instead of Tyr (Y158F/Y248V) behaved like an OCBS and showed β-replacement activity only with substrate OAS. Perhaps CBS evolved from bacterial OCBS to take L-serine instead of OAS as its substrate, with evidence for this speculation including the observed lack of the enzyme to produce OAS in higher organisms and the measured retention of ancestral OCBS activity in the ScCBS mutants.

Nevertheless, to complete the reverse transsulfuration pathway, another enzyme known as cystathionine γ-lyase (CGL) is required (Banerjee et al., 2003, Miles & Kraus, 2004). In H. pylori, the hp0106 gene (annotated as cystathionine γ-synthase) is in the same gene cluster as that encoding OCBS but has been predicted to function as a CGL (Doherty et al., 2010). Our
investigation also revealed the ability of HP0106 to efficiently convert cystathionine to cysteine (data not shown). Thus functional OCBS and CGL are present in *H. pylori* and a complete reverse transsulfuration pathway is functional in this pathogen.

**Experimental procedures**

*In silico analysis*

Various OCBS protein sequences were retrieved using PSI-Blast with *B. anthracis* O-acetylserine-dependent CBS (*BaOCBS*) as the search template (Devi *et al.*, 2017). Furthermore, a few resultant sequences were used as search templates to retrieve additional distantly related OCBSs present in various phyla of bacteria. We used here only a few protein sequences from any particular genus. This means that there are many such proteins present in different species of the same genus. CBS proteins present in different organisms were retrieved using *H. sapiens* CBS (Ereno-Orbea *et al.*, 2014) as a template in PSI-Blast. However, annotated CBSs present in a few bacteria were retrieved from the NCBI database manually. Similarly, OASS sequences from bacteria and plants were also extracted using *E. histolytica* OASS (PDB ID: 2PQM) as a template sequence in PSI-Blast (Chinthalapudi *et al.*, 2008). Finally, a total of 154 sequences were used for the construction of the phylogenetic tree. Initially a multiple sequence alignment was done using Clustal Omega with all of the sequences. The aligned sequences were then used as input into MEGA 5.1 software and a phylogenetic tree was generated using maximum likelihood based on the default parameters with 250 bootstraps (Tamura *et al.*, 2011).

Furthermore, multiple sequence alignment was done using the MULTALIN tool to obtain a consensus sequence for the members of the CBS, OASS and OCBS families (Corpet, 1988). All sequences from one particular family were input into the server for the multiple sequence alignment, which used the BLOSUM62 comparison table. The output file gave the aligned sequences along with the consensus sequence, which included residues conserved in more than 50% of the sequences.

**Amplification, cloning and site-directed mutagenesis of HP0107 and ScCBS**

The *hp0107* gene was amplified by performing the polymerase chain reaction (PCR) on the genomic DNA of *H. pylori* 26695 as a template using gene specific primers (Table 5) and Phusion High-Fidelity DNA Polymerase (Thermo Scientific™). The amplicon of the 921 bp gene was first purified using a PCR purification kit (Qiagen, Germany) and digested with NcoI and XhoI at 37°C for five hours. The pET28b expression vector was also digested with NcoI and XhoI, and the digested vector was purified using a gel extraction kit (Qiagen, Germany). The purified *hp0107* gene was ligated, using T4 DNA ligase, into the pET28b expression vector at the NcoI and XhoI sites with a C-terminal (His)$_6$ tag. The positive construct was selected and screened using a Luria-Bertani (LB) agar plate supplemented with kanamycin (50 μg·ml$^{-1}$) and colony PCR, respectively. The site-directed mutagenesis was performed on a pET28b-*hp0107* plasmid using mutagenic primers for the corresponding mutations listed in Table 5. In brief, pET28b-*hp0107* was used as a template for PCR to
generate variants. Following the amplification of the whole plasmid, parental methylated DNA was digested using DpnI enzyme. The digested mixture was directly transformed in the DH5α strain of E. coli. All of the intended mutations were confirmed by carrying out whole gene sequencing.

Similarly, amplification of the catalytic core region of the Sccbs gene was done by PCR on genomic DNA of S. cerevisiae with the primers listed in Table 5, followed by cloning of Sccbs in pET28b at the NheI and XhoI sites. Furthermore, site-directed mutagenesis was performed on the Sccbs gene as described above.

