Burkholderia pseudomallei D-alanine-D-alanine ligase
DíazSáez, Laura; Torrie, Leah S.; McElroy, Stuart P.; Gray, David; Hunter, William M.

Published in: FEBS Journal

DOI: 10.1111/febs.14976

Publication date: 2019

Document Version
Publisher's PDF, also known as Version of record

Link to publication in Discovery Research Portal

Citation for published version (APA):

General rights
Copyright and moral rights for the publications made accessible in Discovery Research Portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from Discovery Research Portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain.
- You may freely distribute the URL identifying the publication in the public portal.

Take down policy
If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.
*Burkholderia pseudomallei* d-alanine-d-alanine ligase: detailed characterisation and assessment of a potential antibiotic drug target

Laura Díaz-Sáez¹, Leah S. Torrie², Stuart P. McElroy³†, David Gray² and William N. Hunter¹

¹ Division of Biological Chemistry and Drug Discovery, School of Life Sciences, University of Dundee, UK
² Drug Discovery Unit, Wellcome Centre for Anti-Infectives Research, School of Life Sciences, University of Dundee, UK
³ European Screening Centre Newhouse, Biocity Scotland, University of Dundee, Newhouse, UK

**Keywords**
antibacterial; enzyme assay; enzyme inhibition; peptidoglycan; target validation

**Correspondence**
W. N. Hunter, Division of Biological Chemistry and Drug Discovery, School of Life Sciences, University of Dundee, Dow Street, Dundee DD1 5EH, UK
Tel: +44 (0)1382385745
E-mail: w.n.hunter@dundee.ac.uk

†Present address BioAscent Discovery Ltd, Bo’ness Road, Newhouse, Lanarkshire, ML1 5UH, UK

(Received 18 February 2019, revised 27 May 2019, accepted 27 June 2019)
doi:10.1111/febs.14976

*Burkholderia pseudomallei* is a serious, difficult to treat Gram-negative pathogen and an increase in the occurrence of drug-resistant strains has been detected. We have directed efforts to identify and to evaluate potential drug targets relevant to treatment of infection by *B. pseudomallei*. We have selected and characterised the essential enzyme d-alanine-d-alanine ligase (*Bp*Ddl), required for the ATP-assisted biosynthesis of a peptidoglycan precursor. A recombinant supply of protein supported high-resolution crystallographic and biophysical studies with ligands (AMP and AMP+D-Ala-D-Ala), and comparisons with orthologues enzymes suggest a ligand-induced conformational change occurring that might be relevant to the catalytic cycle. The detailed biochemical characterisation of the enzyme, development and optimisation of ligand binding assays supported the search for novel inhibitors by screening of selected compound libraries. In a similar manner to that observed previously in other studies, we note a paucity of hits that are worth follow-up and then in combination with a computational analysis of the active site, we conclude that this ligase represents a difficult target for drug discovery. Nevertheless, our reagents, protocols and data can underpin future efforts exploiting more diverse chemical libraries and structure-based approaches.

**Introduction**

*Burkholderia pseudomallei*, a serious pathogen of humans and animals, causes melioidosis. This Gram-negative bacterium is widely distributed in the environment and is classified as a Category B bioweapon agent by the U.S. Centers for Disease Control and Prevention (CDC, www.cdc.gov). Treatment of melioidosis involves intravenous high-intensity application of cephalosporins for up to 2 weeks then an eradication phase to prevent recurrence that can involve the use of antifolate combinations for up to 20 weeks. This is an expensive treatment often complicated by side effects and a further issue is that strains resistant to the current drugs have been identified [1,2]. New antibacterial strategies are being considered [3] and to this end, we sought to investigate the potential of key enzymes as new therapeutic targets and to generate the data, reagents and protocols that support an assessment of these targets for early stage drug discovery.

The peptidoglycan (PG) layer contributes an important protective shield for Gram-negative organisms and antibiotics, such as *β*-lactams and glycopeptides, indeed the key drug for melioidosis, ceftazidime, target one or more enzymes required for its biosynthesis.

**Abbreviations**

BLI, biolayer interferometry; BSA, buried surface area; DCS, d-cycloserine; Ddl, d-alanine-d-alanine ligase; HTP, high throughput; PEP, phosphoenolpyruvate; PG, peptidoglycan; PK/LDH, phosphoglycerate kinase/lactate dehydrogenase.
Due to the success of targeting parts of the PG biosynthetic pathway [4–7], we focused on the assessment of D-alanine-D-alanine (D-Ala-D-Ala) ligase as a potential target for the development of new drugs for *B. pseudomallei* infections.

D-Alanine (D-Ala) contributes a key structural role in the PG layer as a dipeptide, and in lantibiotics [8–10]. The biosynthesis of D-Ala-D-Ala involves an ATP-dependent ligase (Ddl, EC 6.3.2.4) [10,11]. In some bacteria, for example *Escherichia coli*, there exist two proteins (EcDdlA and EcDdlB) [12] with a sequence identity of about 30%. At the time of writing, there are 587 reviewed Ddl sequences in UniProt (www.uniprot.org), and 18 bacteria presenting two Ddl proteins with sequence identities from 26% to 58%. A search for DdlA and DdlB sequences, performed using NCBI BLAST [13], identified only DdlB in *B. pseudomallei* (*Bp*Ddl).

Structures of DdlB from several bacteria have been determined in the apo-form and ligand-bound complexes [14,15]. Ddl presents the ATP-grasp family fold [16], and the catalysed reaction follows a sequential ordered mechanism [17,18]. Known inhibitors of Ddl include d-cycloserine (DCS) [17–19], phosphinates [15,20,21], diazenocarboxamides [22], hydroxyethylamine [23] and boron derivatives of D-Ala [24]. However, there has been little progress in developing highly potent inhibitors of Ddl with efficient engagement of the cellular target. This might be due to the polar nature of these inhibitors that would require active transport for uptake.

Our objective was to characterise and assess *Bp*Ddl as a potential antibacterial drug target and as part of that assessment to initiate a search for new inhibitor scaffolds. We report an efficient recombinant protein production system, purification and crystallisation protocols, two high-resolution ligand-bound *Bp*Ddl structures (AMP and AMP+D-Ala-D-Ala) together with comparisons with orthologues, the optimisation of two enzymatic assays for the functional characterisation of the enzyme and development of high throughput (HTP) assays that were applied to selected compound libraries. We used biolayer interferometry (BLI) and enzymatic assays to assess the outcomes of the HTP library screens and a computational assessment of the *Bp*Ddl active site properties.

