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Interactions between 2′-fluoro-(carbamoylpyridinyl)deschloroepibatidine analogues and acetylcholine-binding protein inform on potent antagonist activity against nicotinic receptors

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Low-nanomolar binding constants were recorded for a series of six 2′-fluoro-(carbamoylpyridinyl)deschloroepibatidine analogues with acetylcholine-binding protein (AChBP). The crystal structures of three complexes with AChBP reveal details of molecular recognition in the orthosteric binding site and imply how the other three ligands bind. Comparisons exploiting AChBP as a surrogate for α4β2 and α7 nicotinic acetylcholine receptors (nAChRs) suggest that the key interactions are conserved. The ligands interact with the same residues as the archetypal nAChR agonist nicotine yet display greater affinity, thereby rationalizing their in vivo activity as potent antagonists of nicotine-induced antinociception. An oxyanion-binding site is formed on the periphery of the AChBP orthosteric site by Lys42, Asp94, Glu170 and Glu210. These residues are highly conserved in the human α4, β2 and α7 nAChR sequences. However, specific sequence differences are discussed that could contribute to nAChR subtype selectivity and in addition may represent a point of allosteric modulation. The ability to engage with this peripheral site may explain, in part, the function of a subset of ligands to act as agonists of α7 nAChR.

1. Introduction

Nicotinic acetylcholine receptors (nAChRs) are cation-selective pentameric ligand-gated ion channels (pLGICs) gated by the neurotransmitters acetylcholine and choline. They are also the targets of non-endogenous molecules, with the best known being the archetypal agonist nicotine. The availability of a variety of subunits enables neuronal nAChRs to assemble in different homopentameric or heteropentameric combinations (Gotti et al., 2009; Taly et al., 2009; Bertrand & Terry, 2018). The most common types in the mammalian central nervous system are an α4β2 combination, which has a high affinity for nicotine, and the α7 homomer (Taly et al., 2009). The pLGIC family presents a number of therapeutic targets for neurological conditions; specifically, nAChR subtypes are key targets for the development of compounds with use in the treatment of nicotine addiction and also of pain (Bertrand et al., 2015; Dineley et al., 2015; Bagdas et al., 2018). The discovery of epibatidine [(1R,2R,4S)-2-(6-chloro-3-pyridinyl)-7-azabicyclo[2.2.1]heptane; Fig. 1], a highly potent but relatively nonselective agonist of nAChR that displays powerful non-opiate-mediated antinociceptive effects, elicited great excitement (Spande et al., 1992; Traynor, 1998). A serious liability due to toxicity rules out therapeutic use; nevertheless, with high ligand efficiency and potency the compound has provided a basis for the development of...
anallogues that target nAChR (Spang et al., 2000; Carroll, 2004; Mu et al., 2006; Ondachi et al., 2016). One study focused on the position of the chloropyridine ring N atom and resulted in conversion from an agonist to an antagonist profile (Spang et al., 2000).

Our interest centres on a series of 2'-fluoro-(carbamoyl-pyridinyl)deschloroepibatidine analogues (compounds 1–6; Fig. 1), characterized as having high affinity (Ki < 1 nM) for α4β2 nAChR, that display differing degrees of subtype selectivity and novel pharmacological effects compared with epibatidine (Ondachi et al., 2016). Electrophysiological measurements of ion-channel activity indicated that compounds 1–6 have little or no agonist activity on recombinant α4β2 or α3β4 nAChRs. In contrast, the parent compound epibatidine is a full agonist. The compound set demonstrated antagonist activity on α4β2 and α3β4 nAChRs. However, puzzling effects were observed on the homomeric α7 nAChR. Compounds 1 and 3 act as mixed partial agonists of α4β2, whilst compound 2 is only an antagonist of α7 nAChR, with no agonist properties detected. Furthermore, compounds 4 and 5 were selective for α4β2 over α7, and in vivo studies indicated that these analogues all antagonize the antinociceptive action of nicotine with, in the case of compound 2, a potency approaching that of varenicline, a well studied partial agonist of α4β2 receptors and a full agonist of α7 nAChR (Ondachi et al., 2016). The differing actions at nAChR subtypes are perplexing and we sought to investigate further.