**Overexpression and purification of HP0107 and ScCBS**

Plasmids of pET28b-hp0107 and pET28b-Sccbs were transformed in the BL21 (DE3) strain of E. coli for the protein expression. The colonies were picked from LB plates containing kanamycin and inoculated into 50 ml of LB media for overnight growth. One percent of this culture was inoculated into one litre of secondary culture containing 50 μg·ml⁻¹ of kanamycin. The culture was incubated at 37°C until the OD₆₀₀ reached 0.6 to 0.8. Gene expression was initiated with the addition of isopropyl β-D-thiogalactoside (IPTG) to a final concentration of 250 μM, the temperature was lowered to 16°C after induction, and the culture was allowed to grow for another 12 h. Cells were harvested at 4°C by subjecting them to centrifugation at 7500 g for 5 minutes and then resuspending the pellet in buffer A containing 50 mM Tris [pH7.5], 150 mM NaCl, 1 mM phenylmethanesulfonyl fluoride (PMSF), 0.1% Triton X-100, 5 mM imidazole, 0.01 mg·ml⁻¹ lysozyme and 5 mM β-mercaptoethanol (βMe). Then, cells were sonicated and centrifuged at 25 000 g for 1 h. The cleared supernatant was applied onto Ni-NTA Sepharose column that had previously been equilibrated with 50 ml buffer B (50 mM Tris [pH 7.5], 150 mM NaCl, 5 mM βMe and 5 mM imidazole). The protein was eluted in buffer C containing 50 mM Tris [pH7.5], 150 mM NaCl, 300 mM imidazole, 5% glycerol and 5 mM βMe.

For further purification and removal of imidazole, the protein was concentrated using a Centricron filter with 30 kDa cut-off and loaded onto a HiLoad 16/60 Superdex 200 column (GE Healthcare) and eluted at a flow rate of 1 ml·min⁻¹. The column that had previously been equilibrated with running buffer D containing 10 mM Tris [pH 7.5], 150 mM NaCl, 5% glycerol and 5 mM βMe. The purity of the desired protein was checked using 12% SDS-PAGE and purified fractions were pooled and concentrated using a Centricron filter (cut-off 30 kDa). Nearly 95 to 99% pure protein was obtained from gel-filtration chromatography (GFC) and purified protein was used to perform all of the experiments.

Purification of the hp0107 mutant proteins were done using a method similar to that used for WT HP0107 (HpOCBS), and an SDS-PAGE analysis of the purified proteins obtained after GFC showed 90 to 99% pure protein (Fig. S7). The expression levels of the HpOCBS A74S and HpOCBS F147Y single mutants were similar to that of WT HpOCBS, each approximately 8 to 10 mg of protein from one litre of culture, while a mass of only 3 to 4 mg of protein was obtained from the same volume of culture for the HpOCBS V226Y mutant.
and the HpOCBS double (F147Y/V226Y) mutant (Fig. S8). The expression level of the HpOCBS triple mutant (A74S/F147Y/V226Y) was 6 to 7 mg from one litre of culture (Fig. S8), better than that of the HpOCBS double mutant but poorer than that of WT HpOCBS. Furthermore, the HpOCBS V226Y mutant and HpOCBS double (F147Y/V226Y) mutant were observed to be less stable that the WT HpOCBS and tended to precipitate when kept at 4°C for more than 3 to 4 days, but the HpOCBS A74S and HpOCBS F147Y mutants were found to be as stable as HpOCBS. The HpOCBS triple mutant (A74S/F147Y/V226Y) was observed to be more stable than the HpOCBS double mutant (F147Y/V226Y) and remained in solution for 5 to 6 days at 4°C. The stability of this triple mutant may have been provided in part by the hydrogen bonding made between Ser at position 74 and the PLP.

WT ScCBS and its ScCBS double (Y158F/Y428V) mutant were expressed at high levels, approximately 25 to 30 mg of protein from 500 mL of culture. No significant difference was observed between the expression and stability of the ScCBS double mutant and those of WT ScCBS (Fig. S9).