**Results and Discussion**

**Overall subunit structure**

Two high-resolution crystal structures have been determined, *Bp*Ddl:AMP and *Bp*Ddl:AMP+D-Ala-D-Ala, with statistics in Table 1. Ddl is a member of the ATP-grasp superfamily presents a characteristic and highly conserved fold [16,25–28]. ATP-grasp proteins contain three well-defined domains [16]. In the case of Ddl, these are the N-terminal, central and C-terminal domains (Fig. 1). *Bp*Ddl secondary structure comprises 10 α-helices, one 3_10-helix, one π-helix and 12 β-strands (Fig. 1A). The N-terminal segment presents an α/β-domain whilst the central and C-terminal domains comprise the ATP-grasp fold; two α/β-domains that generate a narrow pocket for the ATP binding. The D-Ala binding part of the active site is located in the C-terminal domain. Two loops are implicated in the binding of the co-factor and the substrate. The P-loop (residues 149–159) is located in the central domain, between β-strands 5 and 6, and interacts with the γ-β phosphates of ATP. The ω-loop is located in the C-terminal domain and presents a π-helix configuration involving residues 198–222 (Fig. 1A). At several positions, gross structural differences are noted when comparing subunits of the two *Bp*Ddl structures (Fig. 1B). The position of the central domains and several helices on the N-terminal and C-terminal domains, that line the active site, adopt different positions. The Cα comparison illustrates the notable conservation of these structural changes in both *Bp*Ddl models (Fig. 1C).

**Quaternary structure**

In solution, at low concentrations, *Bp*Ddl behaves as a monomer as indicated by SEC. Native-PAGE on a more concentrated sample indicated monomer as well as the presence of a higher molecular mass species, possibly a dimer (Fig. 2). The crystal structures present a dimer in the asymmetric unit with each subunit aligned perpendicularly to the other (Fig. 1A). The *Bp*Ddl dimer is formed by about 12–13% of the accessible surface area (ASA) of a subunit, which involves residues from α3, α4, α5 and the C-terminal region of α8. Comparisons with structures from different organisms (Table 2) indicate that the dimer arrangement is shared (data not shown), and the ASA percentage involved in the dimerisation, also called buried surface area (BSA), is similar. These BSA values are lower than expected for a surface with a high probability of forming a multimeric assembly [29,30]. In contrast, the P-value for the probability of having a similar observed solvation free energy gain (ΔG) in other surface areas is lower than 0.5, indicating a higher level of hydrophobicity than expected and it is likely this property that drives oligomerisation [29]. *Mycobacterium tuberculosis* Ddl (M1Ddl) [27] has a reduced contact area, with no interactions involving residues...
Table 1. Crystallographic statistics

<table>
<thead>
<tr>
<th>Structure</th>
<th>BpDdl:AMP</th>
<th>BpDdl:AMP::D-Ala::D-Ala</th>
</tr>
</thead>
<tbody>
<tr>
<td>PDB code</td>
<td>5NRH</td>
<td>5NRH</td>
</tr>
<tr>
<td>Space group</td>
<td>P2₁</td>
<td>P2₁</td>
</tr>
<tr>
<td>Wavelength (Å)</td>
<td>0.9174</td>
<td>0.9174</td>
</tr>
<tr>
<td>Unit cell dimensions</td>
<td>69.65, 61.15, 69.67</td>
<td>69.64, 61.13, 69.67</td>
</tr>
<tr>
<td>a, b, c (Å), β (°)</td>
<td>90.17</td>
<td>90.31</td>
</tr>
<tr>
<td>Resolution range (Å)</td>
<td>70.08 – 1.30</td>
<td>69.97 – 1.50</td>
</tr>
<tr>
<td>No. reflections</td>
<td>672,251</td>
<td>283,077</td>
</tr>
<tr>
<td>Unique reflections</td>
<td>141,049</td>
<td>90,609</td>
</tr>
<tr>
<td>Completeness (%)</td>
<td>97.6 (95.7)</td>
<td>96.6 (98.3)</td>
</tr>
<tr>
<td>Completeness (%)</td>
<td>97.6 (95.7)</td>
<td>96.6 (98.3)</td>
</tr>
<tr>
<td>Rmerge (%)</td>
<td>0.04 (0.66)</td>
<td>0.05 (0.45)</td>
</tr>
<tr>
<td>CC1/2 (%)</td>
<td>0.99 (0.74)</td>
<td>0.99 (0.77)</td>
</tr>
<tr>
<td>Redundancy</td>
<td>4.8</td>
<td>3.1</td>
</tr>
<tr>
<td>Bond lengths (Å)/angles (°)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Average B-factors [Å²]</td>
<td>23.9</td>
<td>25.2</td>
</tr>
<tr>
<td>AMP average B-factors [Å²]</td>
<td>13.3</td>
<td>13.7</td>
</tr>
<tr>
<td>α-Ala–β-Ala average B-factors [Å²]</td>
<td></td>
<td>32.4</td>
</tr>
<tr>
<td>Protein residues</td>
<td>1412</td>
<td>1345</td>
</tr>
<tr>
<td>Water molecules</td>
<td>785</td>
<td>709</td>
</tr>
<tr>
<td>Metal ions</td>
<td>1 (Mg²⁺)</td>
<td>1 (Mg²⁺)</td>
</tr>
<tr>
<td>Ligands</td>
<td>7 (2 AMP, 2 SO₂⁻, 3 ethylene glycol)</td>
<td>15 (2 AMP, 1 α-Ala, 3 SO₂⁻, 8 ethylene glycol, 1 poly-ethylene glycol)</td>
</tr>
</tbody>
</table>

Ramachandran analyses

| Favourable regions (%) | 98 | 98 |
| Allowed regions (%) | 99.8 | 99.8 |

*Values in parentheses refer to the highest resolution shell.

1Rmerge = ∑hkl(Ihkl) – <I(hkl)>/∑hkl|Ihkl|, where Ihkl is the intensity of the ihth measurement of reflection hkl and <I(hkl)> is the mean value of Ihkl for all i measurements.

2Rfree = ∑hkl(Fo – |Fo|)/∑hkl|Fo|, where Fo is the observed structure factor and Fc is the calculated structure factor.

3Rmerge = ∑hkl(Fo – |Fo|)/∑hkl|Fo|, where Rfree is the same as Rwork except calculated with a subset, 5%, of data that are excluded from the refinement calculations.

4Diffraction Precision Index. 5Protein atoms.