The biological target nAChRs follow the standard structural arrangement of pLGICs, with five subunits creating a central ion pore. Each subunit possesses an extracellular domain (ECD) followed by four transmembrane α-helices and intracellular contributions from the inter-helical segments (Taly et al., 2009; Bertrand & Terry, 2018). The orthosteric binding site, where agonists and competitive antagonists bind, is created by contributions from the ECD at the interface between two subunits. One subunit contributes the principal (+) face, which is formed of three loops known as A, B and C. On an adjacent subunit, loops D, E, F and G comprise the complementary (−) side of the binding site (Sixma & Smit, 2003; Corringer et al., 2012; daCosta & Baenziger, 2013; Nys et al., 2013; Sauguet et al., 2015). Acetylcholine-binding protein, which is found in the cholinergic synapse of gastropods including Aplysia californica (AcAChBP), shares 20–25% sequence identity with the ECD of nAChR sequences. In AcAChBP, 44 residues are involved in the orthosteric binding site and the identities of these residues in human nAChRs range from 32% (β2) to 45% (α4 and α7). In addition, AChBP and nAChR display closely related structures and similar ligand-binding properties (Celie et al., 2004; Hansen et al., 2005; Lemoine et al., 2012; Rucktooa et al., 2012; Shahsavari et al., 2016). Despite the hugely impressive developments in studies of membrane-bound pLGIC forms, for example the cryo-EM structure of the heteromeric human α4β2 nAChR (Walsh et al., 2018), AcAChBP remains a valued surrogate for the study of ligand–receptor interactions. This is due to the convenience of working with a stable, soluble protein for which efficient purification protocols exist and where ordered single crystals can be obtained.

We sought to exploit AcAChBP in this manner to investigate the interactions and affinities of epibatidine analogues 1–6 (Fig. 1). Biolayer interferometry (BLI) provided Ki values, and the crystal structures of three complexes revealed the mode of binding and interactions of compounds 1–3, providing models for complexes with compounds 4–6. Comparisons and modelling using nAChR sequences and structures allowed us to consider the bioactivity of the compound series and to comment on structure–function relationships.

![Figure 1](image_url)

The structures of epibatidine, nicotine and analogues 1–6 used in this work. In all cases the protonated form, which is most likely under physiological conditions, is shown. The first aromatic ring after the azabicycloheptane (a fluoropyridine) is labelled A and the second ring labelled B is a benzylcarboxamide in compound 1 and a pyridinecarboxamide in compounds 2–6.
that may be exploited in the design of new nAChR ligands as chemical tools and/or with therapeutic potential.

2. Materials and methods
2.1. Recombinant protein production

A gene encoding AcAChBP with a C-terminal His<sub>6</sub> tag was cloned into the pFastBac1 vector (Thermo Fisher). The amino-acid sequence was derived from the <i>A. californica</i> genome (https://www.broadinstitute.org/aplysia/aplysia-genome-project) and is similar to UniProt entry Q8WSF8 except that two alanine residues are replaced by Val<sub>60</sub> and Val<sub>155</sub>. This construct was expressed using the Bac-to-Bac system (Thermo Fisher) in <i>Spodoptera frugiperda</i> (Sf<sub>9</sub>) cells maintained in shaker flasks at 27°C using Insect-XPRESS medium (Lonza) supplemented with 2 mM t-glutamine and 100 U ml<sup>-1</sup> penicillin/streptomycin (Thermo Fisher). The AcAChBP baculovirus was generated through transfection of 500 ng bacmid medium via centrifugation (1000 g) followed by the removal of non-immobilized protein (600 s) and baseline measurements were obtained at 25°C for six concentrations of each compound (1.7–410 nM) diluted in buffer A plus 1% DMSO. BLI assays were conducted with a baseline measurement in buffer (60 s), an association measurement in the well containing the compound (120 s) and a dissociation measurement in buffer (120 s). Data processing and analysis were performed with Octet RED Data Analysis version 7.1 (Forté-Bio), where the background response of immobilized AcAChBP in buffer was subtracted.