Biochemical characterisation

O-acetylserine-dependent CBS (OCBS) assay

An O-acetylserine-dependent CBS assay was conducted using a colorimetric method as described earlier for CBS (Kashiwamata & Greenberg, 1970). An OCBS enzyme assay was performed in a final volume of 200 µl with final concentrations of 0.367 µM for HpOCBS, 0.25 mM for pyridoxal 5’-phosphate and various concentrations for OAS in 50 mM HEPES [pH 7.5] buffer, while keeping the L-homocysteine concentration constant at 5 mM, to determine the K_m for OAS. In each experiment, the reaction mixture was incubated for 5 minutes at 37°C, and then the reaction was stopped with the addition of ninhydrin reagent directly into the reaction mixture. The reaction solution was heated for 5 minutes on a boiling water bath and then incubated for 3 minutes on ice, after which it was left to develop colour for 20 minutes at 25°C. The absorbance of the solution was measured at a wavelength of 451 nm and from this absorbance value was subtracted the absorbance of an enzyme-free blank. Initially, this assay was performed with various buffers of different pH values (50 mM each of sodium acetate [pH 4.0 to 5.0], Bis-Tris [pH 5.5 to 6.5], HEPES [pH 7.0 to 8.1] and Tris-HCl [pH 8.5 to 8.8]) to determine the optimal pH for the OCBS activity. In initial assays, the concentrations of L-homocysteine and OAS were kept constant at 1 mM and 5 mM, respectively. The amount of cystathionine produced in a reaction was calculated from the standard curve of the cystathionine (50 to 5000 µM). All results shown represent mean ± SD of three (n=3) independent experiments. Data obtained from all of the above assays were fitted by the Michaelis-Menten equation $v = \frac{V_{max}[OAS]}{(K_m+[OAS])}$ using GraphPad Prism 5 to calculate the K_m and V_max values of the reactions.

Cystathionine β-synthase (CBS) assay

CBS assay was also carried out with substrate L-serine and L-homocysteine as described above for the OCBS assay. All of the reactions with HpOCBS and its variants were

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performed in 50 mM HEPES [pH 7.5] with a final enzyme concentration of 0.367 µM. Reactions were performed with a constant amount of homocysteine (5 mM) and various concentrations of freshly prepared L-serine along with 0.25 mM of PLP. Reaction mixtures were incubated at 37 °C for 5 minutes and stopped with the addition of ninhydrin.

The CBS and OCBS assays performed with the ScCBS protein used 0.30 µM of the ScCBS enzyme, 0.25 mM pyridoxal 5′-phosphate and 5 mM of L-homocysteine in 50 mM of Tris [pH 8.6] buffer as done earlier (Lodha et al., 2009).

Data obtained from the CBS assay were fitted by the Michaelis-Menten equation $v = \frac{V_{\text{max}} [\text{L-Ser}]}{K_m+[\text{L-Ser}]}$ using GraphPad Prism 5 to calculate the $K_m$ and $V_{\text{max}}$ values of the reactions while the data for the OCBS assay were fitted by the equation $v = \frac{V_{\text{max}}[\text{OAS}]}{K_m+[\text{OAS}]}$ using GraphPad Prism 5 to calculate the $K_m$ and $V_{\text{max}}$ values of these reactions. Each result shown is the mean ± SD of three (n=3) independent experiments.

H$_2$S production assay

OCBSs and CBSs produce H$_2$S via the β-replacement reaction with L-cysteine and L-homocysteine as substrates (Devi et al., 2017, Banerjee et al., 2003, Matoba et al., 2017). The production of H$_2$S catalyzed by HpOCBS was monitored by measuring the absorption (at $\lambda=390$ nm) due to lead sulfide formed from the reaction of H$_2$S with lead acetate. A reaction mixture of 400 µl containing 0.4 mM lead acetate, various concentrations of L-homocysteine, and a fixed concentration of L-cysteine (1 mM) in 50 mM HEPES [pH 7.5] was preincubated at 25°C. The reaction was started by adding 10 µg (final concentration 0.367µM) of enzyme, and the absorption was monitored at a wavelength of 390 nm for 2 minutes. A molar extinction coefficient of 5,500 M$^{-1}$cm$^{-1}$ was used to determine the amount of lead sulfide produced (Chiku et al., 2009). Each result shown is the mean ± SD of three (n=3) independent experiments.

The Michaelis-Menten equation $v = \frac{(V_{\text{max}} [\text{L-homocysteine}])}{(K_m+[\text{L-homocysteine}])}$ was used to fit the data obtained from the H$_2$S production assay and hence calculate the $K_m$ and $V_{\text{max}}$ values of the reaction.

O-acetylserine sulphydrylase (OASS) assay

The OASS activity was determined as described previously (Kumar et al., 2011). Briefly, the reaction mixtures (final volume 400 µL) contained 1.58 µM (final concentration) of enzyme, 100 µM of 5-thio-2-nitrobenzoate TNB (alternative substrate) and various concentrations of OAS (6.75 to 250 µM) in 50 mM HEPES [pH 7.5] buffer. The reaction was initiated with the addition of OAS to the mixture of enzyme and TNB, and readings were taken immediately at a wavelength 412 nm for 3 minutes. The extinction coefficient of the TNB ($A_{412}$; ε=13,600 M$^{-1}$cm$^{-1}$) was used to calculate the rates of the reactions (Bonner et al., 2005). All of the experiments were done at 25°C and the results shown each represented a mean ± SD of three (n=3) independent experiments.