Also, the P-value is higher for MtDdl than the other structures. Ddl structures can be grouped according to the additional number of amino acids, compared to BpDdl, at the C-terminal end of α10. When extra residues are present, α10 is elongated towards the partner subunit and an increased number of inter-subunit contacts assist dimer formation. This is noted for Bacillus anthracis (BaDdl), Streptococcus mutans (SmDdl) [31], Staphylococcus aureus (SaDdl) [32], Coxiella burnetii (CbDdl) [27] and Xanthomonas oryzae (XoDdl) [12], where between 8 and 20 additional residues are observed. Table 2 also presents the sequence identity shared with proteins of known structure, which ranges from about 30% to 90%. Despite a wide range of overall sequence identity compared to BpDdl we note that residues contributing to...
dimer formation are well conserved, with Gly77, Gly81, Val95, Ala99 and Asp103 present in 12–13 structures of the 14 analysed and three of these residues are also located near the active site binding pockets (Gly77, Val95 and Asp103). Despite differences elsewhere in the subunit–subunit interface, the dimer arrangement is a recurrent feature in Ddl.

Co-factor and substrate-binding pockets

Co-factor and substrate-binding sites are identified in Figs 1 and 3, the latter also showing electron density associated with the ligands. Whilst the complete AMP is well-orderd the N, Cα, Cβ atoms of d-Ala1 are not. The residues that contribute to the formation of these binding sites are highly conserved in Ddl sequences. Indeed, the ATP-binding site residues (Lys104, Gly153, Glu184, Asp262, Glu275 and Asn277) are also conserved within different functional proteins from the ATP-grasp superfamily such as acetyl-CoA carboxylase [16]. The recognition of ATP involves residues from the central and C-terminal domains. These include Lys104, Lys148 and Glu275 binding to the phosphate, and Glu184, Lys185, Ile187, Glu192, Phe214 and Tyr215 that are involved in adenosine recognition. In the case of L-Ala–L-Ala, the key interacting residues are Tyr215, Tyr221, Asp277, Arg260, Gly281, Ser286, Leu287 and also a Mg2+–bound water molecule.

In the substrate-binding pocket, Lys220, Arg260, Asp277 and Gly281 are strictly conserved and Leu287 in 90% of sequences. These are key residues for substrate binding and catalysis according to the proposed reaction mechanism [17,18]. Conservation is also noted for Phe214, Tyr215 and Tyr221 (40–50% identity) with other aromatic residues. These residues appear to be involved in placing ATP and d-Ala in the correct position for catalysis. In the ATP-binding pocket, Lys104, Lys148, Glu184, Glu192 and Glu275 are conserved in between 98% and 100% of the sequences highlighting their importance to co-factor binding.

When BpDdl:AMP:d-Ala–d-Ala was compared with other product and substrate-bound structures (from EcDdl: PDB codes: 1IOV, 4CS5, 4CSC; from TrDdl: 2ZDH and 2ZDQ, including ADP+phosphate, ADP+d-Ala, ATP+d-Ala–d-Ala, ADP+d-Ala, ATP+d-Ala respectively), differences in the substrate and product binding were found (Fig. 4). The Cα overlay of BpDdl:AMP:d-Ala–d-Ala (subunit A) with these ligand-bound structures showed the location of d-Ala–d-Ala, though similar, participates in different interactions compared to previously characterised proteins. Additionally, the relative position of the Mg2+ is different. In other Ddl structures (e.g. EcDdl, PDB code 4CSC), two Mg2+ ions interact with Asp262, Glu275 and γ-β phosphates from ATP. However in BpDdl: AMP:d-Ala–d-Ala only one Mg2+ is observed, interacting with His70. The overall BpDdl:AMP:d-Ala–d-Ala conformation presents a more open form than in other structures, suggesting it is an intermediate conformation, but it cannot be distinguished if this corresponds to a substrate binding or product release phase.

Conformational changes

Ligand-induced conformational changes can be observed when comparing the AMP loaded structure with an apo-form extracted from the PDB (Fig. 5). Binding of AMP generates a more compressed conformation where the central domain moves towards the centre of the structure and the α-loop rotates and generates the active site pocket. ATP binding therefore is likely to stabilise the α-loop conformation enforcing interactions with Tyr215 to create a closed conformation. Phe214 is able to interact with a hydrophobic area of the central

Fig. 2. SEC and Native-PAGE 3-12% results. (A) SEC chromatogram for BpDdl. The plot shows the equilibration curve (blue) and BpDdl peak (orange dot). The expected value for monomers is marked with an arrow and dimer with a dashed arrow. (B) Native-PAGE. Lane 1: Molecular weight standards. Lane 2: 10 ng of BpDdl. The arrow indicates the monomer and * the expected value.
domain, and Glu213 with Lys160 to assist closing the ATP-binding pocket. The conformational change in the central domain involves a movement/shift of $\alpha_7$ of about 12.5 and 7.6 Å for $\alpha_6$ (average distances from both subunits) favoured by the AMP and $\omega$-loop interactions. Additionally, the conformation of the P-loop is stabilised by AMP interactions, the $\omega$-loop and the N-terminal domain residue.
is disordered and not included in the model but in the AMP-bound structures this loop is ordered with an average $B$-factor of 36.7 Å² (the overall average $B$-factor value is 23.9 Å²). This higher $B$-value suggests a greater degree of flexibility in the loop.

There are apo and ligand-bound structures for $SaDdl$ (2I8C, 2I80, apo: 2I87) [32] and $TtDdl$ (2YZM, 2YZN, 2ZDH, 2ZDG, 2ZDQ, apo: 2YZG). In both cases, the conformational changes noted when comparing different states do not appear as pronounced as in $BpDdl$. Movement of the $TtDdl$ central domains involves an $a_7$ translation of 4.7 Å, whereas no differences in $SaDdl$ are observed. This tends to suggest that the conformational changes are distinct in different organisms. As indicated in ESC library screening in Results and Discussion, purified $BpDdl$ contained AMP and we were unable to obtain an apo-$BpDdl$ crystal form. With this in mind, electron density maps of the apo-$BpDdl$, $SaDdl$ and $TtDdl$ structures were inspected to check for missed co-factors but in all three there was no evidence of anything bound in the ATP pockets.

A comparison of the electrostatic surfaces in and around the active sites of apo-$BpDdl$ and $BpDdl$:AMP reveals significant differences (Fig. 6). The $\delta$-Ala pocket is more electropositive in the AMP-bound structure. Therefore, at pH 7.5, $\delta$-Ala ($pI = 6.01$) would preferably bind to the $BpDdl$:AMP conformation over the apo-form. This suggests that the $\delta$-Ala pocket is formed after the $\omega$-loop conformational change. The residues involved in determining the electrostatic properties of the different conformations (Glu22, Asp262, Glu275 and Asn277 in apo-$BpDdl$, and His70, Arg260 and Asn277 in $BpDdl$:AMP) are highly conserved. Also, as described previously, His70, Arg260 and Asn277 are key amino acids for Ddl catalysis. This is consistent with the hypothesis that the generation of a favourable environment for catalysis is dependent on ATP binding, in agreement with the sequential ordered mechanism previously characterised [17,18] where ATP is the first component to bind Ddl.