2.4. Co-crystallization of AcAChBP with compounds

AcAChBP (4 mg ml<sup>-1</sup> in buffer A) was incubated with 2 mM of compounds 1–6 for 1 h at room temperature and this mixture was then used in crystallization trials. Compounds 4–6 did not yield suitable crystals. Small well formed crystals of AcAChBP with compounds 1–3 were obtained using hanging drops consisting of 2 μl complex solution plus 1 μl reservoir solution equilibrated against 800 μl reservoir solution for 24 h at 18°C. The reservoirs consisted of 0.2 M NaCl, 0.1 M phosphate–citrate pH 4.2 and PEG 8000 at 8% (compound 2), 10% (compound 3) or 12% (compound 1). These conditions allowed us to prepare microseed stocks by transferring crystals to microcentrifuge tubes containing the appropriate reservoir solution, 40% glycerol and a Secd Bead (Hampton Research), and vortexing the sample for 1 min. A cleaned human eyelash was dipped into the seed stock and then passed through freshly assembled crystallization drops. Well ordered multi-faced prisms, with maximum dimensions of about 20 μm, appeared after four days.

2.5. Crystallographic analyses

Crystals were harvested using a nylon loop, cryoprotected with reservoir solution adjusted to contain 20% ethane-1,2-diol and then flash-cooled in liquid nitrogen. Diffraction was recorded on Diamond Light Source (DLS) microfocus beamline I24 using a PILATUS 6M-F detector (Dectris) and the images were indexed and integrated using XDS (Kabsch, 2010). The data were scaled using AIMLESS (Evans & Murshudov, 2013) from the CCP4 suite (Winn et al., 2011) and the structures were solved by molecular replacement with Phaser (McCoy et al., 2007), utilizing wild-type protein in complex with strychnine at 1.91 Å resolution (PDB entry 2xys; Brams et al., 2011) as the model for AcAChBP–1 and the complex with nicotine at 2.20 Å resolution (PDB entry 5o87; Dawson et al., 2019) as that for AcAChBP–2 and AcAChBP–3. Ligand models and restraints were generated with the grade server (Global Phasing: http://grade.globalphasing.org/cgi-bin/grade/server.cgi). Multiple rounds of automated restrained refinement were completed using REFMAC5 (Murshudov et al., 2011), with manual refinement and model building in Coot (Emsley et al., 2010). The epibatidine analogues were well defined in electron and difference density maps (Supplementary Fig. S1). Asn91 is glycosylated and N-acetyl-d-glucosamine (NAG) was modelled onto several subunits. It became clear that additional ligands were present and these were assigned and refined satisfactorily as phosphate, ethane-1,2-diol and oxalate. The latter is likely to be present as a contaminant in PEG 8000 (Fyfe et al., 2010). Strict noncrystallographic symmetry restraints were applied throughout to
AcChBP-2 and AcChBP-3 but were relaxed towards the end of the refinement for AcChBP-1. Ligand 3D solvent-accessible surface areas were calculated using AREAIMOL, with a probe solvent radius of 1.4 Å and 100 surface points per Å². Omit maps were generated by removal of the ligands and bulk-solvent corrections before recalculating the \( F_o - F_c \) maps presented in Supplementary Fig. S1. Graphics were rendered using the PyMOL molecular-graphics system (Schrödinger). Crystallographic statistics are presented in Table 1. The amino-acid sequences used include human \( \alpha 4 \) (UniProt code P43681), \( \alpha 7 \) (P36544) and \( \alpha 2 \) (P17787) nAChR subtypes.