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Data obtained from the OASS assay were fitted by the Michaelis-Menten equation \( v = \frac{V_{\text{max}}[\text{OAS}]}{K_m+[\text{OAS}]} \) using GraphPad Prism 5 to calculate the \( K_m \) and \( V_{\text{max}} \) values of the reactions.

**Methionine binding**

Binding of methionine to OASS results in the formation of an external aldimine with PLP. Methionine binding to \( Hp\text{OCBS} \) was measured using a spectrofluorometer to monitor the changes in fluorescence spectra of this complex during the course of the binding. Spectra were taken from 450 nm to 600 nm, upon excitation at wavelength 412 nm, while keeping the slit width at 5 nm. Titration of \( Hp\text{OCBS} \) (100 µg) with methionine (0 to 700 µM) was performed in 50 mM HEPES [pH 7.5] buffer and an appropriate concentration of methionine was employed to achieve the saturation curve. All of the experiments were performed at 25°C. The maximum change in the fluorescence was observed at 516 nm. The binding constant \( (K_d) \) of methionine was calculated by fitting to the data a reversible two-state model of binding using the equation \( \Delta F = \frac{\Delta F_{\text{max}}[M]}{K_d + [M]} \), where \( \Delta F \) represents the change in fluorescence at a particular wavelength upon addition of methionine at concentration \([M]\) which is corrected for dilution (Bonner et al., 2005, Banerjee et al., 2011). \( \Delta F_{\text{max}} \) represents the maximum change in the fluorescence at that particular wavelength. The binding constant \( (K_d) \) of methionine for \( Hp\text{OCBS} \) was calculated to be 0.252 ± 0.055 mM (Fig. S3).

**Crystallization and X-ray diffraction data collection**

The purified \( Hp\text{OCBS} \) protein was concentrated up to 8 mg·ml\(^{-1}\) in a solution containing 10 mM Tris [pH 7.5] with 150 mM NaCl, 5% glycerol and 5 mM βMe. After continuous optimization using the hanging-drop vapour-diffusion method, crystals appeared in 3 to 4 days at 16°C using a mixture of 15% PEG 20000, 15% PEG MME 550, and 0.03 M NPS (sodium nitrate, disodium hydrogen phosphate and ammonium sulfate) at pH 6.5 (0.1 M MES/imidazole) as the precipitant, this condition was similar to the Morpheus crystallization screen condition number C1 (Gorrec, 2009).

The needle-shaped crystals that formed were carefully picked up, mounted in cryoloops, and flash frozen in liquid nitrogen at 100 K. These flash-frozen crystals diffracted X-rays (\( λ=0.978Å \)) to a resolution of 1.9 Å at ESRF BM14, Grenoble, France (Table 2). The data were indexed and processed using HKL2000 (Otwinowski & Minor, 1997). These crystals were shown to belong to the space group P2\(_2\)1\(_2\)1 with two \( Hp\text{OCBS} \) protomers per asymmetric unit.

**Structure solution and refinement**

The \( Hp\text{OCBS} \) structure was determined by carrying out molecular replacement using the structure of O-acetylserine sulfhydrylase from *Arabidopsis thaliana* (AtCS) (PDB ID: 1Z7W, 43% sequence identity). This AtCS structure and the aligned sequences of \( Hp\text{OCBS} \) and AtCS were input into CHAINSAW (Schwarzenbacher et al., 2004) to produce the specific search model used for molecular replacement, which was then carried out using Phaser
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Docking of L-homocysteine at the active site of HpOCBS structure

The Met-bound HpOCBS structure was used for the docking of L-homocysteine at the active site. First, we prepared the HpOCBS protein structure for the docking using the Schrodinger Protein Preparation Wizard program with the default parameters. This program was used to add missing hydrogen atoms, remove water molecules beyond 5 Å of HET groups, determine the protonated state for histidine residues, and highlight the missing residues. Further energy minimization and optimization of the protein structure was done using an OPLS2005 force field with a maximum RMSD of 0.30 Å for atom displacement to terminate the minimization. A ligand-binding site or ligand receptor grid was created in the energy-minimized structure of HpOCBS near the PLP. The ligand receptor grid covered all of the expected residues of the active site from the N- and C-terminal domains of HpOCBS. The L-homocysteine substrate was prepared using Schrodinger Ligprep Wizard before docking; here, hydrogen atoms were added, followed by energy minimization. Finally, docking of substrate in the HpOCBS receptor grid was performed using the program GLIDE with the extra precision (XP) module.

Accession number
The accession number (PDB ID) for HpOCBS structure reported in this paper is 6AHI

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Declarations of interest
All authors declare no conflict of interest.