**Biochemical characterisation**

**Enzyme kinetics**

Enzyme activity was investigated using a coupled spectrophotometric assay and a BIOMOL Green assay at
pH 7.5 and 8 (Table 3). Using the coupled assay, inhibition of the enzyme activity was observed at high concentrations of ATP and $K_{i,ATP}$ was also determined. In the BIOMOL Green assay, high ATP concentrations decrease the signal-to-noise ratio due to nonenzymatic hydrolysis and consequently, ATP concentrations did not exceed 1 mM. Nevertheless, the $K_{i,ATP}$ could not be accurately determined with the BIOMOL assay. The values for $K_m2$ were consistent between the coupled and the BIOMOL assays. Comparing the $K_m$ values with other Ddl enzymes, we note more similarity between $Bp$ Ddl and $Mt$ Ddl or $Ec$ Ddl than $Y$ pestis Ddl or $Hp$ Ddl (Table 1). Indeed, $Hp$ Ddl seems to be the least related Ddl, not only displaying a lower sequence identity but also different kinetic parameters.

We showed that our $Bp$ Ddl samples had AMP-bound, so the $K_{ATP}$ determination is compromised. No information about the presence of AMP or similar compounds in $Hp$ Ddl, $Ec$ Ddl and $Mt$ Ddl has been reported. Purification of $Mt$ Ddl [17] and $Hp$ Ddl [26] was carried out using nickel-affinity columns whereas $Ec$ Ddl used ion exchange [20,33]. It is possible that $Mt$ Ddl and $Hp$ Ddl also co-purified with the co-factor as similar protocols to those we used for $Bp$ Ddl were employed; however direct confirmation would have to be sought.

The pH optimum for $Bp$ Ddl was determined. Differences in the activity due to the change in the pH are observed in the $K_{m2}$ and the $V_{max}$ determinations. At a lower pH there is an increase of 1.2 times in the $K_{m2}$ and 1.7 times decrease in the $V_{max}$. To further explore this, activity was measured over a pH range of 5.1–9.5. Regarding this, a test at different pH values has been carried out (Fig. 7). The peak of the maximum activity is observed at pH 8 (100% relative activity) and the activity decreases around 1.7 times at pH 7.5 (56% relative activity). This is consistent with the results from the $K_m$ experiments.

Determination of the $K_i$ and the mechanism of inhibition for DCS were assessed (Fig. 8). DCS appears to be a competitive inhibitor of substrate with a $K_{i,DCS}$ value of 100 lM. At the highest DCS concentration tested, 1 mM, a different $V_{max}$ was observed what may

### Table 3. Kinetic parameters for Ddl. $K_m$ values for two α-Ala and ATP, the $K_i$ for ATP and the $V_{max}$ of the reaction

<table>
<thead>
<tr>
<th>Detection method</th>
<th>pH</th>
<th>$K_m1$ (mM)</th>
<th>$K_m2$ (mM)</th>
<th>$K_{ATP}$ (mM)</th>
<th>$K_{i,ATP}$ (mM)</th>
<th>$V_{max}$ (nmol NAD$^+$/min$^{-1}$)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Coupled</td>
<td>8</td>
<td>0.37 ± 0.085</td>
<td>4.19 ± 0.71</td>
<td>0.07 ± 0.01</td>
<td>3.16 ± 0.59</td>
<td>21.85 ± 0.98</td>
</tr>
<tr>
<td>Coupled</td>
<td>7.5</td>
<td>0.35 ± 0.03</td>
<td>5.12 ± 1.09</td>
<td>–</td>
<td>–</td>
<td>12.92 ± 1.38</td>
</tr>
<tr>
<td>Biomol</td>
<td>8</td>
<td>–</td>
<td>4.9</td>
<td>0.13</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>Biomol$^{TCEP/Tween}$</td>
<td>8$^a$</td>
<td>–</td>
<td>3.2</td>
<td>0.2</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>Biomol$^{200 \mu M \text{ATP}}$</td>
<td>8$^b$</td>
<td>–</td>
<td>2.3</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>Biomol$^{TCEP/Tween}$</td>
<td>7.5$^c$</td>
<td>–</td>
<td>5.4</td>
<td>0.27</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>$Hp$ Ddl [26]</td>
<td>8</td>
<td>1.89</td>
<td>627$^c$</td>
<td>0.000087</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>$Ec$ Ddl [21]</td>
<td>7.8</td>
<td>0.0012</td>
<td>1.13</td>
<td>0.049</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>$Mt$ Ddl [18]</td>
<td>8</td>
<td>0.075 ± 0.01</td>
<td>3.6 ± 0.6</td>
<td>0.31 ± 0.05</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>$Y$ pestis Ddl</td>
<td>7.8</td>
<td>0.006</td>
<td>0.07</td>
<td>3.2</td>
<td>–</td>
<td>–</td>
</tr>
</tbody>
</table>

$^a$In the presence of 1 mM TCEP and 0.01% (w/v) Tween-20.

$^b$In the presence of 200 μM ATP instead of 500 μM ATP.

$^c$Maximum α-Ala concentration tested. (—) Not determined.
indicate that the dynamics of binding changes. DCS could be binding both d-Ala pockets and so display different binding behaviour. For *Mt* Ddl (*K*_i, *DCS* of 25 μM) and *Ec* Ddl (*K*_i,CDS of 185 μM), it was reported that slow binding of DCS occurs [19] but not at a high concentration (1000 μM). This DCS concentration dependent behaviour appears to also apply to *Bp* Ddl.

**Quantification of AMP**

Characterisation of structures derived from crystals grown in the presence of different ligands, or none, revealed all had AMP-bound. This, we presume is derived from the expression host and we decided to quantify the level of AMP in the purified sample. Denatured and folded samples were compared.

