



**University of Dundee**

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Meng, Yutong; Li, Wenfeng; Pan, Xiangliang; Gadd, Geoffrey Michael

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# Applications of nanozymes in the environment

**Abstract:** Nanozymes are inorganic nanoparticles that mimic the enzyme-like properties in redox reactions, possessing both unique properties of nanomaterials and a catalytic function. Because of high catalytic activity, stability and multifunctionality, nanozyme are of increasingly wide interest in the fields of environmental science and technology. In this article, we review the most recent advances of nanozyme research for environmental pollutant detection and treatment. Nanozymes can be used to detect ions, molecules and organic compounds both qualitatively and quantitatively. They have also been applied for destruction multi-drug resistant bacteria and the degradation of various organic pollutants. Despite the apparent potential of nanozymes in environmental science and technology, current research and application is still limited, and so future challenges and research prospects have been highlighted.

**Keywords:** nanozyme; heavy metals; organic pollutants; antibacterial substances; organic pollutant degradation

## 1. Introduction

### 1.1 Definition of nanozyme

Most life processes in nature involve “enzymes”. Natural enzymes are a class of biomolecules that process catalytic functions.<sup>1</sup> They are mainly proteins, and their catalytic activity on the substrate can be very efficient and high specific.<sup>2</sup> However, since most natural enzymes are inhibited and denatured by non-physiological or adverse conditions, such as heat, acids, and alkalis, they are prone to degeneration and can lose their function.<sup>3-5</sup> With the rapid development of nanoscience,

23 nanotechnology and the development of nanomaterials have entered various branches of the life  
24 sciences.<sup>6, 7</sup>

25 In early research, the superoxide dismutase (SOD) mimicking activities of fullerene derivatives was  
26 discovered.<sup>8, 9</sup> Subsequently it was found that the inorganic nanomaterial Fe<sub>3</sub>O<sub>4</sub> possessed a biological  
27 activity similar to that of the natural enzyme, horseradish peroxidase (HRP).<sup>10, 11</sup> Since then, the  
28 “nanozyme” concept has developed and the number of increasing numbers of research publications.<sup>12-24</sup>

29 Nanozymes are inorganic nanoparticles that mimic enzyme-like properties in redox reactions,  
30 therefore possessing both unique properties of nanomaterials and a catalytic function.<sup>14, 23, 25-27</sup> Because  
31 of high catalytic activity, stability and multifunctionality, nanozymes have found increasingly wide  
32 potential applications in fields, such as medicine, chemical engineering, agriculture and the  
33 environment.<sup>21, 22, 28-32</sup> In fact, nanomaterials were originally considered as being a chemically inert  
34 material with no intrinsic biological effects, a nanozyme was originally defined as an enzyme or  
35 enzymic catalytic group associated with the nanomaterial surface and termed a nanomaterial  
36 hybridizing enzyme.<sup>33, 34</sup> For example, azacrown was modified onto gold nanoparticles (AuNP) by  
37 chelation with Zn<sup>2+</sup> to imbue catalytic activity in shearing phosphodiester bonds mimicking the  
38 function of RNase<sup>35</sup>. Thus the catalytic activity arises from surface-modified components and not from  
39 the nature of the nanomaterials themselves. However, with the further development and application of  
40 nanomaterials, several nanomaterials have been found to possess inherent enzymic catalytic properties,  
41 and therefore, the definition of nanozyme has become broader to include all nanomaterials that possess  
42 intrinsic enzyme-like activities.<sup>36</sup>

43

## 44 1.2 Classification and catalytic mechanisms of nanozyme

45 Nanozymes vary in structure and composition, and include metal oxides,<sup>37-52</sup> metals,<sup>17, 53-71</sup> and  
 46 carbon-based nanomaterials.<sup>72-78</sup> However, most of the catalytic reactions mediated by nanozymes  
 47 employ the following four kinds of enzymic reactions: oxidase (OXD), peroxidase (POD), catalase  
 48 (CAT) and superoxide dismutase (SOD) (Table 1).

49

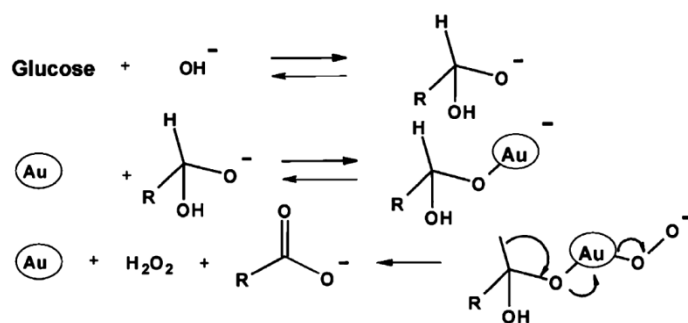
50 **Table 1** Current nanomaterials as enzyme mimics and their typical applications and representative references (GO: graphene oxide, PANI:  
 51 polyaniline, rGO: reduced graphene oxide, C-dots: carbon dots, NPs: nanoparticles, CAT-NP: catalytic nanoparticles, Dex-NZM:  
 52 dextran-coated iron oxide nanoparticles termed nanozymes, GQD: Graphene quantum dot, MNP@CTS: ferromagnetic chitosan  
 53 nanozyme, OXD: oxidase, POD: peroxidase, CAT: catalase, SOD: superoxide dismutase, HRP: Horseradish Peroxidase)

Application	Nanomaterial	Enzyme type	Reference
Detection of ions	MoS <sub>2</sub>	POD	104
	Fe <sub>3</sub> O <sub>4</sub>	POD	105
	Au	POD	106
	CoO <sub>x</sub> H-GO	POD	108
Detection of organic pollutants	PdAu	POD	112
	PANI/rGO	CAT	115
	MnO <sub>2</sub>	OXD	117
Antimicrobial and antifouling treatments	MoS <sub>2</sub>	POD	133
	C-dots	OXD	134
	C <sub>3</sub> N <sub>4</sub> @AuNPs	POD	123
	CAT-NP with Fe <sub>3</sub> O <sub>4</sub>	POD	135
	CuO NPs	POD	138
	CuO nanorods	POD	139
	Nanoceria	SOD and CAT	140
	Dex-NZM	POD	136
	CeO <sub>2-x</sub> nanorods	HRP	141
GQD	POD	72,143	
Treatment of organic pollutants	MNP@CTS	POD	149
	MnP	POD	150
	LiP	POD	153
	ZnO/CuO	POD	157
	Au/TiO <sub>2</sub>	POD	158
	Nano-eco-enzyme	POD	159,160

54

55 Natural OXDs are enzymes that catalyze an oxidation-reduction reaction, especially those involving  
 56 dioxygen (O<sub>2</sub>) as the electron acceptor.<sup>79</sup> Nanozymes based on gold<sup>80-86</sup> and copper<sup>87, 88</sup> are typical

57 representatives of OXD mimic enzymes.<sup>34</sup> The proposed mechanism of molecular activation for AuNP  
 58 catalysis is shown in Figure 1. The hydrated glucose anion is first adsorbed onto the surface of the  
 59 AuNP, and the interaction with gold surface atoms forms an electron-rich gold species which  
 60 effectively activates molecular oxygen by nucleophilic attack and produces a dioxo-gold intermediate.  
 61 The Au<sup>+</sup>-O<sub>2</sub><sup>-</sup> or Au<sup>2+</sup>-O<sub>2</sub><sup>2-</sup> couples of the dioxo-gold intermediate serve as a bridge to transfer electrons  
 62 from glucose to dioxygen. Finally, gluconic acid and H<sub>2</sub>O<sub>2</sub> are produced.<sup>33, 34</sup>



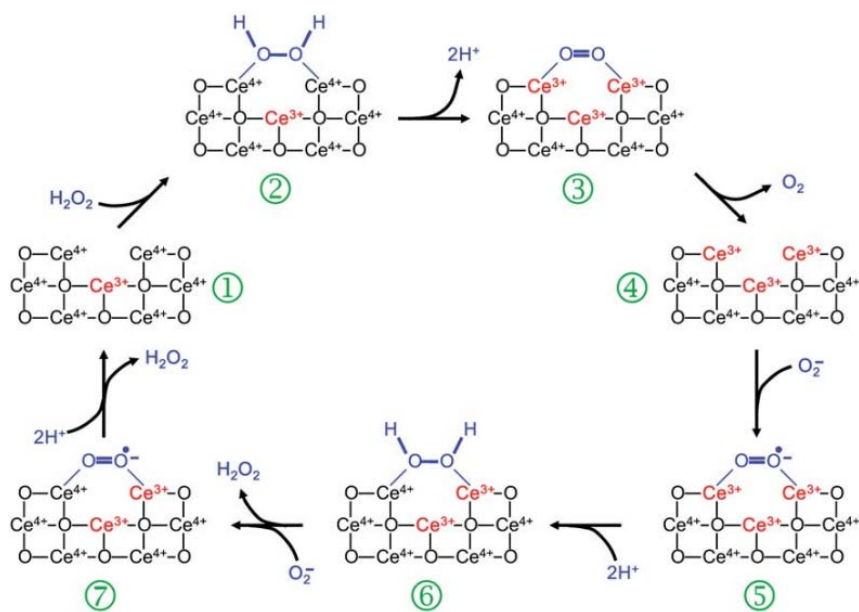
63

64 **Fig. 1** Gold nanoparticles exhibit a catalytic mechanism that mimics glucose oxidase (reproduced with permission from ref. 86, Copyright  
 65 2006 Wiley-VCH).

66 PODs can commonly attack peroxides.<sup>89</sup> Fe<sub>3</sub>O<sub>4</sub> magnetic materials possess intrinsic POD  
 67 properties<sup>10</sup> and these have been applied for the detection of both H<sub>2</sub>O<sub>2</sub> and glucose.<sup>90</sup> Other iron  
 68 oxide-based POD mimics have also been studied, including Fe<sub>3</sub>O<sub>4</sub>,<sup>91, 92</sup> Fe<sub>2</sub>O<sub>3</sub>,<sup>93, 94</sup> and doped ferrites.<sup>95</sup>  
 69 Since POD-like iron oxide has a Fenton and/or Haber-Weiss reaction mechanism (possibly  
 70 involving  $\cdot\text{OH}$  /  $\text{HO}_2\cdot$ ), nanozymes can be used for organic pollutant degradation by combining free  
 71 radical production with magnetic properties of iron oxide.<sup>34</sup>

72 CATs catalyze the decomposition of hydrogen peroxide to water and oxygen.<sup>96</sup> CeO<sub>2</sub> NPs can serve  
 73 as CAT mimic enzymes.<sup>97</sup> A series of catalytic effects is achieved by a redox reaction that forms  
 74 catalytically effective Ce<sup>4+</sup>. Ce<sup>4+</sup> is reduced by H<sub>2</sub>O<sub>2</sub> to form Ce<sup>3+</sup> and produces protons and O<sub>2</sub>. After  
 75 that, another H<sub>2</sub>O<sub>2</sub> molecule can combine with the oxygen vacancy in reaction (5), oxidizing Ce<sup>3+</sup> to

76  $\text{Ce}^{4+}$ , and releasing  $\text{H}_2\text{O}$  (Fig. 2).<sup>33, 98</sup>



77

78 **Fig. 2** Mechanism of  $\text{CeO}_2$  nanoparticles acting as catalase mimic enzymes (reproduced with permission from ref. 98, Copyright 2011

79 Royal Society of Chemistry).

80 SOD is an enzyme that alternately catalyzes the dismutation (or partitioning) of the superoxide ( $\text{O}_2^-$ )

81 radical into either molecular oxygen ( $\text{O}_2$ ) or hydrogen peroxide ( $\text{H}_2\text{O}_2$ ).<sup>99</sup> Taking Cu SOD as an