Authors’ contributions
SG, SD and KFT conceived of the aim of the work and designed the experiments. KFT, SD and SAAR solved the HpOCBS structure. SD and MFA performed all experiments. SD, KFT and MFA wrote the manuscript taking input from SG and SAAR. All of the authors reviewed the manuscript.
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### TABLES

**Table 1. Various kinetics parameters of the reactions catalyzed by HpOCBS**

<table>
<thead>
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<th>Substrate</th>
<th>$V_{\text{max}}$ (µM s$^{-1}$)</th>
<th>$K_{m}$ (mM)</th>
<th>$k_{\text{cat}}$ (s$^{-1}$)</th>
<th>$k_{\text{cat}}/K_{m}$ (mM$^{-1}$s$^{-1}$)</th>
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<tr>
<td><strong>OCBS activity</strong>$^a$</td>
<td>O-acetylserine</td>
<td>13.51 ± 0.52</td>
<td>2.92 ± 0.28</td>
<td>36.8 ± 1.4</td>
<td>12.6 ± 1.31</td>
</tr>
<tr>
<td><strong>CBS activity</strong>$^a$</td>
<td>L-serine</td>
<td>NA $^4$</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td><strong>H$_2$S-production activity</strong>$^b$</td>
<td>L-homocysteine</td>
<td>2.22 ± 0.05</td>
<td>0.086 ± 0.007</td>
<td>6.05 ± 0.13</td>
<td>70.33 ± 5.98</td>
</tr>
<tr>
<td><strong>OASS activity</strong>$^c$</td>
<td>O-acetylserine</td>
<td>0.17 ± 0.006</td>
<td>0.025 ± 0.003</td>
<td>0.108 ± 0.004</td>
<td>4.42 ± 0.17</td>
</tr>
</tbody>
</table>

$^4$ NA- No detectable activity. $^a$OCBS and CBS kinetics experiments were carried out in 50 mM HEPES [pH 7.5] buffer at 37°C. $^b$ H$_2$S-production activity experiments were carried out in 50 mM HEPES [pH 7.5] buffer at 25°C. $^c$ OASS activity was determined in 50mM HEPES [pH 7.5] buffer at 25°C. Each result shown is the mean ± SD of three (n=3) independent experiments.
Table 2. *HpOCBS* crystallographic statistics

<table>
<thead>
<tr>
<th>Data statistics</th>
<th><em>HpOCBS</em></th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Crystallographic data</strong></td>
<td></td>
</tr>
<tr>
<td>Diffraction source</td>
<td>ESRF beamline BM14</td>
</tr>
<tr>
<td>Wavelength (Å)</td>
<td>0.978</td>
</tr>
<tr>
<td>Space group</td>
<td><em>P2</em>2<em>1</em>2<em>1</em></td>
</tr>
<tr>
<td><em>a</em>, <em>b</em>, <em>c</em> (Å)</td>
<td>71.4, 82.7, 96.1</td>
</tr>
<tr>
<td><em>α</em>, <em>β</em>, <em>γ</em> (°)</td>
<td>90, 90, 90</td>
</tr>
<tr>
<td>Resolution range (Å)</td>
<td>50–1.9 (1.93–1.9)*¶</td>
</tr>
<tr>
<td><em>R</em>&lt;sub&gt;r.i.m.&lt;/sub&gt;</td>
<td>0.121 (0.88)</td>
</tr>
<tr>
<td><em>R</em>-meas</td>
<td>0.117 (0.862)</td>
</tr>
<tr>
<td>Completeness (%)</td>
<td>100 (100)</td>
</tr>
<tr>
<td>Redundancy</td>
<td>16.3 (15.8)</td>
</tr>
<tr>
<td>⟨<em>I</em>/σ(<em>I</em>)⟩</td>
<td>47 (4.4)</td>
</tr>
<tr>
<td>Overall <em>B</em> factor from Wilson plot (Å&lt;sup&gt;2&lt;/sup&gt;)</td>
<td>24.4</td>
</tr>
<tr>
<td>CC1/2</td>
<td>0.99 (0.97)</td>
</tr>
<tr>
<td>No. of molecules per asymmetric unit</td>
<td>2</td>
</tr>
<tr>
<td><strong>Refinement</strong></td>
<td></td>
</tr>
<tr>
<td>Total No. of reflections</td>
<td>741356</td>
</tr>
<tr>
<td>No. of unique reflections</td>
<td>45368</td>
</tr>
<tr>
<td>Final <em>R</em>&lt;sub&gt;crys&lt;/sub&gt;</td>
<td>0.183</td>
</tr>
<tr>
<td>Final <em>R</em>&lt;sub&gt;free&lt;/sub&gt;</td>
<td>0.207</td>
</tr>
<tr>
<td>No of protein/water atoms</td>
<td>4667/132</td>
</tr>
<tr>
<td><strong>R.m.s. deviations</strong></td>
<td></td>
</tr>
<tr>
<td>Bonds (Å)</td>
<td>0.010</td>
</tr>
<tr>
<td>Angles (°)</td>
<td>1.469</td>
</tr>
<tr>
<td>Average <em>B</em> factors (Å&lt;sup&gt;2&lt;/sup&gt;)</td>
<td></td>
</tr>
<tr>
<td>Protein</td>
<td>37.5</td>
</tr>
<tr>
<td>Water</td>
<td>33.4</td>
</tr>
<tr>
<td><strong>Ramachandran plot</strong></td>
<td></td>
</tr>
<tr>
<td>Favoured (%)</td>
<td>96.4</td>
</tr>
<tr>
<td>Allowed (%)</td>
<td>3.6</td>
</tr>
</tbody>
</table>