Firstly, controls *BaKynB*, *BaKynB* in the presence of 2 μM AMP and *BpDdl* itself were denaturated using (a) pH extremes, (b) an elevated temperature and (c) chymotrypsin digestion. The experimental and expected results are shown in Fig. 9. The negative control (in the absence of protein and AMP) and positive control (10 μM AMP in the absence of protein) results were similar to expected values. *BaKynB* likewise, confirmed there is no AMP-bound [34]. The *BaKynB* data indicate that the different sample treatments are not interfering with the AMP-glo assay. The results for *BpDdl* demonstrate the sample has been co-purified with AMP. The presence of AMP without pretreatment of *BpDdl* indicates there is some free material in the protein solution (around 0.4 μM) probably due to AMP exchange/release. The sample treated with chymotrypsin presents an AMP concentration of about 2 μM that indicates 1.6 μM (approximately 15 nmol) AMP remains bound to *BpDdl*. Taking into account that the concentration of *BpDdl* is 25 mg·mL⁻¹ (about 25 mg·mL⁻¹) AMP, the calculated AMP concentration is approximately 0.06 mg·mL⁻¹ (about 60 nmol).

**Fig. 7.** Effect of pH on enzyme activity. The relative activity is set to 100% at maximum level, at pH 8.

**Fig. 8.** DCS *K* determination. (A) Double reciprocal plot for the DCS inhibition and the corresponding equation for competitive inhibition. (B) Secondary representation for inhibitory data. Plot of the slopes from the reciprocal plot against the concentration of DCS. From the equation of the slope of this line the *K*_i,DCS can be determined as shown in the accompanying equation. *V*, reaction velocity (nmol·min⁻¹); *K*_m, Michaelis–Menten constant (M); *V*_max, maximum velocity of the reaction (nmol·min⁻¹); [S], substrate concentration (M); [I], inhibitor concentration; *K*_i, inhibitory constant.
20 nmol), then the ratio AMP:BpDdl suggests almost full occupancy of BpDdl by the monophosphate. This is consistent with the structural data that show one, well-ordered AMP per BpDdl subunit.

The presence of AMP in the purified enzyme, presumably acquired from the bacterial expression system has implications for our analyses. The $K_{ATP}$ determination and the compound screens therefore have been done mainly with the binary complex BpDdl:AMP instead of apo-BpDdl. Additionally, during the spectrophotometric assay, a retarded phase was observed at the onset of the reaction (Fig. 10). This could be due to diffusion of AMP that has to first occur prior to ATP loading. Also, the fragment screen using BLI was carried out with the closed BpDdl conformation and that might explain why a relatively low ATP-binding signal was measured.

An apo-BpDdl structure (PDB code 4EGQ) has been determined by the Seattle Structural Genomics Center for Infectious Disease. Electron and difference density maps in the ATP-binding sites were inspected to ascertain if a ligand might be present. There were no significant features so we conclude this is a bone fide apo-BpDdl form. It is unclear why in our structures AMP is present and this may be due to different purification or crystallisation protocols. The PDB entry stipulates that the apo-form crystal was grown at a basic pH (0.1 M CHES pH 9.5, 1 M Na$^+$-K$^+$-tartrate and 0.2 M Li$_2$SO$_4$) whereas the crystals obtained in our work were obtained at pH 5.5. There are no details of the apo-form purification. Whilst it is not uncommon for a purified protein to acquire ligands during recombinant protein purification as exemplified by human PPAR-β/δ binding endogenous fatty acids of E. coli [35], it is intriguing in this case. We speculate that this might be indicative of a mechanism whereby the levels of AMP:ATP could be networked to the control exercised by the second messenger cyclic di-AMP to regulate peptidoglycan biosynthesis [36]. The role of AMP and derivatives is coming under increased scrutiny in this aspect [37].

**Compound library screens**

We screened three compound libraries. BLI was used in conjunction with a fragment library whilst the malachite green-based assay was applied to the ESC and small diversity set libraries.

**BLI: Fragment library screen**

BpDdl was interrogated with a fragment library of around 700 compounds, using a BLI binding assay. A total of 11 hits were selected for a secondary inhibition assay at 300 μM. A single compound (Fig. 11) gave enzyme inhibition with a $K_i$ value of 10 μM. Attempts to obtain a structure of the hit in complex with the enzyme by co-crystallisation failed.
ESC library screening

The optimised HTP assay had a Z’ score of 0.8 and a signal-to-noise ratio of 5 after 80 min of reaction. Initially, the experiments were run in the absence of TCEP and Tween-20. To target the substrate and cofactor binding sites, a concentration of n-Ala and ATP close to the K_m values was used. A library of 6775 compounds, comprising well-known drugs, and, for control purposes, known aggregators, was tested. A total of 74 hits were selected and re-tested using the BIOMOL Green assay. Results showed no interference by any of the compounds but that 61 appeared to be false positives leaving 13 compounds for further investigation. The dose–response experiment for the hits was performed in the presence of 1 mM TCEP and 0.01% (w/v) Tween-20 and no inhibition was observed. Thus the redox environment may affect the stability of the compounds and/or induce aggregation. Indeed, inspection of the chemical structures suggested they could also be aggregators [38] and this was further supported by the dose–response profile from one of the compounds (SAM001247071, Fig. 12), which is characteristic of this issue [39]. Although our assay appears to be reliable no useful hits were obtained.

DDU small diversity set screening

Due to the high similarity of the ATP-binding pockets between different functional proteins (glutathione synthetase [40], ATP-citrate lyase [41], succinyl-CoA ligase [42] and kinases [43]), targeting this pocket might result in inhibition of human enzymes with toxic side effects. To avoid this, the inhibitor search was focused on obtaining new inhibitors targeting the n-Ala binding pocket. The DDU diversity set screen was selected to inform on this part of the active site. A new optimisation of the HTP assay was carried out to establish screening conditions to target this pocket. The reaction was linear for 20 min, the signal-to-noise ratio 6 and the Z’ score 0.8–0.9. From the 15 667 compounds tested, four hits were selected on the basis of a 30% inhibitory effect. However, the follow-up dose–response experiment identified them as false positives.

Experimental procedures

Recombinant protein production

An oligonucleotide encoding BpDdl (Uniprot code: Q63Q9J, protein mass 33 kDa) was purchased from GeneWiz (Takeley, Essex, UK) and inserted into a modified pET15b vector (Novagen, Madison, WI, USA), that adds an N-terminal hexahistidine tag followed by a tobacco etch virus (TEV) protease cleavage site before the resulting target protein. The restriction enzymes used were Ndel (at the 3’ DNA end) and BamHI (5’ end). The integrity of the construct was confirmed by sequencing. Recombinant protein production was achieved with BL21 (DE3) cells, 1 mM IPTG (final concentration) for induction and incubating overnight (O/N) at 20 °C. Purification involved use of metal ion affinity chromatography with a Ni²⁺-NTA column (GE Healthcare, Chicago, IL, USA) using buffer A (20 mM Tris-HCl pH 7.4, 200 mM NaCl, 5 mM MgCl₂) for sample loading and buffer B (20 mM Tris-HCl pH 7.4, 200 mM NaCl, 5 mM MgCl₂, 800 mM imidazole) for elution. The hexahistidine tag was cleaved, during dialysis back into buffer A, using 1 mg of TEV protease per 10 mg of BpDdl, incubated O/N at 4 °C. A second round of affinity chromatography with a Ni²⁺-NTA column removed the tag and the TEV protease, itself carrying a hexahistidine tag, and fractions carrying the purified protein were pooled. Protein purity and molecular weight were assessed by SDS/PAGE.