82 example, as a reducing agent,  $\text{O}_2^-$  is finally turned into  $\text{O}_2$ :  $\text{Cu}^{2+}\text{-SOD} + \text{O}_2^- \rightarrow \text{Cu}^+\text{-SOD} + \text{O}_2$ .<sup>100</sup>

83 These kinds of redox reactions can produce (or eliminate) free radicals and regulate levels of reactive

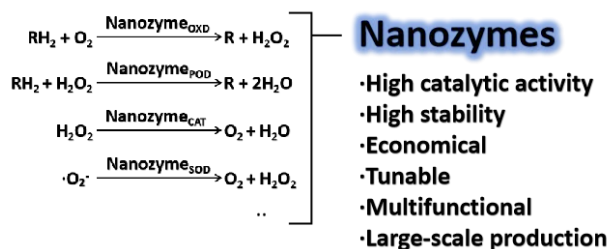
84 oxygen species (ROS). Because ROS can attack nucleic acids, proteins, polysaccharides, lipids and

85 other biological molecules, ROS releasing nanozymes can have excellent antimicrobial properties.<sup>101</sup> In

86 addition, POD-mimic nanozymes can produce hydroxyl radicals with oxidative properties. Due to these

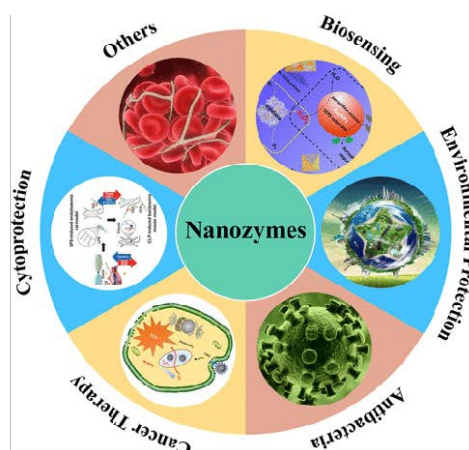
87 characteristics, nanozymes have been widely applied to environmental problems. Nanozymes can

88 detect the presence of certain pollutants, and also be used for effective treatment.



## Nanozymes

- High catalytic activity
- High stability
- Economical
- Tunable
- Multifunctional
- Large-scale production



89

90 **Fig. 3** Natural enzymic properties, advantages and applications of nanozymes (R: substrate, OXD: oxidase, POD: peroxidase, CAT:

91 catalase, SOD: superoxide dismutase) (reproduced with permission from ref. 33, Copyright 2019, American Chemical Society).

92

93 **Table 2** Nanomaterials for detection as enzyme mimics and their targets and agents (PANI: polyaniline, rGO: reduced graphene oxide,  
94 OXD: oxidase, POD: peroxidase, CAT: catalase) (adapted from refs.104-108,112,115,117)

Nanomaterial	Enzyme type	Agent	Target
MoS <sub>2</sub>	POD	2, 3-diaminophenazine	Fe <sup>2+</sup>
Fe <sub>3</sub> O <sub>4</sub>	POD	4-chloro-1-naphthol	Ag <sup>+</sup>
Au	POD	4-nitrophenol	Hg <sup>2+</sup>
CoO <sub>x</sub> H-GO	POD	Amplex Red	CN <sup>-</sup>
PdAu	POD	O-phenylenediamine	Malathion
PANI/rGO	CAT	Electrical signal	Kanamycin
MnO <sub>2</sub>	OXD	Oligonucleotides	Ochratoxin A

95

## 96 2. Nanozymes for the detection of environmental pollutants

97 Nanozymes can be used in place of natural enzymes for environmental monitoring. By employing

98 the catalytic activity of POD nanozymes, the content of hydrogen peroxide in rainwater, acid rain, and

99 heavy metals, including mercury, can be detected in environmental samples. Such nanozyme detection

100 methods are also suitable for operation under various environmental conditions and are relatively

101 simple and inexpensive, and can be easily applied to screening of, e.g. pesticides, organophosphorus

102 compounds, and other substances.

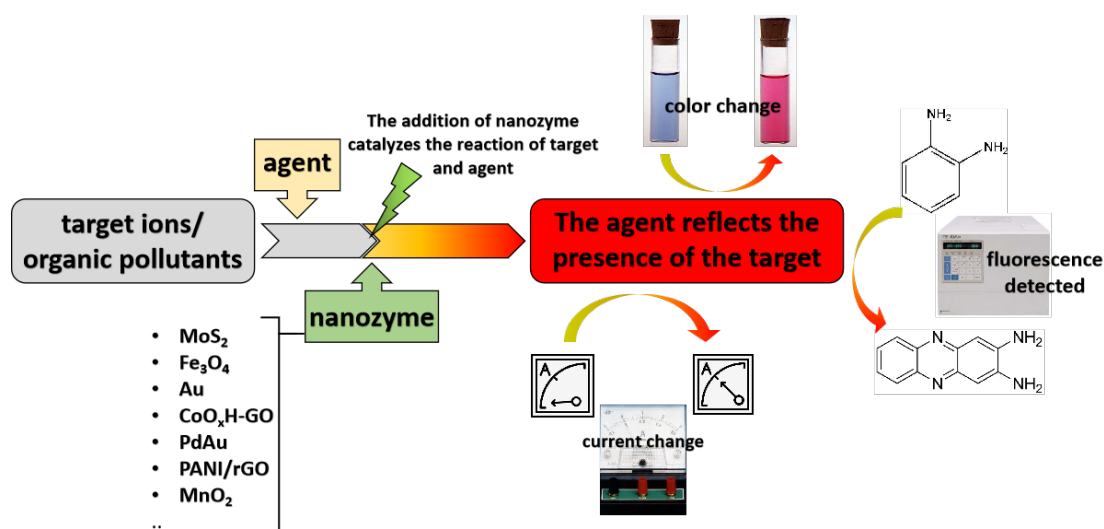
103 Whether the pollutants are ions, metals or organic compounds, nanozymes can only detect them  
104 indirectly. The basic principle of detection is that the target activates a reaction between the nanozyme  
105 and the agent, or the presence of the nanozyme causes the target to undergo a change in chemical  
106 properties and reacts with the agent to be detected (Fig. 4). An agent is usually a colorimetric sensor  
107 although there is some use of sensors such as electric current monitors (Table 2).

## 108 **2.1 Detection of Ions**

109 In the field of environmental pollution, the term heavy metals, mainly refers to such potentially  
110 biotoxic elements as mercury, cadmium, lead, chromium and arsenic. Heavy metals cannot be degraded,  
111 only transformed into different chemical species by abiotic or biotic mechanism, and they can be  
112 biologically amplified through food webs, causing harm to ecosystems and organisms, including  
113 humans.<sup>102, 103</sup>

114 A novel layered molybdenum disulfide ( $\text{MoS}_2$ ) nanosheet POD mimetic-based fluorescent catalytic  
115 biosensor was developed for the sensitive and selective detection of  $\text{Fe}^{2+}$  over the range of 0.005-0.20  
116  $\mu\text{M}$ .<sup>104</sup> The catalyst  $\text{MoS}_2$ , was synthesized from  $\text{Na}_2\text{MoO}_4 \cdot 2\text{H}_2\text{O}$ , and exhibits POD activity.  
117 O-phenylenediamine acts as the substrate and was converted to 2, 3-diaminophenazine by  $\text{MoS}_2$  in the  
118 presence of  $\text{Fe}^{2+}$ , becoming an indicator of fluorescence detection. Since the fluorescence intensity was  
119 proportional to the  $\text{Fe}^{2+}$  concentration,  $\text{Fe}^{2+}$  was successfully detected with high selectivity and  
120 sensitivity.





121

122 Fig. 4 The basic principle of nanozyme detection of ions/organic compounds (PANI: polyaniline, rGO: reduced graphene oxide) (adapted  
123 from refs.104-106,108,112,115,117)

124 Based on the POD-like properties of histidine-modified Fe<sub>3</sub>O<sub>4</sub> (his-Fe<sub>3</sub>O<sub>4</sub>) nanozyme, a simple,  
125 low-cost means to detect Ag<sup>+</sup> was developed with ultralow detection limit of 18 fg/mL.<sup>105</sup> An electron  
126 transfer sensor was conjugated to the highly active nanozyme his-Fe<sub>3</sub>O<sub>4</sub> in the presence of Ag<sup>+</sup> via a  
127 specific reaction. 4-chloro-1-naphthol was used as the substrate, nanozyme his- Fe<sub>3</sub>O<sub>4</sub> was used as the  
128 catalyst, and H<sub>2</sub>O<sub>2</sub> was used as an oxidant. When Ag<sup>+</sup> was present, the POD enzyme activity of his-  
129 Fe<sub>3</sub>O<sub>4</sub> was activated and this changed the substrate to generate insulating precipitation of  
130 benzo-4-chlorohexadienone. The insulating products attenuate the photocurrent signal which reflected  
131 the presence of Ag<sup>+</sup>. His-Fe<sub>3</sub>O<sub>4</sub> nanozymes could make photoelectrochemical immunoassays chemical  
132 easier and less expensive.

133 Mercury in the environment exists in many chemical forms, with divalent mercury being one of the  
134 most stable. Because of the toxicity and bioaccumulation potential of mercury, detection in the  
135 environment is very important. One approach harnessed the strong affinity between AuNP and Hg<sup>2+</sup>,  
136 meaning that mercury can attach to the surface of gold nanozymes to form a gold amalgam. As a POD  
137 nanozyme, AuNP activity was further enhanced by precipitation of Hg<sup>2+</sup>, and NaBH<sub>4</sub> supplied as a

138 reducing agent, reduce the substrate 4-nitrophenol more quickly. 4-nitrophenol reduction produces a  
139 colour change with ultra-high sensitivity to  $\text{Hg}^{2+}$  and detection limits down to 1.45 nM. This AuNP  
140 nanozyme was stable and recyclable.<sup>106, 107</sup>

141 A cobalt oxide/oxide-modified graphene oxide ( $\text{CoO}_x\text{H-GO}$ ) nanozyme having POD-like catalytic  
142 activity has been used as an effective detecting agent for  $\text{CN}^-$  (selectivity>100-fold).<sup>108</sup> The principle  
143 relied on the significant inhibitory effect of  $\text{CN}^-$  on the catalytic activity of the  $\text{CoO}_x\text{H-GO}$  nanozyme.  
144 When Amplex Red was used as a substrate and colorimetric reagent, with the  $\text{CoO}_x\text{H-GO}$  nanozyme as  
145 catalyst and  $\text{H}_2\text{O}_2$  as oxidant, the increase in  $\text{CN}^-$  concentration was clearly reflected by a decrease in  
146 red colour. This method can also be applied to complex wastewaters (e.g. sea water) at low cost.