*¶* Values for the highest-resolution shell are shown in parentheses.
Table 3. Comparisons of the structure and sequence of HpOCBS with those available for OCBS, OASS and CBS. Table shows RMSDs of Cα atoms with number of equivalent residues and percentage sequence identity values.

<table>
<thead>
<tr>
<th></th>
<th>PDB ID</th>
<th>RMSD (Å)</th>
<th>Equivalent residues</th>
<th>Sequence identity (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>OCBS</td>
<td>5B1H (Matoba et al., 2017)</td>
<td>1.15</td>
<td>301</td>
<td>58.1</td>
</tr>
<tr>
<td></td>
<td>5XW3 (Devi et al., 2017)</td>
<td>3.06</td>
<td>257</td>
<td>54.9</td>
</tr>
<tr>
<td>OASS</td>
<td>1OAS (Burkhard et al., 2000)</td>
<td>1.98</td>
<td>299</td>
<td>37.5</td>
</tr>
<tr>
<td></td>
<td>1VE1</td>
<td>1.69</td>
<td>298</td>
<td>37.2</td>
</tr>
<tr>
<td></td>
<td>1Z7W (Bonner et al., 2005)</td>
<td>1.80</td>
<td>303</td>
<td>40.2</td>
</tr>
<tr>
<td></td>
<td>2JC3 ( Chattopadhyay et al., 2007)</td>
<td>1.88</td>
<td>289</td>
<td>34.9</td>
</tr>
<tr>
<td></td>
<td>2PQM (Chinthalapudi et al., 2008)</td>
<td>1.87</td>
<td>304</td>
<td>35.4</td>
</tr>
<tr>
<td></td>
<td>2Q3B ( Schnell et al., 2007)</td>
<td>1.96</td>
<td>297</td>
<td>40.8</td>
</tr>
<tr>
<td></td>
<td>3SPX ( Raj et al., 2012)</td>
<td>1.85</td>
<td>302</td>
<td>35.2</td>
</tr>
<tr>
<td></td>
<td>5JIS (Dharavath et al., 2017)</td>
<td>2.01</td>
<td>302</td>
<td>33.9</td>
</tr>
<tr>
<td>CBS</td>
<td>1JBQ (Meier et al., 2001)</td>
<td>1.93</td>
<td>301</td>
<td>28.8</td>
</tr>
<tr>
<td></td>
<td>3PC2 (Koutmos et al., 2010)</td>
<td>1.98</td>
<td>305</td>
<td>23.2</td>
</tr>
<tr>
<td></td>
<td>5OHX (Gimenez-Mascarell et al., 2017)</td>
<td>2.00</td>
<td>305</td>
<td>27.1</td>
</tr>
<tr>
<td></td>
<td>6C2H(Tu et al., 2018)</td>
<td>2.10</td>
<td>304</td>
<td>33.8</td>
</tr>
</tbody>
</table>

[^]O-acetylserine sulfhydrylase from *Thermus thermophilus HB8*
Table 4. Kinetic parameters of *HpOCBS* variants, and of *ScCBS* WT and its variants.