Protein concentration was determined using the theoretical molar extinction coefficient ε(BpDdl) = 27 515 M⁻¹cm⁻¹ as calculated in PROTPARAM [44]. Quaternary structure was investigated using native-PAGE (Novex, ThermoFisher Scientific, Waltham, MA, USA) and size-exclusion chromatography (SEC) using buffer A and the HiLoad 16/60 superdex 200 prep grade column (GE Healthcare).

Crystallisation

The protein sample was concentrated to 25 mg·mL⁻¹ in buffer A. Initially, thin needle shaped crystals grew in sitting drops plates at 18 °C after incubating the protein (at different protein : reservoir ratios) with a reservoir containing 0.2 m Li₂SO₄, 0.1 m Bis-Tris pH 5.5 and 25% (w/v) PEG 3350. These crystals were crushed to provide micro-seeds. Hanging drops were set up using a protein : reservoir ratio of 1 : 1 (in a final volume of 4 µL) plus 0.5 µL of crystal seeds. Crystals grew at 18 °C under various conditions: 0.05–0.3 m Li₂SO₄, 0.1 m Bis-Tris pH 5.5 and 15–30% (w/v) PEG 3350. Co-crystallisation with different ligand combinations was attempted using BpDdl incubated for 30 min on ice in the presence of different ligand
combinations (e.g. ATP, DCS, d-Ala+ADP, d-Ala-d-Ala+AMP, DCS+AMP, DCS+AMPPNP). Numerous crystals were analysed but only two structures are reported, a binary complex with AMP and a ternary complex with d-Ala-d-Ala+AMP. In all other cases only AMP was observed in the active site and this will be discussed further below.

**Diffraction, data collection and structural analyses**

Optimised crystals, grown in the presence of ATP, were tested in-house using a M007HF X-ray generator with a Saturn 944HG+ CCD detector (Rigaku-Americas, Woodlands, TX, USA) and subsequently used to record a 2.0 Å resolution data set on ESRF beamline BM30-A. The data were indexed and integrated using xds [45], then scaled and analysed using AIMLESS [46]. The Matthews coefficient (V_m of 2.27 Å³ Da⁻¹ with bulk solvent of about 50%) suggested two molecules in the asymmetric unit. Two subunits were positioned with PHASERMR [47] using only the protein components of the 2.2 Å resolution apo-BpDdl structure (PDB code 4EGQ, subunit A). Rigid body refinement followed by cycles of restrained refinement using REFMAC [48], electron and difference density map inspections, and model manipulations were made with COOT [49]. AMP, not ATP, was observed in the active site and the end result was a 2.0 Å resolution structure of this binary complex. Other crystals, of putative apo-enzyme and grown in the presence of different ligands provided 13 data sets on Diamond beamline I04-1. Data were processed using XDSgui and IMOLSLFLM [50], and scaled and merged with AIMLESS. All crystals were isomorphous with the initial structure. These data produced two structures. In one case, that of a crystal obtained in the presence of ADP and d-Ala, data extending to 1.3 Å resolution were recorded and molecular replacement was performed with the refined subunit A from the binary complex. This structure only showed AMP-bound in the active site and hence, due to a superior resolution, supersedes the first; this structure is BpDdl:AMP. A 1.5 Å resolution structure of a ternary complex of BpDdl:AMP: d-Ala-d-Ala was the second structure. This was refined starting from the BpDdl:AMP model. Tight, noncrystallographic symmetry (NCS) restraints were employed at the onset of the refinements and these were released once solvents and ligands were included. Side-chain rotamers, water molecules and ligands, for example, ethylene glycol, were added when the protein models were completed and then anisotropic thermal parameters were included in the model. In the case of the ternary complex, a difference Fourier map suggested the presence of d-Ala-d-Ala. A model with full occupancy for the complete ligand resulted in negative difference density at the N-terminus of the dipeptide. However, if only the second d-Ala was refined, positive difference density was observed. We conclude that the first d-Ala (d-Ala1) is not fully ordered and although included in the model we assigned zero occupancy. A Mg²⁺ ion is present near His70 (subunit A). Two strong difference density features, in BpDdl:AMP (near residues 293 from subunit A, 221 from subunit B) were interpreted as SO₄²⁻ ions. Three such examples were noted in BpDdl:AMP: d-Ala-d-Ala (near residues 293 from subunit A, 221 and 290 from subunit B). The sulphates near residue 221 occupy the d-Ala binding pocket. Refinement was concluded when no improvements resulted for R_free and R_work values, when density maps indicated no further changes were justified and that satisfactory model geometry was obtained. For the other data sets, omit maps were inspected after some refinement and AMP was noted in all structures, irrespective of the ligands that had been added or when an apo-structure was sought. This indicated that AMP might be present in the protein solution, a point we had to investigate (see below).

**Model analyses and informatics**

MOLPROBIT [51] was used to assess model geometry. Secondary structure was inspected using DSSP [30], the dimer interface was analysed using PISA [29,52] and MOLSOFT ICM (www.molsoft.com), and B-factors analysed using Baverage from the CCP4 suite. Figures were prepared using ALINE [53], PYMOL (www.pymol.org) and MOLSOFT ICM. The DALI server [54] was used for comparative purposes whilst electrostatic surfaces and properties of the enzyme active site were determined using MOLSOFT ICM.
A comparison of Ddl sequences was carried out. First, a search for relevant sequences in UniProt (www.uniprot.org) was conducted. The identified sequences were grouped by 50% identity and aligned using CLUSTAL OMEGA [55] with group representatives (based on 588 sequences).

Enzymatic assays

A coupled spectrophotometric assay was initially optimised for characterising the enzyme. To support the HTP screening of compound libraries, a malachite green-based assay was developed.