147

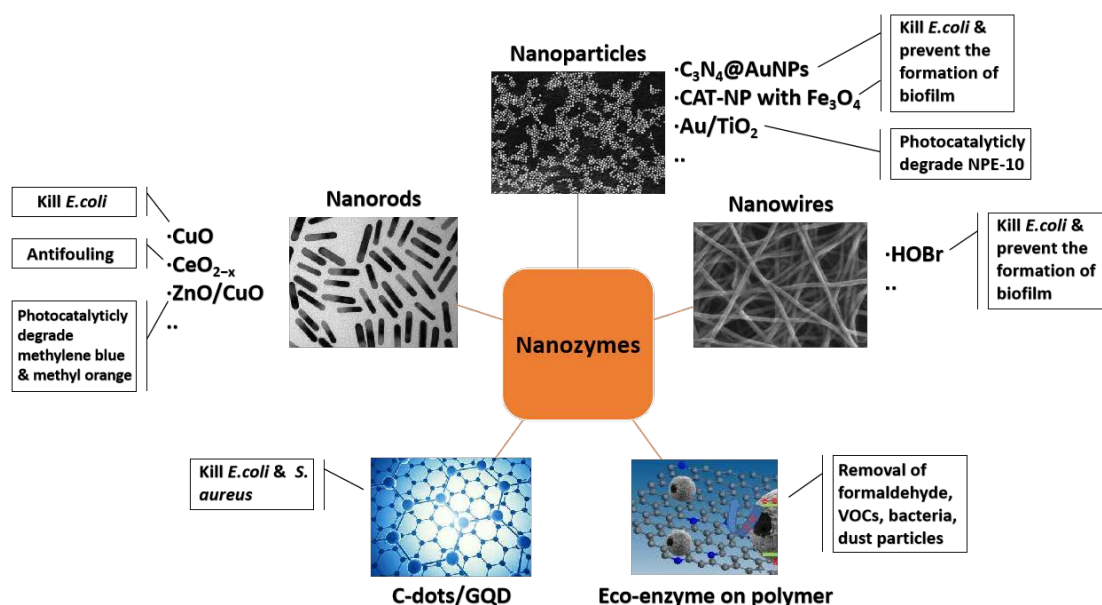
## 148 **2.2 Detection of organic pollutants**

149 There are several kinds of organic pollutants in the environment, and many are difficult to detect.  
150 Nanozymes can be used to detect pesticides, antibiotics and other toxic organic compounds.<sup>109-111</sup>

151 Pesticide pollution refers to pollution caused by pesticides, and their toxic metabolites,  
152 degradation products and impurities remaining in organisms, agricultural by-products and the  
153 environment after pesticide use has exceeded the maximum residue limits. The toxicity of residual  
154 pesticides to living organisms is called the pesticide residue, and this can contaminate the soil,  
155 atmosphere and groundwater. In order to detect pesticide residues, various nanozymes have been  
156 examined. For example, O-phenylenediamine was used as the substance to be oxidized, and  
157 palladium-gold (PdAu) bimetallic nanozyme as the catalyst. Because the POD activity of nanozyme  
158 was selectively quenched with increasing concentrations of malathion, is PdAu nanozyme could be  
159 used for detection of is low-toxic insecticide. This assay was highly sensitive (60 ng/ml) and of low

160 cost.<sup>112-114</sup>

161 Kanamycin is an antibiotic which can be detected by an innovative gas pressure-based biosensing  
162 platform.<sup>115, 116</sup> A polyaniline nanowire functionalized reduced graphene oxide (PANI / rGO) framework  
163 was used as a CAT-like nanozyme that catalyzed the reduction of hydrogen peroxide to produce  
164 oxygen. The existence of kanamycin triggered strand displacement amplification which affected the  
165 CAT nanozyme and reflected the presence of kanamycin as an electrical signal which increased with  
166 increasing kanamycin concentration.<sup>115, 116</sup>



167

168 **Fig. 5** Different forms of nanozymes and their applications in environmental biotechnology (C-dots: carbon dots, NPs: nanoparticles,

169 CAT-NP: catalytic nanoparticles, GQD: Graphene quantum dot, *E.coli*: *Escherichia coli*, *S.aureus*: *Staphylococcus aureus*, NPE-10:

170 nonylphenyl poly (oxyethylene) ethers, VOCs: volatile organic compounds) (adapted from refs. 72, 133-141,149-160).

171 Ochratoxin is a mycotoxin that has attracted worldwide attention. It is one of a group of important,

172 food-contaminating mycotoxins produced by seven species of *Aspergillus* and six species of

173 *Penicillium*, four of which are the most toxic and widely distributed in the agricultural products. The

174 most widely polluting and damaging to human health is ochratoxin A (OTA). One detection method

175 was based on the biotin-streptavidin reaction.<sup>117, 118</sup> Here, 3, 3', 5, 5'-tetramethylbenzidine (TMB) was

176 used as a substrate and a colorimetric reagent, and MnO<sub>2</sub> nanosheets were used as an OXD nanozyme  
177 to oxidize TMB to a blue colour TMB Ox. However, when OTA was present, acid-2-phosphate was  
178 converted to ascorbic acid, which reduced the MnO<sub>2</sub> nanosheet to Mn<sup>2+</sup> which cannot oxidize TMB.  
179 This method for detecting OTA possessed high sensitivity, with a limit of detection being 0.069 nM.<sup>117,</sup>

180 <sup>118</sup>

### 181 **3. Application of nanozymes in environmental treatment**

#### 182 **3.1 Nanozymes as antimicrobial and antifouling agents**

183 Environmental antibiotic resistance is a rapidly increasing problem in recent years. Abuse of  
184 antibiotics can lead to the emergence of multi-drug resistant bacteria as well as causing environment  
185 pollution. Therefore, it is important to develop new antimicrobial agents that are highly effective,  
186 environmentally-friendly and which avoid or minimize drug resistance. Effects of antibacterial  
187 nanomaterials are multifaceted, which can make it difficult for bacteria to develop drug resistance.  
188 Compared with traditional nanomaterials, nanozymes have possess higher biosafety and show promise  
189 as an effective antibacterial material.<sup>119, 120</sup>

190 Reactive oxygen species (ROS) play an important role in cellular defence against pathogen  
191 invasion<sup>121, 122</sup>, and nanozymes have the ability to regulate level of ROS free radicals.<sup>123</sup> This ability  
192 confers an antibacterial function. For instance, hydrogen peroxide is a commonly used disinfectant  
193 because it can be decomposed to generate free radicals, thereby attacking cellular components of  
194 bacteria, such as membranes, proteins and nucleic acids.<sup>124, 125</sup> However, the efficiency of generating  
195 free radicals is low, and the addition of a catalyst greatly accelerates the reaction. Nanomaterials with  
196 POD mimicking enzyme activity can be used as such a catalyst to improve the transformation of  
197 hydrogen peroxide to free hydroxy radicals and thus enhance sterilization.<sup>126-129</sup> It was found that in the

198 presence of low concentrations of hydrogen peroxide, trace amounts of nanozymes could kill 100%  
199 *Escherichia coli* (*E. coli*), while the sterilization efficiency of hydrogen peroxide alone was less than  
200 15%.<sup>130</sup>This study also found that vanadium pentoxide nanowires with vanadium haloperoxidase  
201 activity effectively inhibited formation of biofilm. In the presence of hydrogen peroxide, this substance  
202 can oxidize bromide ions to produce hypobromous acid (HOBr) and singlet oxygen, which has strong  
203 antibacterial activity. Applying vanadium pentoxide nanowires to the surface of stainless steel inhibited  
204 microbial adhesion, thus effectively preventing the formation of biofilm, and therefore has potential in  
205 antifouling applications, e.g. for ship hulls.

206 Photocatalytic cooperation with nanozymes can kill bacteria very effectively.<sup>131, 132</sup> POD activity  
207 of a MoS<sub>2</sub> nanozyme was activated by lowering the pH which altered the surface charge of MoS<sub>2</sub> from  
208 negative to positive. The activated MoS<sub>2</sub> nanozyme catalyzed the decomposition of H<sub>2</sub>O<sub>2</sub>, and the  
209 resulting •OH destroyed cell integrity to achieve an antibacterial effect. The advantage of this method is  
210 that this antibacterial treatment can be accomplished simply by controlling the light.<sup>133</sup> Light-driven  
211 carbon dots (C-dots) were used as OXD nanozymes to kill *E. coli* and *S. aureus* by photosensitization,  
212 both ambient light and UV irradiation being tested.<sup>134</sup>

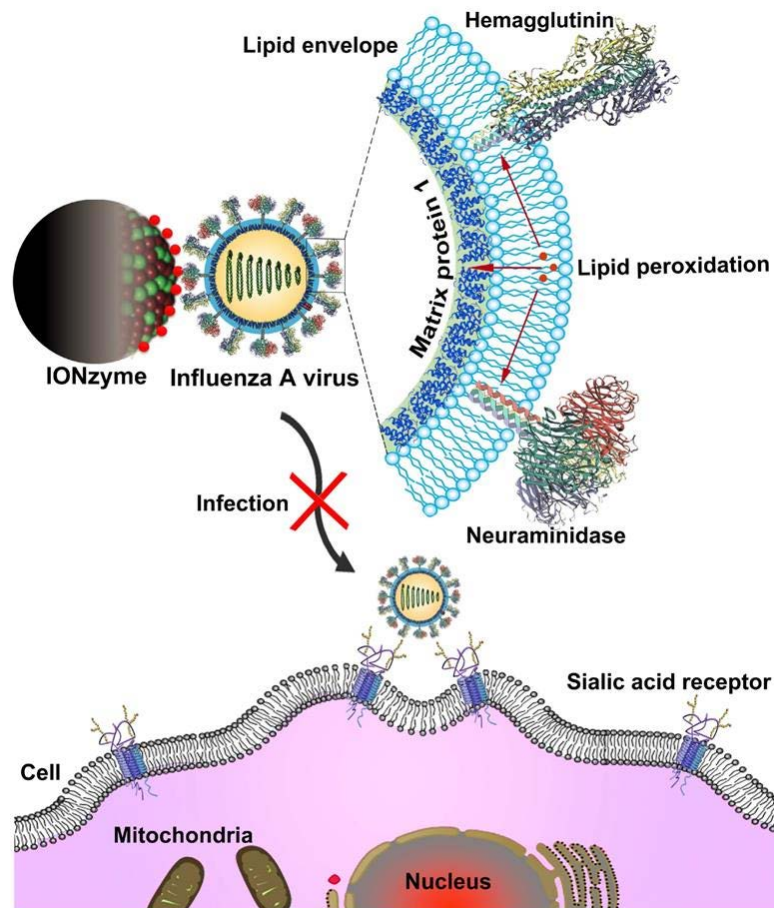
213 Ultra-thin graphite carbon nitride (g-C<sub>3</sub>N<sub>4</sub>) AuNPs can be used as POD to efficiently catalyze the  
214 decomposition of H<sub>2</sub>O<sub>2</sub> into •OH to kill drug-resistant Gram-negative and Gram-positive bacteria. This  
215 method was also very effective in destroying and preventing biofilm regeneration.<sup>123</sup> Catalytic  
216 nanoparticles containing biocompatible Fe<sub>3</sub>O<sub>4</sub> and dextran coated iron oxide NPs also have POD  
217 activity and can be used to decompose H<sub>2</sub>O<sub>2</sub> to produce •OH. This method degrades the biofilm matrix  
218 and kills bacteria quickly.<sup>135, 136</sup>

219 Nano-CuO, as an artificial POD, is another kind of antibacterial material.<sup>137</sup> Nanozyme based

220 hydrogel (nanozyme-gel) CuO NPs and CuO nanorods (NRs) can be used to catalyze the  
221 decomposition of H<sub>2</sub>O<sub>2</sub> to kill *E. coli*. Interestingly, the catalytic activity of CuO NRs showed  
222 significant catalytic enhancement under visible light irradiation, allowing light control of antibacterial  
223 activity.<sup>138, 139</sup>

224 Nanoceria has a variety of enzymic properties. A nanoceria was discovered with both CAT and  
225 SOD properties, which was controlled by the Ce<sup>3+</sup>/Ce<sup>4+</sup> ratio.<sup>140</sup> When the Ce<sup>3+</sup>/Ce<sup>4+</sup> ratio and its  
226 activity are regulated, the cerium oxide nanozyme converts between these two properties. The  
227 superoxide anion is converted to H<sub>2</sub>O<sub>2</sub> like SOD which is then further converted to H<sub>2</sub>O and O<sub>2</sub> like  
228 CAT, and was effective in killing *E. coli* and *S. aureus*. CeO<sub>2-x</sub> nanorods can somehow prevent  
229 biofouling in an aqueous environment because of haloperoxidase activity. One CeO<sub>2-x</sub> NRs is stable in  
230 water, including marine environments, and can reduce bacteria adhesion by 70% compared to  
231 conventional PVA fibres.<sup>96-99, 141, 142</sup>

232 Graphene quantum dots (GQD) also have multiple enzymatic properties like nanoceria. GQD can  
233 exhibit POD properties, e.g. addition of GQD can significantly improve the antibacterial activity of  
234 H<sub>2</sub>O<sub>2</sub>, while GQD/AgNP hybrids can express an OXD function.<sup>72, 143</sup>



235

236 Fig.6 Schematic of viral lipid peroxidation by IONzymes for virus inactivation. IONzymes directly contact with IAVs particles

237 and collapses the viral lipid envelope by enhancing the level of lipid peroxidation, which further produces free radicals to destroy

238 neighbouring proteins, including haemagglutinin, neuraminidase, and matrix protein 1, and impaires various viral structures and

239 functions resulting in failed infection. (IONzymes: iron oxide nanozymes, IAVs: inactivated A viruses) (reproduced with permission

240 from ref. 144, Copyright 2019, Ivyspring International Publisher) .