<table>
<thead>
<tr>
<th>Protein</th>
<th>Variant</th>
<th>Substrate</th>
<th>$V_{\text{max}}$ (µM s$^{-1}$)</th>
<th>$K_{\text{m}}$ (mM)</th>
<th>$k_{\text{cat}}$ (s$^{-1}$)</th>
<th>$k_{\text{cat}}/K_{\text{m}}$ (mM$^{-1}$ s$^{-1}$)</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>HpOCBS</em>&lt;sup&gt;a&lt;/sup&gt;</td>
<td>A74S</td>
<td>OAS</td>
<td>8.43 ± 0.56</td>
<td>1.78 ± 0.39</td>
<td>22.96 ± 1.53</td>
<td>12.89 ± 2.96</td>
</tr>
<tr>
<td></td>
<td>L-serine</td>
<td>NA&lt;sup&gt;b&lt;/sup&gt;</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td></td>
<td>F147Y</td>
<td>OAS</td>
<td>5.77 ± 0.32</td>
<td>1.57 ± 0.29</td>
<td>15.72 ± 0.87</td>
<td>10.04 ± 1.92</td>
</tr>
<tr>
<td></td>
<td>L-serine</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td></td>
<td>V226Y</td>
<td>OAS</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td></td>
<td>L-serine</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td></td>
<td>F147Y/V226Y</td>
<td>OAS</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td></td>
<td>L-serine</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td></td>
<td>A74S/F147Y/V226Y</td>
<td>OAS</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td></td>
<td>L-serine</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td><em>ScCBS</em>&lt;sup&gt;b&lt;/sup&gt;</td>
<td>WT</td>
<td>OAS</td>
<td>6.89 ± 0.67</td>
<td>1.82 ± 0.43</td>
<td>22.50 ± 2.20</td>
<td>12.35 ± 3.13</td>
</tr>
<tr>
<td></td>
<td>L-serine</td>
<td>17.03 ± 1.15</td>
<td>2.26 ± 0.37</td>
<td>55.65 ± 3.75</td>
<td>24.68 ± 4.41</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Y158F/Y248V</td>
<td>OAS</td>
<td>14.67 ± 1.06</td>
<td>2.69 ± 0.53</td>
<td>47.94 ± 3.47</td>
<td>17.85 ± 9.47</td>
</tr>
</tbody>
</table>

NA-No detectable activity. *OCBS and CBS kinetics experiments were carried out in 50 mM HEPES [pH 7.5] buffer at 37 °C for WT *HpOCBS* and mutant proteins. *OCBS and CBS kinetics experiments of *ScCBS* and its mutant were carried out in 50 mM Tris-HCl [pH 8.6] buffer at 37 °C. Each result shown is the mean ± SD of three (n=3) independent experiments.

Table 5. Primers sequences used in present study.

<table>
<thead>
<tr>
<th>Primer</th>
<th>Sequences</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>HpOCBS_Fp</em>&lt;sup&gt;c&lt;/sup&gt;</td>
<td>5'-CATGCCATGGAATGATTATCATACCACAATG-3'</td>
</tr>
<tr>
<td><em>HpOCBS_Rp</em>&lt;sup&gt;c&lt;/sup&gt;</td>
<td>5'-CCGGCTCGAGTAAATAATACCTTTTGAGAGATAA-3'</td>
</tr>
<tr>
<td><em>HpOCBS_A74S_Fp</em></td>
<td>5'-CATCATGTAGGCTACCAACGGCAGACCGCAGGTATCG-3'</td>
</tr>
<tr>
<td><em>HpOCBS_A74S_Rp</em></td>
<td>5'-CGATGCCGGTGATTTGCCGCTTCAATGATG-3'</td>
</tr>
<tr>
<td><em>HpOCBS_F147Y_Fp</em></td>
<td>5'-AGCTATTTGCCCTTACAATATGAGCAGCAAG-3'</td>
</tr>
<tr>
<td><em>HpOCBS_F147Y_Rp</em></td>
<td>5'-GGGATTATCAGGTTTTCATATTGAAAGGCAGCAATGCT-3'</td>
</tr>
<tr>
<td><em>HpOCBS_V226Y_Fp</em></td>
<td>5'-AGATAGGGGATCTTGTAGCTATCCTCCTACTT-3'</td>
</tr>
<tr>
<td><em>HpOCBS_V226Y_Rp</em></td>
<td>5'-AAAGGAGGGGTAGAACTCAGCAGCCAATGCCCTCAATCT-3'</td>
</tr>
<tr>
<td><em>ScCBS_Fp</em>&lt;sup&gt;c&lt;/sup&gt;</td>
<td>5'-CTAGCTAGGATCGATGAGAATCTGGAGCAGACG-3'</td>
</tr>
<tr>
<td><em>ScCBS_Rp</em>&lt;sup&gt;c&lt;/sup&gt;</td>
<td>5'-GGGATGCTAGCAGCGTATGAGTGAAGGCTAAGAACG-3'</td>
</tr>
<tr>
<td><em>ScCBS_Y158F_Fp</em></td>
<td>5'-GTATATCTACTGACAATACAGCTAAGGGCAATACG-3'</td>
</tr>
<tr>
<td><em>ScCBS_Y158F_Rp</em></td>
<td>5'-GGGGATGCTAGCAGCGTATGAGTGAAGGCTAAGAACG-3'</td>
</tr>
<tr>
<td><em>ScCBS_Y248V_Fp</em></td>
<td>5'-AGTGGAGGGGTAGAACTCAGCAGCCAATGCCCTCAATCT-3'</td>
</tr>
<tr>
<td><em>ScCBS_Y248V_Rp</em></td>
<td>5'-GGGGATGCTAGCAGCGTATGAGTGAAGGCTAAGAACG-3'</td>
</tr>
</tbody>
</table>