A coupled spectrophotometric assay

$K_m$ determination

The activity of BpDdl was measured using a spectrophotometric assay [17]. The assay buffer contained 100 mM Tris-HCl pH 8, 100 mM KCl, 10 mM MgCl$_2$, 2.5 mM phosphoenolpyruvate (PEP), 0.4 mM NADH, 5 μL·m$^{-1}$ phosphoglycerate kinase/lactate dehydrogenase (PK/LDH) (Merck & Co, Kenilworth, NJ, USA) and 30 μM BpDdl. Additionally, to determine $K_m$ for D-Ala and ATP, 500 μM ATP and variable concentrations of D-Ala (0.1, 0.3, 0.5, 0.7, 1, 1.5, 2, 2.5, 3, 5, 10, 20 and 100 mM) or 10 mM D-Ala and variable concentrations of ATP (0.1, 0.2, 0.5, 1.2 and 3 mM) were added respectively. Ten times (10×) concentrated D-Ala/ATP stocks were prepared for each assay so the volume added was constant. The final reaction volume was 100 μL, and the reaction started by adding the enzyme. All components were incubated for 30 min at 30°C and then the assay was performed in triplicate. Negative controls included the master mix with BpDdl and without substrate, and the master mix without BpDdl and with D-Ala/ATP. No decrease in absorbance was detected in the negative controls during the assay period. The positive control was the PK/LDH combination and instead of ATP, 0.4 mM ADP was added. Initial velocities were obtained by measuring the steady-state rate for 60 s and determination of the $K_m$ values ($K_{m1}$(D-Ala), $K_{m2}$(D-Ala), $K_m$(ATP)) was carried out as previously described [17]. Inhibition of the enzyme activity at high ATP concentrations was observed, and the $K_{i,ATP}$ calculation was carried out with GRAPHPAD (www.graphpad.com).

BpDdl inhibition

Inhibition experiments were performed in the presence of different inhibitor concentrations to determine $K_{i1}$(D-Ala). In the case of DCS the concentrations used were 100, 200, 250, 300, 500 and 1000 μM. Stocks (10×) for each inhibitor concentration were prepared. Each $K_i$ determination was conducted for about 12 h. To check for time-dependent loss of BpDdl activity, an assay without inhibitors was performed before and after the inhibition studies. $K_i$ calculations were performed using double reciprocal plots [17]. To check for inhibition of any of the coupled enzymes, a cross-validation assay was performed. The assay buffer in this case contained 100 mM Tris-HCl pH 8, 100 mM KCl, 10 mM MgCl$_2$, 2.5 mM PEP, 0.4 mM NADH, 5 μL·m$^{-1}$ PK/LDH mix and 0.5 mM ADP. Concentrations of 0.5 and 10 mM DCS (from stock 1 m) and a positive control (without inhibitor) were tested. No inhibition was detected. A potential inhibitor was identified (see below) and the $K_i$ determination was carried out as for DCS using final concentrations of 200, 500, 2000 and 5000 μM.

The pH profile of BpDdl activity

The reaction mixture contained 100 mM of the selected buffer, 10 mM KCl, 10 mM MgCl$_2$, 2.5 mM PEP, 0.4 mM NADH, 5 μL·m$^{-1}$ PK/LDH mix, 0.5 mM ATP, 10 mM D-Ala and 20 μM BpDdl (added to start the reaction). The buffers used are: Bis-tris pH 5.5, Tricine pH 5.1, MES pH 6.0, HEPES pH 7.5, Tris-HCl pH 7.5, Tris-HCl pH 8.0, Tris-HCl pH 8.5, HEPES pH 8.5, BICINE pH 9.0 and CHES pH 9.5. The maximum reaction rate was used to define 100% activity.

Malachite green-based assay

$K_m$ determination

The release of phosphate after ATP hydrolysis can be measured using malachite green (BIOMOL green, ENZO life sciences, Farmingdale, NY, USA). This set of experiments and a compound library screen (see ESC library screen using the malachite green-based assay in Experimental procedures) were carried out at the European Screening Centre (ESC, Newhouse, UK). The assays were run at room temperature in a final volume of 30 μL, in clear bottom 384 well plates (Greiner, Kremsmunster, Austria). The assay buffer to determine the $K_{m1}$(D-Ala) and $K_m$(ATP) contained 100 mM Tris-HCl pH 8, 10 mM KCl, 10 mM MgCl$_2$, 0.4 mM BpDdl, variable concentrations of D-Ala (serial 1/4 dilutions from top concentration of 10 mM) or ATP (serial 1/4 dilutions from top concentration of 0.5 mM) and 0.5 mM ATP or 10 mM D-Ala respectively. The time points used in these experiments were as follows: 10 s, 30 s, 1, 1.5, 2, 3, 7, 10, 15 and 20 min. To avoid nonspecific ATP hydrolysis due to the low pH of the BIOMOL green reagents, tri-sodium citrate was added to each reaction at a final concentration of 0.65% (w/v). Each experiment included four replicates. Absorbance was recorded at 620 nm using a PerkinElmer EnVision plate reader (PerkinElmer, Waltham, MA, USA). For the calculations of D-Ala $K_m$ only $K_{m2}$ was taken into account and data were analysed with the Michaelis–Menten equation. High ATP concentrations are incompatible with the BIOMOL Green assay hence enzyme inhibition by this
compound was not a factor and $K_{\text{ATP}}$ was determined using the Michaelis–Menten equation. Regarding the $K_{\text{m}}$(d-Ala) and $K_{\text{m}}$(ATP) results, the chosen HTP assay conditions were as follows: 100 mM Tris-HCl pH 8, 10 mM KCl, 10 mM MgCl$_2$, 0.4 mM $Bp$Ddl, 200 $\mu$M ATP and 5 mM d-Ala.

To determine whether the assay was linear under the chosen assay conditions, a time course experiment was carried out. A total of 24 reactions were started at time points 2.5 min apart during 90 min, and eight replicates included. Additionally, the same experiment was done in the absence of enzyme. These results were used to determine the $Z'$ score (Eqn 1) and the signal-to-noise ratio of the experiment.

$$Z' = 1 - \frac{3(\text{SD}_{\text{max}} + \text{SD}_{\text{min}})}{\text{mean (max) – mean (min)}},$$

where $\text{SD}$ = standard deviation, $\text{max}$ = maximum signal, $\text{min}$ = minimum signal.

### Determination of DMSO tolerance and $\alpha$-cycloserine IC$_{50}$

Using the HTP assay conditions, a dose–response experiment was performed using variable concentrations of DMSO (from 0% to 27%) or DCS (0–300 $\mu$m). A 2% inhibition of enzyme activity was noted at 6% DMSO.