241 Most recently, iron oxide nanozymes (IONzymes) have been demonstrated to effectively

242 inactivate A viruses (IAVs) by inducing envelope lipid peroxidation and destruction of the integrity of

243 neighbouring proteins, including haemagglutinin, neuraminidase, and matrix protein 1 (Fig. 6).

244 Furthermore, IONzymes possess broad-spectrum antiviral activity against 12 subtypes of IAVs

245 (H1~H12).<sup>144</sup>

246

### 247 3.2 Treatment of organic pollutants in water

248 Nanozymes have shown excellent qualities for the treatment of contaminated wastewater because  
249 they (i) can treat compounds that are normally difficult to biodegrade, (ii) can operate independently of  
250 the contaminant concentration, (iii) can operate over a wide range of pH, temperature and salinities, (iv)  
251 are not subject to inhibition from biofouling, (v) are relatively simple and easy to control, and (vi)  
252 possess high stability and are recyclable.<sup>145-148</sup>

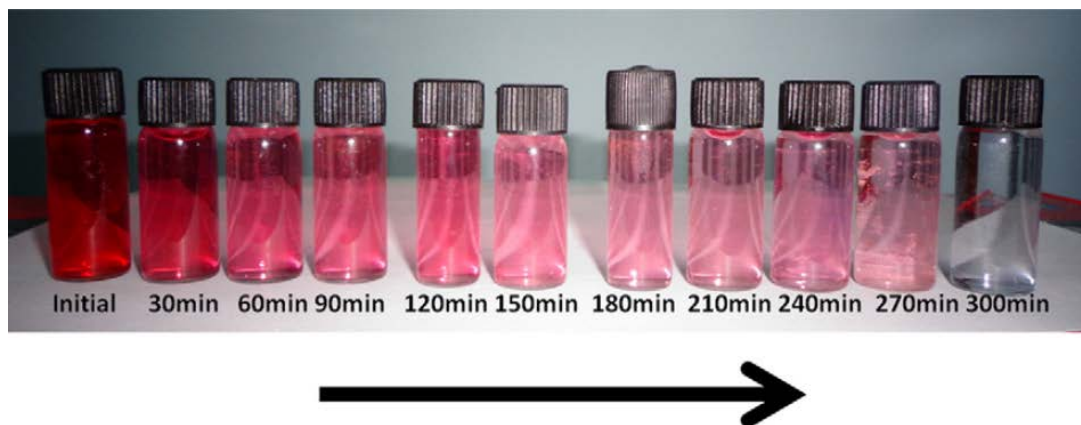
253 Ferromagnetic chitosan nanozymes (MNP@CTS) has been synthesized which are more  
254 catalytically active than conventional ferromagnetic nanozymes for the degradation of phenol.<sup>149</sup> This  
255 type of nanozyme has excellent POD activity and can be prepared and regenerated easily at a lower  
256 cost than conventional horseradish peroxidase (HRP). MNP@CTS removed over 95% phenol from an  
257 aqueous solution within 5 h under the optimum conditions (pH range 2-10). What is more important is  
258 that the MNP@CTS are very stable and could be regenerated for reuse for at least ten cycles. In  
259 otherwork Mn(III)-chelate arising from manganese POD was pumped into a reaction vessel containing  
260 organic contaminants.<sup>150</sup> It was found that the Mn(III)-chelate efficiently oxidized 2, 4-dichlorophenol  
261 and 2, 4, 6-trichlorophenol. The Mn(III) was oxidized to Mn(III) at an initial rate of 78% under  
262 optimized conditions. This nanozyme had the advantage of retaining 88% of the initial MnP nanozyme  
263 activity after about 24 hours of continuous operation.

264 Aromatic compounds are a class of compounds possessing benzene ring structures. They are  
265 normally structurally stable, resistant to degradation, and often highly toxic, causing serious pollution  
266 to the environment and injurious to human health.<sup>151, 152</sup> Lignin peroxidase (LiP) from a variety of  
267 sources can degrade a variety of recalcitrant aromatic compounds including polycyclic aromatic and  
268 phenolic compounds. In addition to exhibiting the normal properties of POD, the enzyme can form a



269 substrate cation radical with non-phenolic aromatic compounds and catalyze high-potential  
270 one-electron oxidation.<sup>153, 154</sup>

271 Dyes are still a major problem in water pollution.<sup>155, 156</sup> A new thermal-decomposition method was  
272 used to prepare ZnO nanorods and ZnO/CuO nanocomposites with different weight ratios (the  
273 maximum efficiency was observed for 5% CuO loaded on ZnO).<sup>157</sup> Organic dyes such as methylene  
274 blue and methyl orange were photocatalytically degraded by addition of composite catalyst under  
275 visible light irradiation. Preparation of this nanozyme is simple, rapid and economical.



276

277 **Fig. 7** Photography image represents change in colour of textile dyes using ZnO/CuO (95:5) catalyst for different exposure time under  
278 visible light irradiation.) (adapted from refs. 157).

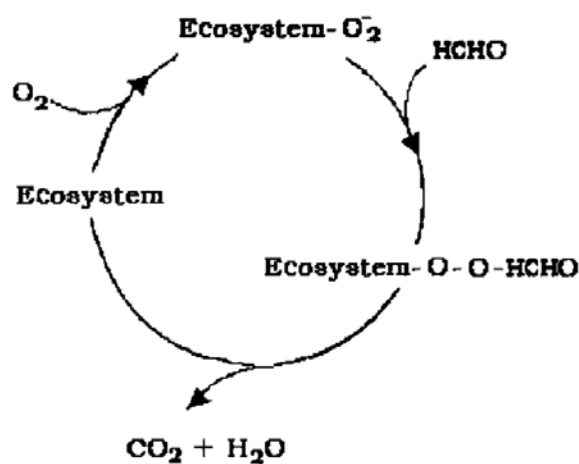
279 In other research, Au/TiO<sub>2</sub> powder was prepared by a sol-gel method, and this could  
280 photocatalytically degrade nonylphenyl poly (oxyethylene) ethers (NPE-10) under sunlight  
281 illumination.<sup>158</sup> The degradation rate of NPE-10 after irradiation for 4h was 91.8%, while that of  
282 TiO<sub>2</sub>-P-25% and undoped TiO<sub>2</sub> was 66% and 52.6%, respectively.

283

### 284 3.3 Treatment of indoor air pollution

285 Nano-ecological-enzyme air purification material, another kind of nanozyme, is a new type of  
286 functional material with high purification efficiency, low wind resistance and sterilizable. It can be used

287 in air purifiers to remove indoor dust, microorganisms, formaldehyde and other volatile organic  
 288 pollutants. The material is made of activated carbon fibre (ACF) and porous polymer composites and  
 289 loaded with nano-silver and eco-enzyme catalyst.<sup>159, 160</sup> Eco-enzymatic catalysts are supported in the  
 290 nanoporous carbon structure and become composite macromolecules with active oxygen carriers.  
 291 Enzymatic catalysts in the nanopores combine with oxygen to form highly active superoxide ions and  
 292 oxidation-reduction active sub-fields, similar to POD that are widely distributed in nature. The contact  
 293 area between the eco-enzyme catalyst and adsorbed formaldehyde in the nanopores is very large. The  
 294 active catalyst molecule quickly binds the formaldehyde molecule and, after a series of  
 295 oxidation-reductase catalytic reactions, different intermediate peroxide molecules are formed. Finally,  
 296 the formaldehyde is oxidized to water and carbon dioxide. The eco-enzyme catalyst quickly returns to  
 297 its original state and can bind again to oxygen molecule in the atmosphere. This process of enzymatic  
 298 oxidation-reduction can therefore be repeated multiple times (Fig. 8). Organic molecules such as  
 299 formaldehyde and microbial propagules in the air can be absorbed by the nanopores through an  
 300 autonomous cycle and thus maintain the long-term purification effect of the composite material. The  
 301 average purification rate of formaldehyde in two hours is 91.9%.<sup>159, 160</sup>



302

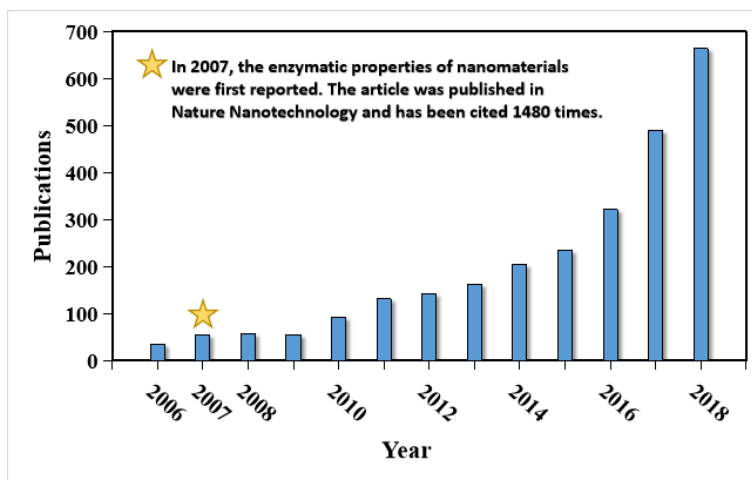
303 **Fig. 8** Degradation of formaldehyde in air by nano-ecological-enzymes (adapted from ref.159).

304

305 **4. Conclusions and research prospects**

306 In the past 10 years, nanozyme research has been carried out in more than 220 laboratories in 26  
307 countries (Fig. 7), and nanozyme applications have also extended to biology, medicine, agriculture,  
308 environmental protection and other fields.<sup>36</sup> In the field of environmental biotechnology, nanozymes  
309 have multi-functional applications ranging from pollutant detection to treatment. Nanozymes can not  
310 only achieve the specificity and efficiency of natural enzymes, but of function independently of various  
311 environmental factors that may affect biotic systems, e.g. extremes of temperature and pH.

312



313

314 **Fig. 9** The number of published papers on nanozymes has annually risen (source of data in the table: Google Scholar, December 31, 2018)

315 (adapted from ref.36).

316

317 Nanozymes also have several limitations, although nanozymes overcome many restrictive factors  
318 active against natural enzymes, most catalytic activities of nanozymes are still much lower than the  
319 corresponding natural enzymes. At present, research on nanozymes mainly concentrates on redox  
320 enzyme mimics, such as OXD, POD, CAT and SOD, with much less attention given to other enzymes  
321 that may be important in the degradation of some polymers. In addition, although the efficiency of

322 pollutant treatment is high, the cost of industrial treatment can be higher than traditional pollution  
323 treatment methods.