### Table 1
Crystalllographic statistics for the AcChBP-ligand complexes.

<table>
<thead>
<tr>
<th>Ligand</th>
<th>AcChBP-1</th>
<th>AcChBP-2</th>
<th>AcChBP-3</th>
</tr>
</thead>
<tbody>
<tr>
<td>PDB code</td>
<td>6qkk</td>
<td>6qqp</td>
<td>6qqo</td>
</tr>
<tr>
<td>Data collection</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>( a, b, c ) (Å)</td>
<td>211.0, 131.6, 131.8</td>
<td>209.5, 136.9, 131.5</td>
<td>209.5, 136.9, 131.5</td>
</tr>
<tr>
<td>Space group</td>
<td>C2</td>
<td>C2</td>
<td>C2</td>
</tr>
<tr>
<td>Source</td>
<td>DLS microfocus beamline I24</td>
<td>DLS microfocus beamline I24</td>
<td>DLS microfocus beamline I24</td>
</tr>
<tr>
<td>Wavelength (Å)</td>
<td>0.96858</td>
<td>0.96858</td>
<td>0.96858</td>
</tr>
<tr>
<td>Subunits per asymmetric unit</td>
<td>10</td>
<td>10</td>
<td>10</td>
</tr>
<tr>
<td>Resolution range (Å)</td>
<td>48.83–2.20 (2.24–2.20)</td>
<td>46.82–2.40 (2.44–2.40)</td>
<td>46.40–2.50 (2.54–2.50)</td>
</tr>
<tr>
<td>Other ligands</td>
<td>NAG, ethane-1,2-diol, phosphate, oxalate</td>
<td>NAG, ethane-1,2-diol, phosphate</td>
<td>NAG, ethane-1,2-diol, phosphate</td>
</tr>
<tr>
<td>Total No. of reflections</td>
<td>611399 (30075)</td>
<td>405440 (19537)</td>
<td>254487 (12673)</td>
</tr>
<tr>
<td>Unique reflections</td>
<td>177603 (8766)</td>
<td>139199 (6884)</td>
<td>109552 (5537)</td>
</tr>
<tr>
<td>Multiplicity</td>
<td>3.4 (3.4)</td>
<td>2.9 (2.8)</td>
<td>2.3 (2.3)</td>
</tr>
<tr>
<td>( R_{merge} )</td>
<td>0.172 (0.888)</td>
<td>0.089 (0.510)</td>
<td>0.118 (0.559)</td>
</tr>
<tr>
<td>( R_{fref} )</td>
<td>0.162 (0.831)</td>
<td>0.088 (0.497)</td>
<td>0.114 (0.539)</td>
</tr>
<tr>
<td>Wilson B factor (Å²)</td>
<td>15.5</td>
<td>29.0</td>
<td>13.6</td>
</tr>
<tr>
<td>Completeness (%)</td>
<td>99.9 (98.4)</td>
<td>98.7 (98.7)</td>
<td>89.7 (91.8)</td>
</tr>
<tr>
<td>( I/\sigma(I) )</td>
<td>6.6 (2.0)</td>
<td>8.2 (2.5)</td>
<td>6.9 (2.6)</td>
</tr>
<tr>
<td>CC&lt;sub&gt;1/2&lt;/sub&gt;</td>
<td>0.95 (0.43)</td>
<td>0.95 (0.65)</td>
<td>0.93 (0.46)</td>
</tr>
<tr>
<td>Refinement</td>
<td>( R_{work}/R_{free} )</td>
<td>0.1990/0.228</td>
<td>0.1880/0.210</td>
</tr>
<tr>
<td>No. of reflections for ( R_{work}/R_{free} )</td>
<td>168732/8867</td>
<td>131926/6903</td>
<td>104105/5421</td>
</tr>
<tr>
<td>Protein residues</td>
<td>2065</td>
<td>2054</td>
<td>2053</td>
</tr>
<tr>
<td>No. of ligands</td>
<td>10</td>
<td>10</td>
<td>10</td>
</tr>
<tr>
<td>No. of water molecules</td>
<td>1287</td>
<td>924</td>
<td>659</td>
</tr>
<tr>
<td>R.m.s.d.s</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Bond lengths (Å)</td>
<td>0.048</td>
<td>0.011</td>
<td>0.012</td>
</tr>
<tr>
<td>Angles (°)</td>
<td>0.97</td>
<td>1.49</td>
<td>1.56</td>
</tr>
<tr>
<td>Ramachandran plot</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Residues in favoured regions</td>
<td>2002</td>
<td>1981</td>
<td>1964</td>
</tr>
<tr>
<td>Residues in allowed regions</td>
<td>35</td>
<td>30</td>
<td>35</td>
</tr>
<tr>
<td>Residues in outlier regions</td>
<td>0</td>
<td>1</td>
<td>0</td>
</tr>
<tr>
<td>Mean B factors (Å²)</td>
<td>18.4/15.7/17.3/19.2/17.3/18.4/20.5/22.2/22.6/19.2</td>
<td>34.2/30.7/33.1/29.9/32.2/37.7/40.4/41.2/34.4/34.0</td>
<td>19.7/17.3/19.1/17.2/18.9/22.2/24.0/23.6/19.3/19.6</td>
</tr>
<tr>
<td>Protein atoms per subunit</td>
<td>22.2</td>
<td>36.1</td>
<td>17.1</td>
</tr>
<tr>
<td>NAG</td>
<td>68.7</td>
<td>112.7</td>
<td>87.6</td>
</tr>
<tr>
<td>Ethane-1,2-diol</td>
<td>34.4</td>
<td>46.7</td>
<td>36.6</td>
</tr>
<tr>
<td>Phosphate</td>
<td>41.0</td>
<td>—</td>
<td>54.9</td>
</tr>
<tr>
<td>Oxalate</td>
<td>41.5</td>
<td>—</td>
<td>—</td>
</tr>
</tbody>
</table>