<sup>c</sup>underlined sequences represent the restriction sites, while Fp and Rp stand for forward and reverse primers, respectively.
FIGURE LEGENDS

Figure 1. Reverse transsulfuration pathway and de novo pathway in Helicobacter pylori. In the reverse transsulfuration pathway, L-homocysteine reacts with L-serine in the presence of CBS to produce the intermediate L-cystathionine, which is further converted to L-cysteine by the action of the enzyme cystathionine γ-lyase (CGL) (Scriver & Kaufman, 2001, Stipanuk, 2004) (shown with green arrow). H. pylori can produce L-cysteine utilizing L-methionine via the reverse transsulfuration pathway. The organism includes OCBS (HP0107) and can produce L-cystathionine from L-homocysteine and OAS (an intermediate in the de novo pathway) (shown with red arrow). HP0106 is annotated as cystathionine γ-synthase but predicted to function as CGL in H. pylori (Doherty et al., 2010). H. pylori also has de novo pathway enzymes, namely serine O-acetyltransferase (SAT) and O-acetylserine sulfhydrylase OASS (OCBS in H. pylori), and can produce L-cysteine. In H. pylori, OASS was shown in the current work to function as an OCBS.

Figure 2. Phylogenetic study of OASS, OCBS and CBS of the PLP-II family. A phylogenetic analysis was carried out with 154 sequences using the maximum likelihood method with 250 bootstraps of the Mega 5.1 software. This analysis yielded a phylogenetic tree made up of three different clads, namely OCBS (red), OASS (green) and CBS (blue), with the OCBS clad containing well-annotated OCBSs from various bacteria, including HpOCBS (PDB ID: 6AHI). Red stars indicate experimentally verified OCBS proteins while blue stars indicate sequences predicted, but not yet shown experimentally, to be OCBS in whole genome sequencing of the corresponding bacterial genome.

Figure 3. WebLogo generated from multiple sequence alignment of OCBSs (55 sequences), CBSs (52 sequences) and OASSs (47 sequences). Residue numbering is according to HpOCBS for the OCBS alignment, H. sapiens CBS for the CBS alignment and OASS of M. tuberculosis for the OASS alignment. Consensus sequences near the active site were divided into 5 conserved blocks and the residues in CBS and OASS differing from those in OCBS are shown with red stars. Height of an individual amino acid in Weblogo indicates level of conservation at the particular portion (Crooks et al., 2004).

Figure 4. Biochemical characterisation of HpOCBS. (a) The OAS-dependent CBS activity of HpOCBS was characterised with 5 mM L-homocysteine and various concentrations of OAS in 50 mM of HEPES [pH 7.5] to determine the K_m and V_max values of the reaction. HpOCBS also catalyzes the production of H_2S from the reaction of L-cysteine with L-homocysteine. (b) Continuous monitoring of hydrogen sulfide production in the presence of 0.4 mM lead acetate upon reaction of L-cysteine (1 mM) with increasing concentrations of L-homocysteine. (c) HpOCBS was also found to display OASS activity, which was measured with various concentrations of OAS in 50 mM HEPES [pH 7.5] in the presence of 100 μM of TNB. Each result shown is the mean ± SD of three (n=3) independent experiments.
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This article is protected by copyright. All rights reserved.
1) \(O\)-acetylserine-dependent cystationine \(\beta\)-synthase reaction

\[
\text{\(O\)-acetylserine} + \text{L-Homocysteine} \rightarrow \text{L-Cystathionine} + \text{L-Cystathionine}
\]

2) Reaction producing hydrogen sulfide

\[
\text{L-Cysteine} + \text{L-Homocysteine} \rightarrow \text{Hydrogen sulfide} + \text{L-Cystathionine}
\]

3) \(O\)-acetylserine sulphydrylase reaction

\[
\text{\(O\)-acetylserine} + \text{Hydrogen sulfide} \rightarrow \text{L-Cysteine} + \text{L-Cystathionine}
\]