324 It is evident that nanozymes show good performance in small-scale experiments, but their  
325 application in environmental engineering is still limited, mainly because catalytic nanozyme devices  
326 require high precision technology, and an extended service life if such shortcomings can be overcome,  
327 applications of nanozymes for pollutant treatment industry may bring huge benefits. In addition,  
328 nanozymes can be combined with a variety of composite materials that allow application to changing  
329 environmental problems, which is also a challenging topic.<sup>161, 162</sup>

330 Although current research on nanozymes is still limited, the future of nanozyme technology  
331 seems promising. In the field of environmental biotechnology, research on nanomaterials is increasing  
332 and new nanozymes continue to emerge significant research questions include how to effectively kill  
333 bacteria under adverse conditions, and how to degrade very recalcitrant organic polymers. Integration  
334 of nanozyme technology in the fields of environment, biology, agriculture and medicine could result in  
335 multi-level benefits for industrial development and human health.

336

### 337 References

- 338 1. U. Bornscheuer, G. Huisman, R. J. Kazlauskas, S. Lutz, J. Moore and K. Robins, Engineering  
339 the third wave of biocatalysis, *Nature*, 2012, **485**, 185.
- 340 2. K.-E. Jaeger and T. Eggert, Enantioselective biocatalysis optimized by directed evolution,  
341 *Current Opinion in Biotechnology*, 2004, **15**, 305-313.
- 342 3. J.-P. Genet, Asymmetric catalytic hydrogenation. Design of new Ru catalysts and chiral  
343 ligands: from laboratory to industrial applications, *Accounts of chemical research*, 2003, **36**,

- 344 908-918.
- 345 4. M. Behrens, F. Studt, I. Kasatkin, S. Kühl, M. Hävecker, F. Abild-Pedersen, S. Zander, F.  
346 Girsdsies, P. Kurr and B.-L. Kniep, The active site of methanol synthesis over Cu/ZnO/Al<sub>2</sub>O<sub>3</sub>  
347 industrial catalysts, *Science*, 2012, **336**, 893-897.
- 348 5. B. Meunier, S. P. de Visser and S. Shaik, Mechanism of oxidation reactions catalyzed by  
349 cytochrome P450 enzymes, *Chemical reviews*, 2004, **104**, 3947-3980.
- 350 6. D. R. Paul and L. M. Robeson, Polymer nanotechnology: nanocomposites, *Polymer*, 2008, **49**,  
351 3187-3204.
- 352 7. C. Guozhong, *Nanostructures and nanomaterials: synthesis, properties and applications*,  
353 World scientific, 2004.
- 354 8. L. L. Dugan, J. K. Gabrielsen, P. Y. Shan, T.-S. Lin and D. W. Choi, Buckminsterfullerenol  
355 free radical scavengers reduce excitotoxic and apoptotic death of cultured cortical neurons,  
356 *Neurobiology of disease*, 1996, **3**, 129-135.
- 357 9. L. L. Dugan, D. M. Turetsky, C. Du, D. Lobner, M. Wheeler, C. R. Almli, C. K.-F. Shen, T.-Y.  
358 Luh, D. W. Choi and T.-S. Lin, Carboxyfullerenes as neuroprotective agents, *Proceedings of*  
359 *the National Academy of Sciences*, 1997, **94**, 9434-9439.
- 360 10. L. Gao, J. Zhuang, L. Nie, J. Zhang, Y. Zhang, N. Gu, T. Wang, J. Feng, D. Yang and S. Perrett,  
361 Intrinsic peroxidase-like activity of ferromagnetic nanoparticles, *Nature nanotechnology*, 2007,  
362 **2**, 577.
- 363 11. J. M. Perez, Iron oxide nanoparticles: hidden talent, *Nature Nanotechnology*, 2007, **2**, 535.
- 364 12. A. Karakoti, S. Singh, J. M. Dowding, S. Seal and W. T. Self, Redox-active radical scavenging  
365 nanomaterials, *Chemical Society Reviews*, 2010, **39**, 4422-4432.

- 366 13. J. Xie, X. Zhang, H. Wang, H. Zheng and Y. Huang, Analytical and environmental  
367 applications of nanoparticles as enzyme mimetics, *TrAC Trends in Analytical Chemistry*, 2012,  
368 **39**, 114-129.
- 369 14. H. Wei and E. Wang, Nanomaterials with enzyme-like characteristics (nanozymes):  
370 next-generation artificial enzymes, *Chemical Society Reviews*, 2013, **42**, 6060-6093.
- 371 15. G. Li-Zeng and Y. Xi-Yun, Discovery and current application of nanozyme, *Progress in*  
372 *Biochemistry and Biophysics*, 2013, **40**, 892-902.
- 373 16. W. He, W. Wamer, Q. Xia, J.-j. Yin and P. P. Fu, Enzyme-like activity of nanomaterials,  
374 *Journal of Environmental Science and Health, Part C*, 2014, **32**, 186-211.
- 375 17. Y. Lin, J. Ren and X. Qu, Nano-gold as artificial enzymes: hidden talents, *Advanced materials*,  
376 2014, **26**, 4200-4217.
- 377 18. Y. Lin, J. Ren and X. Qu, Catalytically active nanomaterials: a promising candidate for  
378 artificial enzymes, *Accounts of chemical research*, 2014, **47**, 1097-1105.
- 379 19. L. J. Prins, Emergence of complex chemistry on an organic monolayer, *Accounts of chemical*  
380 *research*, 2015, **48**, 1920-1928.
- 381 20. L. Zheng, J. Zhao, X. Niu and Y. Yang, Nanomaterial-based peroxidase enzyme mimics with  
382 applications to colorimetric analysis and electrochemical sensor, *Materials Review*, 2015, **29**,  
383 129.
- 384 21. X. Wang, Y. Hu and H. Wei, Nanozymes in bionanotechnology: from sensing to therapeutics  
385 and beyond, *Inorganic Chemistry Frontiers*, 2016, **3**, 41-60.
- 386 22. L. Gao and X. Yan, Nanozymes: an emerging field bridging nanotechnology and biology, *Sci*  
387 *China Life Sci*, 2016, **59**, 400-402.

- 388 23. R. Ragg, M. N. Tahir and W. Tremel, Solids go bio: inorganic nanoparticles as enzyme mimics,  
389 *European Journal of Inorganic Chemistry*, 2016, **2016**, 1906-1915.
- 390 24. E. Kuah, S. Toh, J. Yee, Q. Ma and Z. Gao, Enzyme mimics: advances and applications,  
391 *Chemistry–A European Journal*, 2016, **22**, 8404-8430.
- 392 25. Q. Wang, H. Wei, Z. Zhang, E. Wang and S. Dong, Nanozyme: An emerging alternative to  
393 natural enzyme for biosensing and immunoassay, *TrAC Trends in Analytical Chemistry*, 2018,  
394 **105**, 218-224.
- 395 26. L. Pasquato, P. Pengo and P. Scrimin, Nanozymes: Functional nanoparticle-based catalysts,  
396 *Supramolecular Chemistry*, 2005, **17**, 163-171.
- 397 27. Y. Xi-Yun, Nanozyme: a new type of artificial enzyme. *Journal*, 2018.
- 398 28. P. Weerathunge, R. Ramanathan, R. Shukla, T. K. Sharma and V. Bansal, Aptamer-controlled  
399 reversible inhibition of gold nanozyme activity for pesticide sensing, *Analytical chemistry*,  
400 2014, **86**, 11937-11941.
- 401 29. J. Golchin, K. Golchin, N. Alidadian, S. Ghaderi, S. Eslamkhah, M. Eslamkhah and A.  
402 Akbarzadeh, Nanozyme applications in biology and medicine: an overview, *Artificial cells,*  
403 *nanomedicine, and biotechnology*, 2017, **45**, 1069-1076.
- 404 30. D. Jiang, D. Ni, Z. T. Rosenkrans, P. Huang, X. Yan and W. Cai, Nanozyme: new horizons for  
405 responsive biomedical applications, *Chemical Society Reviews*, 2019.
- 406 31. U. Wille and E. Juaristi, *Encyclopedia of Physical Organic Chemistry, 6 Volume Set*, John  
407 Wiley & Sons, 2017.
- 408 32. D. Duan, K. Fan, D. Zhang, S. Tan, M. Liang, Y. Liu, J. Zhang, P. Zhang, W. Liu and X. Qiu,  
409 Nanozyme-strip for rapid local diagnosis of Ebola, *Biosensors and Bioelectronics*, 2015, **74**,

- 410 134-141.
- 411 33. Y. Huang, J. Ren and X. Qu, Nanozymes: classification, catalytic mechanisms, activity  
412 regulation, and applications, *Chemical reviews*, 2019.
- 413 34. J. Wu, X. Wang, Q. Wang, Z. Lou, S. Li, Y. Zhu, L. Qin and H. Wei, Nanomaterials with  
414 enzyme-like characteristics (nanozymes): next-generation artificial enzymes (II), *Chemical  
415 Society Reviews*, 2019, **48**, 1004-1076.
- 416 35. P. Rossi, F. Felluga, P. Tecilla, F. Formaggio, M. Crisma, C. Toniolo and P. Scrimin, An  
417 azacrown-functionalized peptide as a metal ion based catalyst for the cleavage of a RNA-  
418 model substrate, *Peptide Science*, 2000, **55**, 496-501.
- 419 36. Yan Xiyun, Nanozyme: a new generation of artificial enzyme, *Progress in Biochemistry and  
420 Biophysics*, 2018, **45**, 101-104.(in Chinese)
- 421 37. F. Natalio, R. André, A. F. Hartog, B. Stoll, K. P. Jochum, R. Wever and W. Tremel, Vanadium  
422 pentoxide nanoparticles mimic vanadium haloperoxidases and thwart biofilm formation,  
423 *Nature nanotechnology*, 2012, **7**, 530.
- 424 38. P. Biparva, S. M. Abedirad and S. Y. Kazemi, ZnO nanoparticles as an oxidase  
425 mimic-mediated flow-injection chemiluminescence system for sensitive determination of  
426 carvedilol, *Talanta*, 2014, **130**, 116-121.
- 427 39. C. Xu and X. Qu, Cerium oxide nanoparticle: a remarkably versatile rare earth nanomaterial  
428 for biological applications, *NPG Asia Materials*, 2014, **6**, e90.
- 429 40. D. Yang, M. Fa, L. Gao, R. Zhao, Y. Luo and X. Yao, The effect of DNA on the oxidase  
430 activity of nanoceria with different morphologies, *Nanotechnology*, 2018, **29**, 385101.
- 431 41. L. Huang, W. Zhang, K. Chen, W. Zhu, X. Liu, R. Wang, X. Zhang, N. Hu, Y. Suo and J.