3. Results and discussion

3.1. Binding properties

The affinity of AcChBP for compounds 1–6 was assessed by BLI (Table 2; representative sensorgrams are shown in Supplementary Fig. S2). All compounds displayed \( K_d \) values in the low-nanomolar range, comparable to that of epibatidine (Ondachi et al., 2016), but show a higher affinity than that seen for nicotine. The ligands can be placed into three groups based on affinity for AcChBP: compounds 1 and 6 display \( K_d \) values close to 10 nM, while the values for compounds 3, 4, 5 are around 30 nM and that for compound 2 is 60 nM. For comparison, the \( K_i \) values determined with a \(^{3}H\)-epibatidine displacement assay against \( \alpha 4\beta 2 \) nAChR are presented in Table 2. Consistent with previous AcChBP ligand-binding studies (Hansen et al., 2005), the affinities are reduced significantly compared with the subnanomolar levels observed with actual \( \alpha 4\beta 2 \) nAChR (Ondachi et al., 2016).

3.2. Crystallographic analyses

We attempted to co-crystallize all six compounds with AcChBP and success with three led to structures of the AcChBP–ligand complexes. Crystallographic statistics for the AcChBP–ligand complexes are presented in Table 1. The amino-acid sequences used include human \( \alpha 4 \) (UniProt code P43681), \( \alpha 7 \) (P36544) and \( \alpha 2 \) (P17787) nAChR subtypes.
crystals obtained in each case are isomorphous, with two
pentameric assemblies comprising the asymmetric unit. A high
degree of noncrystallographic symmetry was evident and was
thus maintained in the refinement calculations, although for
AcAChBP-1, the highest resolution structure, these restraints
were released in the final calculations. The ligands of interest
are well defined in the electron density observed in each of
the ten binding sites of the structures (Supplementary Fig. S1) and
and were refined with average B factors that were lower than or
close to the values noted for their associated subunits
(Table 1). Within each complex, the orientation of the ligand
and the pattern of interactions within each binding site are
essentially identical and it is only necessary to describe one.
Enantiomeric mixtures of compounds 1–6 were used, and in
the structures with compounds 1–3 we tested both forms in
modelling to the electron density. The resolution of the crystal
structures was insufficient to distinguish (+) and (−) enan-
tomers or whether a mixture was present, so we used the
former to match that of the naturally occurring form of
epibatidine, the parent compound. Note that the (+) and (−)
forms of epibatidine have very similar binding and biological
properties (Mu et al., 2006; Dallanoce et al., 2012) and we
judge it likely that this also applies to compounds 1–6.