### Compound library screens

#### Fragment library screen using BLI

A fragment library from the Drug Discovery Unit (DDU, University of Dundee) was screened using an OctetRED384 (ForteBio-Molecular Devices, Fremont, CA, USA). The protein was immobilised on hexahistidine tag high affinity sensors (ForteBio), and the experiment carried out in black plates (Greiner). We used TEV protease as a control and to identify promiscuous binders, and ATP as the control ligand. During the assay performance, we noticed a low binding signal (lower than 0.1 nm) in the presence of ATP. Selected hits were tested for enzyme inhibition using the coupled enzymatic assay at a fixed concentration of 500 $\mu$m, in the presence of 2 mM d-Ala. One compound with an inhibition effect greater than 20% was followed up for $K_i$ determination.

#### ESC library screen using the malachite green-based assay

A small selection of compounds from the BioAscent compound cloud and the NCC library of drug compounds from the NIH were tested at a concentration of 10 $\mu$m. Each screening plate included positive (normal reaction without compounds added) and negative controls (without enzyme). Compounds were dispensed using a Labcyte Echo acoustic dispenser. The assay was carried out using the HTP conditions and allowed to proceed for 1 h before stopping the reaction by adding BIOMOL green. Data were analysed using ActivityBase XE (www.idbs.com). A compound was considered a hit when the normalised percentage effect was higher than the median plus three times the standard deviation. The final DMSO concentration during screening reached 0.075% so the assay has not been compromised. Selected hits were re-tested using the HTP assay conditions in the presence and absence of enzyme with a final concentration of 20 $\mu$m to filter false positives. Compounds inhibiting more than 30% of $Bp$Ddl activity were selected for IC$_{50}$ determination. The dose–response experiment was carried out using the same conditions and in the presence or absence of 0.01% (w/v) Tween-20 and 1 mM TCEP. Data were analysed using ActivityBase XE and Microsoft EXCEL.

#### Small diversity set screen using the malachite green-based assay

A small diversity set library was screened under HTP assay conditions modified to target the d-Ala pocket, at pH 7.5, and including 0.01% (w/v) Tween-20 and 1 mM TCEP in the assay buffer. Consequently, further assay optimisation was required. $K_{\text{m2}}$(d-Ala) and $K_{\text{m}}$(ATP) at pH 7.5 were determined as described before using 1 mM $Bp$Ddl. The time course experiment was performed with 3, 2 and 1 mM d-Ala and 500 $\mu$m ATP. The compound set was used at a final concentration of 30 $\mu$m. The reaction was carried out with 1 mM d-Ala and 0.5 mM ATP, and incubated for 12 min then absorbance was measured using a PHERAstar (BMG Labtech, Ortenberg, Germany). Data were analysed using ActivityBase XE and Microsoft EXCEL and any compound providing 30% inhibition was considered a hit. Validation of hits was performed by dose–response experiments using different concentrations (1/10 dilutions starting at 100 $\mu$m) in duplicate.

### AMP detection

The crystallographic analyses revealed the presence of AMP-bound to the protein even when no nucleotide was added to the sample. Therefore, the AMP-Glo kit from Promega (Madison, WI, USA) was used to quantify the presence of AMP in the purified $Bp$Ddl. The reaction was performed in a 96-well white plate (Greiner), using 25 $\mu$L of sample, in triplicate. The signal was measured with a FLUOstar OPTIMA microplate reader (BMG Labtech). The control protein, selected because it does not bind AMP/ADP/ATP, was an amidase called BaKynB [34]. Samples were denatured before AMP quantification using three different treatments. These included heating (20 min at 100 °C prior to the assay performance), exposure to extreme pH (2 min incubation at pH 2, then pH 10, then 7) and digestion with chymotrypsin. The samples used were $Bp$Ddl, $Ba$KynB and...
\textit{B. pseudomallei} \textit{d}-alanine-\textit{d}-alanine ligase

\textit{B. pseudomallei} in the presence of 2 \textmu m AMP (\textit{BkynB-AMP}) as a control. Chymotrypsin was added to a final ratio chymotrypsin : sample of 1 : 20 and the mixtures incubated O/N at 25 °C. Then, the samples were filtered using an Amicon ULTRA 3K concentrator (Merck & Co). The protein concentration was 4 mg mL\(^{-1}\) for \textit{BkynB} and 25 mg mL\(^{-1}\) for \textit{BpDdl}. Data are presented in Fig. 9.

**Conclusions**

We set out to assess, and potentially validate, \textit{BpDdl} as a target for the development of the new antibacterial drugs for \textit{B. pseudomallei}. Two high-resolution crystal structures of \textit{BpDdl} in complex with AMP and AMP+ \textit{d}-\textit{Ala}-\textit{d}-Ala were obtained. Comparative studies indicate aspects of Ddl that are well conserved allow us to describe conformational differences that result from ligand binding. The \textit{BpDdl} structures provided templates for a computational druggability assessment, but also identified that the enzyme was retaining AMP during isolation. This was confirmed using an AMP detection assay. The co-purification with AMP suggests that we have assayed a binary complex not the apo-enzyme. This had not been reported previously and might also apply in other Ddl studies. The \textit{BpDdl} activity was measured using two different enzyme assays and showed similar kinetic parameters to two other Ddl enzymes but different from a third. We noted a retarded phase during assay that was probably due to the presence of AMP which first has to vacate the co-factor binding site prior to catalysis.

High-throughput binding and enzymatic assays were developed to conduct compound library screens. Despite screening more than 22 500 compounds, we did not identify any good starting points for further study. Whilst our study was underway, AstraZeneca reported a larger compound library screen for \textit{BpDdl} but no lead compounds were found [56], which is consistent with our results.

Our overall assessment is that \textit{BpDdl} represents a challenging target for drug discovery. However, our progress in enabling ligand-binding studies of the enzyme means that in future a structure-based rational approach [57] combining crystallography with computational docking and modelling calculations would be feasible. In similar fashion, further HTP screens can be prosecuted but the composition of the library would have to be carefully considered.

**Acknowledgements**

We thank Helen Atkins and Phil Ireland from the Defense Science and Technology Laboratory (Dstl) and Alice Dawson for advice and support, staff at the European Screening Centre, and in the Drug Discovery Unit for advice and support, and the European Synchrotron Radiation Facility and Diamond Light Source for beam time. This work was funded by Dstl, SULSA and The Wellcome Trust (grant numbers 082596, 094090).

**Conflict of interest**

The authors declare no conflict of interest.

**Author contributions**

Participated in research design, performed the data analysis and contributed to the writing of the manuscript: LD-S, LST, SPM, DG, WNH; Conducted experiments: LD-S, LST, SPM.

**References**

9 Neuhaus FC & Baddiley J (2003) A continuum of anionic charge: structures and functions of D-alanyl-


34 Díaz-Sáez L, Srikanthnasas V, Zoltner M & Hunter WN (2014) Structures of bacterial kynurenine formamidase reveal a crowded binuclear zinc catalytic