- 432 Wang, Facet-selective response of trigger molecule to CeO<sub>2</sub> {1 1 0} for up-regulating  
433 oxidase-like activity, *Chemical Engineering Journal*, 2017, **330**, 746-752.
- 434 42. J. Liu, L. Meng, Z. Fei, P. J. Dyson, X. Jing and X. Liu, MnO<sub>2</sub> nanosheets as an artificial  
435 enzyme to mimic oxidase for rapid and sensitive detection of glutathione, *Biosensors and*  
436 *Bioelectronics*, 2017, **90**, 69-74.
- 437 43. X. Liu, Q. Wang, H. Zhao, L. Zhang, Y. Su and Y. Lv, BSA-templated MnO<sub>2</sub> nanoparticles as  
438 both peroxidase and oxidase mimics, *Analyst*, 2012, **137**, 4552-4558.
- 439 44. X. Yan, Y. Song, X. Wu, C. Zhu, X. Su, D. Du and Y. Lin, Oxidase-mimicking activity of  
440 ultrathin MnO<sub>2</sub> nanosheets in colorimetric assay of acetylcholinesterase activity, *Nanoscale*,  
441 2017, **9**, 2317-2323.
- 442 45. N. Singh, M. A. Savanur, S. Srivastava, P. D'Silva and G. Mugesh, A redox modulatory  
443 Mn<sub>3</sub>O<sub>4</sub> nanozyme with multi-enzyme activity provides efficient cytoprotection to human cells  
444 in a Parkinson's disease model, *Angewandte Chemie International Edition*, 2017, **56**,  
445 14267-14271.
- 446 46. X. Zhang and Y. Huang, Evaluation of the antioxidant activity of phenols and tannic acid  
447 determination with Mn<sub>3</sub>O<sub>4</sub> nano-octahedrons as an oxidase mimic, *Analytical Methods*,  
448 2015, **7**, 8640-8646.
- 449 47. H. Li, T. Wang, Y. Wang, S. Wang, P. Su and Y. Yang, Intrinsic triple-enzyme mimetic activity  
450 of V<sub>6</sub>O<sub>13</sub> nanotextiles: mechanism investigation and colorimetric and fluorescent detections,  
451 *Industrial & Engineering Chemistry Research*, 2018, **57**, 2416-2425.
- 452 48. X. Zhang, S. He, Z. Chen and Y. Huang, CoFe<sub>2</sub>O<sub>4</sub> nanoparticles as oxidase mimic-mediated  
453 chemiluminescence of aqueous luminol for sulfite in white wines, *Journal of agricultural and*

- 454 *food chemistry*, 2013, **61**, 840-847.
- 455 49. A. A. Vernekar, T. Das, S. Ghosh and G. Mugesh, A remarkably efficient MnFe<sub>2</sub>O<sub>4</sub>-based  
456 oxidase nanozyme, *Chemistry–An Asian Journal*, 2016, **11**, 72-76.
- 457 50. L. Su, W. Dong, C. Wu, Y. Gong, Y. Zhang, L. Li, G. Mao and S. Feng, The peroxidase and  
458 oxidase-like activity of NiCo<sub>2</sub>O<sub>4</sub> mesoporous spheres: Mechanistic understanding and  
459 colorimetric biosensing, *Analytica chimica acta*, 2017, **951**, 124-132.
- 460 51. Y.-B. Feng, L. Hong, A.-L. Liu, W.-D. Chen, G.-W. Li, W. Chen and X.-H. Xia,  
461 High-efficiency catalytic degradation of phenol based on the peroxidase-like activity of cupric  
462 oxide nanoparticles, *International Journal of Environmental Science and Technology*, 2015,  
463 **12**, 653-660.
- 464 52. K. Korschelt, R. Schwidetzky, F. Pfitzner, J. Strugatchi, C. Schilling, M. von der Au, K.  
465 Kirchhoff, M. Panthöfer, I. Lieberwirth and M. Tahir, CeO<sub>2-x</sub> nanorods with intrinsic  
466 urease-like activity, *Nanoscale*, 2018, **10**, 13074-13082.
- 467 53. J. Fan, J.-J. Yin, B. Ning, X. Wu, Y. Hu, M. Ferrari, G. J. Anderson, J. Wei, Y. Zhao and G.  
468 Nie, Direct evidence for catalase and peroxidase activities of ferritin–platinum nanoparticles,  
469 *Biomaterials*, 2011, **32**, 1611-1618.
- 470 54. X. Shen, W. Liu, X. Gao, Z. Lu, X. Wu and X. Gao, Mechanisms of oxidase and superoxide  
471 dismutation-like activities of gold, silver, platinum, and palladium, and their alloys: A general  
472 way to the activation of molecular oxygen, *Journal of the American Chemical Society*, 2015,  
473 **137**, 15882-15891.
- 474 55. G.-L. Wang, L.-Y. Jin, X.-M. Wu, Y.-M. Dong and Z.-J. Li, Label-free colorimetric sensor for  
475 mercury (II) and DNA on the basis of mercury (II) switched-on the oxidase-mimicking

476 activity of silver nanoclusters, *Analytica chimica acta*, 2015, **871**, 1-8.

477 56. C.-J. Yu, T.-H. Chen, J.-Y. Jiang and W.-L. Tseng, Lysozyme-directed synthesis of platinum  
478 nanoclusters as a mimic oxidase, *Nanoscale*, 2014, **6**, 9618-9624.

479 57. M. Cui, Y. Zhao, C. Wang and Q. Song, The oxidase-like activity of iridium nanoparticles, and  
480 their application to colorimetric determination of dissolved oxygen, *Microchimica Acta*, 2017,  
481 **184**, 3113-3119.

482 58. G.-J. Cao, X. Jiang, H. Zhang, T. R. Croley and J.-J. Yin, Mimicking horseradish peroxidase  
483 and oxidase using ruthenium nanomaterials, *RSC Advances*, 2017, **7**, 52210-52217.

484 59. G.-L. Wang, L.-Y. Jin, Y.-M. Dong, X.-M. Wu and Z.-J. Li, Intrinsic enzyme mimicking  
485 activity of gold nanoclusters upon visible light triggering and its application for colorimetric  
486 trypsin detection, *Biosensors and Bioelectronics*, 2015, **64**, 523-529.

487 60. M. Deng, S. Xu and F. Chen, Enhanced chemiluminescence of the luminol-hydrogen peroxide  
488 system by BSA-stabilized Au nanoclusters as a peroxidase mimic and its application,  
489 *Analytical Methods*, 2014, **6**, 3117-3123.

490 61. Y. Jv, B. Li and R. Cao, Positively-charged gold nanoparticles as peroxidase mimic and their  
491 application in hydrogen peroxide and glucose detection, *Chemical communications*, 2010, **46**,  
492 8017-8019.

493 62. H. Jiang, Z. Chen, H. Cao and Y. Huang, Peroxidase-like activity of chitosan stabilized silver  
494 nanoparticles for visual and colorimetric detection of glucose, *Analyst*, 2012, **137**, 5560-5564.

495 63. L. Jin, Z. Meng, Y. Zhang, S. Cai, Z. Zhang, C. Li, L. Shang and Y. Shen, Ultrasmall Pt  
496 nanoclusters as robust peroxidase mimics for colorimetric detection of glucose in human  
497 serum, *ACS applied materials & interfaces*, 2017, **9**, 10027-10033.

- 498 64. Z. Gao, M. Xu, L. Hou, G. Chen and D. Tang, Irregular-shaped platinum nanoparticles as  
499 peroxidase mimics for highly efficient colorimetric immunoassay, *Analytica chimica acta*,  
500 2013, **776**, 79-86.
- 501 65. J. Lan, W. Xu, Q. Wan, X. Zhang, J. Lin, J. Chen and J. Chen, Colorimetric determination of  
502 sarcosine in urine samples of prostatic carcinoma by mimic enzyme palladium nanoparticles,  
503 *Analytica chimica acta*, 2014, **825**, 63-68.
- 504 66. L. Hu, Y. Yuan, L. Zhang, J. Zhao, S. Majeed and G. Xu, Copper nanoclusters as peroxidase  
505 mimetics and their applications to H<sub>2</sub>O<sub>2</sub> and glucose detection, *Analytica chimica acta*, 2013,  
506 **762**, 83-86.
- 507 67. W. Huang, H. Wu, X. Li and T. Chen, Facile One-Pot Synthesis of Tellurium Nanorods as  
508 Antioxidant and Anticancer Agents, *Chemistry–An Asian Journal*, 2016, **11**, 2301-2311.
- 509 68. F. Wang, E. Ju, Y. Guan, J. Ren and X. Qu, Light-Mediated Reversible Modulation of ROS  
510 Level in Living Cells by Using an Activity-Controllable Nanozyme, *Small*, 2017, **13**,  
511 1603051.
- 512 69. W. He, Y.-T. Zhou, W. G. Wamer, M. D. Boudreau and J.-J. Yin, Mechanisms of the pH  
513 dependent generation of hydroxyl radicals and oxygen induced by Ag nanoparticles,  
514 *Biomaterials*, 2012, **33**, 7547-7555.
- 515 70. J. Gao, B. Jiang, C. Ni, Y. Qi and X. Bi, Enhanced reduction of nitrate by noble metal-free  
516 electrocatalysis on P doped three-dimensional Co<sub>3</sub>O<sub>4</sub> cathode: Mechanism exploration from  
517 both experimental and DFT studies, *Chemical Engineering Journal*, 2019, 123034.
- 518 71. J. Gao, B. Jiang, C. Ni, Y. Qi, Y. Zhang, N. Oturan and M. A. Oturan, Non-precious  
519 Co<sub>3</sub>O<sub>4</sub>-TiO<sub>2</sub>/Ti cathode based electrocatalytic nitrate reduction: Preparation, performance and

520 mechanism, *Applied Catalysis B: Environmental*, 2019, **254**, 391-402.

521 72. S. Chen, Y. Quan, Y.-L. Yu and J.-H. Wang, Graphene quantum dot/silver nanoparticle hybrids  
522 with oxidase activities for antibacterial application, *ACS Biomaterials Science & Engineering*,  
523 2017, **3**, 313-321.

524 73. J. Wang, P. Su, D. Li, T. Wang and Y. Yang, Fabrication of CeO<sub>2</sub>/rGO nanocomposites with  
525 oxidase-like activity and their application in colorimetric sensing of ascorbic acid, *Chemical*  
526 *Research in Chinese Universities*, 2017, **33**, 540-545.

527 74. Y. Guo, Y. Tao, X. Ma, J. Jin, S. Wen, W. Ji, W. Song, B. Zhao and Y. Ozaki, A dual  
528 colorimetric and SERS detection of Hg<sup>2+</sup> based on the stimulus of intrinsic oxidase-like  
529 catalytic activity of Ag-CoFe<sub>2</sub>O<sub>4</sub>/reduced graphene oxide nanocomposites, *Chemical*  
530 *Engineering Journal*, 2018, **350**, 120-130.

531 75. S. Zhang, H. Li, Z. Wang, J. Liu, H. Zhang, B. Wang and Z. Yang, A strongly coupled Au/Fe<sub>3</sub>  
532 O<sub>4</sub>/GO hybrid material with enhanced nanozyme activity for highly sensitive colorimetric  
533 detection, and rapid and efficient removal of Hg<sup>2+</sup> in aqueous solutions, *Nanoscale*, 2015, **7**,  
534 8495-8502.

535 76. M. Vázquez-González, W.-C. Liao, R. m. Cazelles, S. Wang, X. Yu, V. Gutkin and I. Willner,  
536 Mimicking horseradish peroxidase functions using Cu<sup>2+</sup>-modified carbon nitride  
537 nanoparticles or Cu<sup>2+</sup>-modified carbon dots as heterogeneous catalysts, *ACS nano*, 2017, **11**,  
538 3247-3253.

539 77. Y.-l. Dong, H.-g. Zhang, Z. U. Rahman, L. Su, X.-j. Chen, J. Hu and X.-g. Chen, Graphene  
540 oxide-Fe<sub>3</sub>O<sub>4</sub> magnetic nanocomposites with peroxidase-like activity for colorimetric  
541 detection of glucose, *Nanoscale*, 2012, **4**, 3969-3976.

- 542 78. G. Darabdhara, B. Sharma, M. R. Das, R. Boukherroub and S. Szunerits, Cu-Ag bimetallic  
543 nanoparticles on reduced graphene oxide nanosheets as peroxidase mimic for glucose and  
544 ascorbic acid detection, *Sensors and Actuators B: Chemical*, 2017, **238**, 842-851.
- 545 79. E. J. Toone, *Advances in enzymology and related areas of molecular biology: Protein*  
546 *evolution*, John Wiley & Sons, 2010.
- 547 80. M. Comotti, C. Della Pina, R. Matarrese and M. Rossi, The catalytic activity of “naked” gold  
548 particles, *Angewandte Chemie International Edition*, 2004, **43**, 5812-5815.
- 549 81. I. V. Delidovich, B. L. Moroz, O. P. Taran, N. V. Gromov, P. A. Pyrjaev, I. P. Prosvirin, V. I.  
550 Bukhtiyarov and V. N. Parmon, Aerobic selective oxidation of glucose to gluconate catalyzed  
551 by Au/Al<sub>2</sub>O<sub>3</sub> and Au/C: impact of the mass-transfer processes on the overall kinetics,  
552 *Chemical engineering journal*, 2013, **223**, 921-931.
- 553 82. C. Ma, W. Xue, J. Li, W. Xing and Z. Hao, Mesoporous carbon-confined Au catalysts with  
554 superior activity for selective oxidation of glucose to gluconic acid, *Green Chemistry*, 2013,  
555 **15**, 1035-1041.
- 556 83. P. J. Miedziak, H. Alshammari, S. A. Kondrat, T. J. Clarke, T. E. Davies, M. Morad, D. J.  
557 Morgan, D. J. Willock, D. W. Knight and S. H. Taylor, Base-free glucose oxidation using air  
558 with supported gold catalysts, *Green Chemistry*, 2014, **16**, 3132-3141.
- 559 84. K. Odrozek, K. Maresz, A. Koreniuk, K. Prusik and J. Mrowiec-Białoń, Amine-stabilized  
560 small gold nanoparticles supported on AISBA-15 as effective catalysts for aerobic glucose  
561 oxidation, *Applied Catalysis A: General*, 2014, **475**, 203-210.
- 562 85. Y. Wang, S. Van de Vyver, K. K. Sharma and Y. Román-Leshkov, Insights into the stability of  
563 gold nanoparticles supported on metal oxides for the base-free oxidation of glucose to

- 564 gluconic acid, *Green Chemistry*, 2014, **16**, 719-726.
- 565 86. M. Comotti, C. Della Pina, E. Falletta and M. Rossi, Aerobic oxidation of glucose with gold  
566 catalyst: hydrogen peroxide as intermediate and reagent, *Advanced Synthesis & Catalysis*,  
567 2006, **348**, 313-316.
- 568 87. H. Liang, F. Lin, Z. Zhang, B. Liu, S. Jiang, Q. Yuan and J. Liu, Multicopper laccase  
569 mimicking nanozymes with nucleotides as ligands, *ACS applied materials & interfaces*, 2017,  
570 **9**, 1352-1360.
- 571 88. M. Chen, Z. Wang, J. Shu, X. Jiang, W. Wang, Z.-H. Shi and Y.-W. Lin, Mimicking a natural  
572 enzyme system: cytochrome c oxidase-like activity of Cu<sub>2</sub>O nanoparticles by receiving  
573 electrons from cytochrome c, *Inorganic chemistry*, 2017, **56**, 9400-9403.
- 574 89. F. Passardi, G. Theiler, M. Zamocky, C. Cosio, N. Rouhier, F. Teixeira, M. Margis-Pinheiro, V.  
575 Ioannidis, C. Penel and L. Falquet, PeroxiBase: the peroxidase database, *Phytochemistry*,  
576 2007, **68**, 1605-1611.
- 577 90. H. Wei and E. Wang, Fe<sub>3</sub>O<sub>4</sub> magnetic nanoparticles as peroxidase mimetics and their  
578 applications in H<sub>2</sub>O<sub>2</sub> and glucose detection, *Analytical Chemistry*, 2008, **80**, 2250-2254.
- 579 91. R. Cheng, G.-q. Li, C. Cheng, L. Shi, X. Zheng and Z. Ma, Catalytic oxidation of  
580 4-chlorophenol with magnetic Fe<sub>3</sub>O<sub>4</sub> nanoparticles: mechanisms and particle transformation,  
581 *RSC Advances*, 2015, **5**, 66927-66933.
- 582 92. Y.-h. Wu, L. Chu, W. Liu, L. Jiang, X.-y. Chen, Y.-h. Wang and Y.-l. Zhao, The screening of  
583 metal ion inhibitors for glucose oxidase based on the peroxidase-like activity of nano-Fe<sub>3</sub>O<sub>4</sub>,  
584 *RSC Advances*, 2017, **7**, 47309-47315.
- 585 93. A. Roy, R. Sahoo, C. Ray, S. Dutta and T. Pal, Soft template induced phase selective synthesis

586 of Fe<sub>2</sub>O<sub>3</sub> nanomagnets: one step towards peroxidase-mimic activity allowing colorimetric  
587 sensing of thioglycolic acid, *RSC Advances*, 2016, **6**, 32308-32318.

588 94. M. Kluncker, M. Nawaz Tahir, R. Ragg, K. Korschelt, P. Simon, T. E. Gorelik, B. Barton, S. I.  
589 Shylin, M. Panthöfer and J. Herzberger, Pd@ Fe<sub>2</sub>O<sub>3</sub> superparticles with enhanced peroxidase  
590 activity by solution phase epitaxial growth, *Chemistry of Materials*, 2017, **29**, 1134-1146.

591 95. T. Zhang, C. Cao, X. Tang, Y. Cai, C. Yang and Y. Pan, Enhanced peroxidase activity and  
592 tumour tissue visualization by cobalt-doped magnetoferritin nanoparticles, *Nanotechnology*,  
593 2016, **28**, 045704.

594 96. P. Chelikani, I. Fita and P. C. Loewen, Diversity of structures and properties among catalases,  
595 *Cellular and Molecular Life Sciences CMLS*, 2004, **61**, 192-208.

596 97. T. Pirmohamed, J. M. Dowding, S. Singh, B. Wasserman, E. Heckert, A. S. Karakoti, J. E.  
597 King, S. Seal and W. T. Self, Nanoceria exhibit redox state-dependent catalase mimetic  
598 activity, *Chemical communications*, 2010, **46**, 2736-2738.

599 98. I. Celardo, J. Z. Pedersen, E. Traversa and L. Ghibelli, Pharmacological potential of cerium  
600 oxide nanoparticles, *Nanoscale*, 2011, **3**, 1411-1420.

601 99. M. Hayyan, M. A. Hashim and I. M. AlNashef, Superoxide ion: generation and chemical  
602 implications, *Chemical reviews*, 2016, **116**, 3029-3085.

603 100. A. Okado-Matsumoto and I. Fridovich, Subcellular distribution of superoxide dismutases  
604 (SOD) in rat liver Cu, Zn-SOD in mitochondria, *Journal of Biological Chemistry*, 2001, **276**,  
605 38388-38393.

606 101. T. Yan, Q. Zhi-Yue, X. Zhuo-Bin and G. Li-Zeng, Antibacterial Mechanism and Applications  
607 of Nanozymes, *Progress in Biochemistry & Biophysics*, 2018, **45**, 118-128.



- 608 102. L. Järup, Hazards of heavy metal contamination, *British medical bulletin*, 2003, **68**, 167-182.
- 609 103. M. K. Raikwar, P. Kumar, M. Singh and A. Singh, Toxic effect of heavy metals in livestock  
610 health, *Veterinary world*, 2008, **1**, 28.
- 611 104. J. Hu, Q. Zhuang, Y. Wang and Y. Ni, Label-free fluorescent catalytic biosensor for highly  
612 sensitive and selective detection of the ferrous ion in water samples using a layered  
613 molybdenum disulfide nanozyme coupled with an advanced chemometric model, *Analyst*,  
614 2016, **141**, 1822-1829.
- 615 105. W. Li, G.-C. Fan, F. Gao, Y. Cui, W. Wang and X. Luo, High-activity Fe<sub>3</sub>O<sub>4</sub> nanozyme as  
616 signal amplifier: A simple, low-cost but efficient strategy for ultrasensitive  
617 photoelectrochemical immunoassay, *Biosensors and Bioelectronics*, 2019, **127**, 64-71.
- 618 106. Z. Chen, C. Zhang, Q. Gao, G. Wang, L. Tan and Q. Liao, Colorimetric signal amplification  
619 assay for mercury ions based on the catalysis of gold amalgam, *Analytical chemistry*, 2015, **87**,  
620 10963-10968.
- 621 107. W. Li, B. Chen, H. Zhang, Y. Sun, J. Wang, J. Zhang and Y. Fu, BSA-stabilized Pt nanozyme  
622 for peroxidase mimetics and its application on colorimetric detection of mercury (II) ions,  
623 *Biosensors and Bioelectronics*, 2015, **66**, 251-258.
- 624 108. C.-W. Lien, B. Unnikrishnan, S. G. Harroun, C.-M. Wang, J.-Y. Chang, H.-T. Chang and C.-C.  
625 Huang, Visual detection of cyanide ions by membrane-based nanozyme assay, *Biosensors and*  
626 *Bioelectronics*, 2018, **102**, 510-517.
- 627 109. K. C. Jones and P. De Voogt, Persistent organic pollutants (POPs): state of the science,  
628 *Environmental pollution*, 1999, **100**, 209-221.
- 629 110. F. Wania and D. Mackay, Peer reviewed: tracking the distribution of persistent organic

630 pollutants, *Environmental science & technology*, 1996, **30**, 390A-396A.

631 111. C. Comninellis, Electrocatalysis in the electrochemical conversion/combustion of organic  
632 pollutants for waste water treatment, *Electrochimica Acta*, 1994, **39**, 1857-1862.

633 112. S. Singh, P. Tripathi, N. Kumar and S. Nara, Colorimetric sensing of malathion using  
634 palladium-gold bimetallic nanozyme, *Biosensors and Bioelectronics*, 2017, **92**, 280-286.

635 113. N. Desneux, A. Decourtye and J.-M. Delpuech, The sublethal effects of pesticides on  
636 beneficial arthropods, *Annu. Rev. Entomol.*, 2007, **52**, 81-106.

637 114. B. A. Croft, *Arthropod biological control agents and pesticides*, John Wiley and Sons Inc.,  
638 1990.

639 115. R. Zeng, Z. Luo, L. Zhang and D. Tang, Platinum nanozyme-catalyzed gas generation for  
640 pressure-based bioassay using polyaniline nanowires-functionalized graphene oxide  
641 framework, *Analytical chemistry*, 2018, **90**, 12299-12306.

642 116. P. S. Stewart and J. W. Costerton, Antibiotic resistance of bacteria in biofilms, *The lancet*,  
643 2001, **358**, 135-138.

644 117. F. Tian, J. Zhou, B. Jiao and Y. He, Nanozyme-based cascade colorimetric aptasensor for  
645 amplified detection of ochratoxin A, *Nanoscale*, 2019.

646 118. K. Van der Merwe, P. Steyn, L. Fourie, D. B. Scott and J. Theron, Ochratoxin A, a toxic  
647 metabolite produced by *Aspergillus ochraceus* Wilh, *Nature*, 1965, **205**, 1112.

648 119. A. P. Magiorakos, A. Srinivasan, R. Carey, Y. Carmeli, M. Falagas, C. Giske, S. Harbarth, J.  
649 Hindler, G. Kahlmeter and B. Olsson-Liljequist, Multidrug-resistant, extensively drug-  
650 resistant and pandrug-resistant bacteria: an international expert proposal for interim standard  
651 definitions for acquired resistance, *Clinical microbiology and infection*, 2012, **18**, 268-281.

- 652 120. M. Rai, S. Deshmukh, A. Ingle and A. Gade, Silver nanoparticles: the powerful nanoweapon  
653 against multidrug-resistant bacteria, *Journal of applied microbiology*, 2012, **112**, 841-852.
- 654 121. B. P. Yu, Cellular defenses against damage from reactive oxygen species, *Physiological*  
655 *reviews*, 1994, **74**, 139-162.
- 656 122. S. Bhattacharya, in *Free radicals in human health and disease*, Springer, 2015, pp. 17-29.
- 657 123. Z. Wang, K. Dong, Z. Liu, Y. Zhang, Z. Chen, H. Sun, J. Ren and X. Qu, Activation of  
658 biologically relevant levels of reactive oxygen species by Au/g-C<sub>3</sub>N<sub>4</sub> hybrid nanozyme for  
659 bacteria killing and wound disinfection, *Biomaterials*, 2017, **113**, 145-157.
- 660 124. E. Cabisco Català, J. Tamarit Sumalla and J. Ros Salvador, Oxidative stress in bacteria and  
661 protein damage by reactive oxygen species, *International Microbiology*, 2000, vol. 3, núm. 1,  
662 p. 3-8, 2000.
- 663 125. O. Choi and Z. Hu, Size dependent and reactive oxygen species related nanosilver toxicity to  
664 nitrifying bacteria, *Environmental science & technology*, 2008, **42**, 4583-4588.
- 665 126. K. Apel and H. Hirt, Reactive oxygen species: metabolism, oxidative stress, and signal  
666 transduction, *Annu. Rev. Plant Biol.*, 2004, **55**, 373-399.
- 667 127. M. P. Murphy, How mitochondria produce reactive oxygen species, *Biochemical journal*, 2009,  
668 **417**, 1-13.
- 669 128. M. L. Circu and T. Y. Aw, Reactive oxygen species, cellular redox systems, and apoptosis,  
670 *Free Radical Biology and Medicine*, 2010, **48**, 749-762.
- 671 129. J. Nordberg and E. S. Arnér, Reactive oxygen species, antioxidants, and the mammalian  
672 thioredoxin system, *Free radical biology and medicine*, 2001, **31**, 1287-1312.
- 673 130. K. Xu, D. Luan, X. Wang, B. Hu, X. Liu, F. Kong and B. Tang, An Ultrasensitive Cyclization-

674 Based Fluorescent Probe for Imaging Native HOBr in Live Cells and Zebrafish, *Angewandte*  
675 *Chemie International Edition*, 2016, **55**, 12751-12754.

676 131. R. A. Damodar, S.-J. You and H.-H. Chou, Study the self cleaning, antibacterial and  
677 photocatalytic properties of TiO<sub>2</sub> entrapped PVDF membranes, *Journal of hazardous*  
678 *materials*, 2009, **172**, 1321-1328.

679 132. T. Paul, M. C. Dodd and T. J. Strathmann, Photolytic and photocatalytic decomposition of  
680 aqueous ciprofloxacin: transformation products and residual antibacterial activity, *Water*  
681 *research*, 2010, **44**, 3121-3132.

682 133. J. Niu, Y. Sun, F. Wang, C. Zhao, J. Ren and X. Qu, Photomodulated nanozyme used for a  
683 Gram-selective antimicrobial, *Chemistry of Materials*, 2018, **30**, 7027-7033.

684 134. J. Zhang, X. Lu, D. Tang, S. Wu, X. Hou, J. Liu and P. Wu, Phosphorescent carbon dots for  
685 highly efficient oxygen photosensitization and as photo-oxidative nanozymes, *ACS applied*  
686 *materials & interfaces*, 2018, **10**, 40808-40814.

687 135. L. Gao, Y. Liu, D. Kim, Y. Li, G. Hwang, P. C. Naha, D. P. Cormode and H. Koo,  
688 Nanocatalysts promote *Streptococcus mutans* biofilm matrix degradation and enhance  
689 bacterial killing to suppress dental caries in vivo, *Biomaterials*, 2016, **101**, 272-284.

690 136. P. C. Naha, Y. Liu, G. Hwang, Y. Huang, S. Gubara, V. Jonnakuti, A. Simon-Soro, D. Kim, L.  
691 Gao and H. Koo, Dextran-Coated Iron Oxide Nanoparticles as Biomimetic Catalysts for  
692 Localized and pH-Activated Biofilm Disruption, *ACS nano*, 2019.

693 137. D. Das, B. C. Nath, P. Phukon and S. K. Dolui, Synthesis and evaluation of antioxidant and  
694 antibacterial behavior of CuO nanoparticles, *Colloids and Surfaces B: Biointerfaces*, 2013,  
695 **101**, 430-433.

- 696 138. Y. Ye, L. Xiao, B. He, Q. Zhang, T. Nie, X. Yang, D. Wu, H. Cheng, P. Li and Q. Wang,  
697 Oxygen-tuned nanozyme polymerization for the preparation of hydrogels with printable and  
698 antibacterial properties, *Journal of Materials Chemistry B*, 2017, **5**, 1518-1524.
- 699 139. M. N. Karim, M. Singh, P. Weerathunge, P. Bian, R. Zheng, C. Dekiwadia, T. Ahmed, S. Walia,  
700 E. Della Gaspera and S. Singh, Visible-Light-Triggered Reactive-Oxygen-Species-Mediated  
701 Antibacterial Activity of Peroxidase-Mimic CuO Nanorods, *ACS Applied Nano Materials*,  
702 2018, **1**, 1694-1704.
- 703 140. M. T. Matter, L. A. Furer, F. H. Starsich, G. Fortunato, S. E. Pratsinis and I. K. Herrmann,  
704 Engineering the Bioactivity of Flame-Made Ceria and Ceria/Bioglass Hybrid Nanoparticles,  
705 *ACS applied materials & interfaces*, 2018, **11**, 2830-2839.
- 706 141. M. Hu, K. Korschelt, M. Viel, N. Wiesmann, M. Kappl, J. r. Brieger, K. Landfester, H.  
707 Thérien-Aubin and W. Tremel, Nanozymes in Nanofibrous Mats with Haloperoxidase-like  
708 Activity To Combat Biofouling, *ACS applied materials & interfaces*, 2018, **10**, 44722-44730.
- 709 142. Z.-Y. Yang, S.-L. Luo, H. Li, S.-W. Dong, J. He, H. Jiang, R. Li and X.-C. Yang, Alendronate  
710 as a robust anchor for ceria nanoparticle surface coating: facile binding and improved  
711 biological properties, *RSC Advances*, 2014, **4**, 59965-59969.
- 712 143. Z. Chen, Z. Wang, J. Ren and X. Qu, Enzyme mimicry for combating bacteria and biofilms,  
713 *Accounts of chemical research*, 2018, **51**, 789-799.
- 714 144. T. Qin, R. Ma, Y. Yin, X. Miao, S. Chen, K. Fan, J. Xi, Q. Liu, Y. Gu and Y. Yin, Catalytic  
715 inactivation of influenza virus by iron oxide nanozyme, *Theranostics*, 2019, **9**, 6920.
- 716 145. C. S. Turchi and D. F. Ollis, Photocatalytic degradation of organic water contaminants:  
717 mechanisms involving hydroxyl radical attack, *Journal of catalysis*, 1990, **122**, 178-192.

- 718 146. R. J. Tayade, P. K. Surolia, R. G. Kulkarni and R. V. Jasra, Photocatalytic degradation of dyes  
719 and organic contaminants in water using nanocrystalline anatase and rutile TiO<sub>2</sub>, *Science and*  
720 *Technology of Advanced Materials*, 2007, **8**, 455.
- 721 147. D. R. Lovley, Bioremediation of organic and metal contaminants with dissimilatory metal  
722 reduction, *Journal of industrial microbiology*, 1995, **14**, 85-93.
- 723 148. L. N. Nguyen, F. I. Hai, W. E. Price, F. D. Leusch, F. Roddick, H. H. Ngo, W. Guo, S. F.  
724 Magram and L. D. Nghiem, The effects of mediator and granular activated carbon addition on  
725 degradation of trace organic contaminants by an enzymatic membrane reactor, *Bioresource*  
726 *technology*, 2014, **167**, 169-177.
- 727 149. J. Jiang, C. He, S. Wang, H. Jiang, J. Li and L. Li, Recyclable ferromagnetic chitosan  
728 nanozyme for decomposing phenol, *Carbohydrate polymers*, 2018, **198**, 348-353.
- 729 150. A. C. Grabski, H. J. Grimek and R. R. Burgess, Immobilization of manganese peroxidase from  
730 *Lentinula edodes* and its biocatalytic generation of MnIII-chelate as a chemical oxidant of  
731 chlorophenols, *Biotechnology and bioengineering*, 1998, **60**, 204-215.
- 732 151. M. Fujihira, Y. Satoh and T. Osa, Heterogeneous photocatalytic oxidation of aromatic  
733 compounds on TiO<sub>2</sub>, *Nature*, 1981, **293**, 206.
- 734 152. M. Blumer, Polycyclic aromatic compounds in nature, *Scientific American*, 1976, **234**, 34-45.
- 735 153. T. Johjima, N. Itoh, M. Kabuto, F. Tokimura, T. Nakagawa, H. Wariishi and H. Tanaka, Direct  
736 interaction of lignin and lignin peroxidase from *Phanerochaete chrysosporium*, *Proceedings of*  
737 *the National Academy of Sciences*, 1999, **96**, 1989-1994.
- 738 154. N. Duran and E. Esposito, Potential applications of oxidative enzymes and phenoloxidase-like  
739 compounds in wastewater and soil treatment: a review, *Applied catalysis B: environmental*,

740 2000, **28**, 83-99.

741 155. B. Padhi, Pollution due to synthetic dyes toxicity & carcinogenicity studies and remediation,  
742 *International Journal of Environmental Sciences*, 2012, **3**, 940.

743 156. Q. Zhou, Chemical pollution and transport of organic dyes in water–soil–crop systems of the  
744 Chinese Coast, *Bulletin of environmental contamination and toxicology*, 2001, **66**, 784-793.

745 157. R. Saravanan, S. Karthikeyan, V. Gupta, G. Sekaran, V. Narayanan and A. Stephen, Enhanced  
746 photocatalytic activity of ZnO/CuO nanocomposite for the degradation of textile dye on  
747 visible light illumination, *Materials Science and Engineering: C*, 2013, **33**, 91-98.

748 158. Z. Du, C. Feng, Q. Li, Y. Zhao and X. Tai, Photodegradation of NPE-10 surfactant by  
749 Au-doped nano-TiO<sub>2</sub>, *Colloids and Surfaces A: Physicochemical and Engineering Aspects*,  
750 2008, **315**, 254-258.

751 159. Yuqun Wang, Xiangyu Jia and Yunlong Zhang, Nanozyme air materials and their applications  
752 in air purifiers, *Jiangsu Architecture*, 2005, **4**, 36-38. (in Chinese)

753 160. Zhang Qingsong, Wang Yuqun, Jia Xiangyi and Zhang Yunlong, Research on air purification  
754 equipment of central air conditioning and ventilation system, *Jiangsu Architecture*, 2006, **6**,  
755 73-76. (in Chinese)

756 161. Q. Wang, X. Zhang, L. Huang, Z. Zhang and S. Dong, One-pot synthesis of Fe<sub>3</sub>O<sub>4</sub>  
757 nanoparticle loaded 3D porous graphene nanocomposites with enhanced nanozyme activity  
758 for glucose detection, *ACS applied materials & interfaces*, 2017, **9**, 7465-7471.

759 162. S. Li, X. Liu, H. Chai and Y. Huang, Recent advances in the construction and analytical  
760 applications of metal-organic frameworks-based nanozymes, *TrAC Trends in Analytical*  
761 *Chemistry*, 2018, **105**, 391-403.