There are three crystal structures containing epibatidine in
the Protein Data Bank (PDB) that are relevant to our study.
These are a low-resolution (3.4 Å) complex with AcAChBP
(PDB entry 2byq; Hansen et al., 2005), a 3.2 Å resolution
structure with α2 (PDB entry 5fw; Li et al., 2011) and a 2.8 Å
resolution complex with an α7 chimera (PDB entry 3sq6;
Kouvatsos et al., 2016). Compounds 1–3 and epibatidine share
the azabicyclo[2.2.1]heptane moiety and this part of the ligand
binds in a similar way in all structures, deep in a hydrophobic
part of the binding site formed primarily on the principal side.
Here, the orthosteric site is dominated by the presence of
aromatic residues (Fig. 2). The protonated amine donates
hydrogen bonds to Tyr110 OH and the Trp164 carbonyl group.
The position of Tyr110 OH is fixed by hydrogen-bonding
interactions with the Ser163 carbonyl and a network of
ordered water molecules that form bridges through to Tyr205,
and Asp214 (not shown). There are van der Waals interactions
with Tyr72, Tyr205, Tyr212, Trp164 and Cys207. The amine is
around 4 Å distant from the face of the Trp164 side chain and
is positioned to suggest the presence of a cation–π interaction.
This interaction is a common and important feature of pLGIC
ligand complexes (Taly et al., 2009; Nys et al., 2013) and
specifically nAChRs (Zhong et al., 1998). The halogen-
substituted pyrimidine ring A, which is common to the series
of compounds and epibatidine, is directed towards the
complementary side of the orthosteric site, with one face of
the aromatic system forming van der Waals interactions with
Val165 from the principal side and the other face interacting
with Ile135 on the complementary side. A side-on interaction
with Cys208 is evident. In the complexes of AcAChBP with
compounds 1, 2 and 3, the fluorine is directed into a shallow
pocket and forms van der Waals interactions with the main
chain of Ala124 and Phe134 and the side chains of Val125 and
Ile135, all from the complementary side. The pyridine N
accepts a hydrogen bond from a water molecule that is at one
end of an ordered solvent chain extending to the surface of the
protein, forming hydrogen bonds to residues in the orthosteric
site, for example to Ile123 O and Trp164 NE1. These parts of
the ligands replicate key features found in AChBP–nicotine
complex structures; in particular, the presence of an ordered
water molecule linking the pyridine N atom to the protein
is noted repeatedly (Hansen et al., 2005; Nys et al., 2013; Dawson
et al., 2019). The available AChBP–epibatidine complex
structures are at low resolution and lack solvent molecules in
the binding sites. However, this hydration pattern is strictly
conserved across the three structures reported here and is
observed in other structures of AChBP complexes (Hansen
et al., 2005; Nys et al., 2013; Dawson et al., 2019). It was
previously thought that the presence of such an ordered water
molecule correlates with agonist activity of the ligand (Nys
et al., 2013), but our structures suggest that such a conclusion
does not apply in all cases, a point that we will revisit below.

The structures in the series 1–6 have a similar substituent: a
phenyl or pyridine ring (termed ring B), with a carboxamide
substituent, at the 4′ position of ring A (Fig. 1). These repres-
ent extensions of the epibatidine scaffold. Compound 1 has
a phenyl group, and compounds 2–6 have pyridines with the N
atom at position 2, 3 or 4. A further variation is a carboxamide
substituent attached to the pyridine N atom at the meta or
para position. The structures of the complexes of AcAChBP
with compounds 1–3 all suggest that ring B participates in van
der Waals interactions with Arg96, Val125 and Met133 from
the complementary side and Cys208 on the principal side
(Fig. 2). In the structure of AcAChBP–1 the alignment and
distances (around 3.3 Å) of the Tyr212 OH group and a
carbon on ring B suggest the possibility of a C—H···O
hydrogen bond (Fig. 2a). In the complexes with compounds 2
and 3 (Fig. 2b) the pyridine N atom participates in a hydrogen
bond to Tyr212 with distances of around 2.6–3.0 Å.

In the AcAChBP–1 and AcAChBP–3 complexes the
carboxamide is directed outwards towards bulk solvent.
The carbonyl groups participate in water-mediated links to Val125,
Thr127 and Ser131 (not shown). The amide groups contribute
to a phosphate-binding site, with the oxyanion also coordi-
nated by the side chains of Asp94 and Arg96 on the

Table 2
Ligand-binding properties.
Values are given with standard errors.

<table>
<thead>
<tr>
<th>Ligand</th>
<th>AcAChBP, Kd (nM)</th>
<th>αβ, nAChR, Kd (nM)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>9.8 ± 0.03</td>
<td>0.12 ± 0.020</td>
</tr>
<tr>
<td>2</td>
<td>60.0 ± 0.14</td>
<td>0.28 ± 0.010</td>
</tr>
<tr>
<td>3</td>
<td>29.0 ± 0.05</td>
<td>0.14 ± 0.070</td>
</tr>
<tr>
<td>4</td>
<td>33.0 ± 0.20</td>
<td>0.07 ± 0.002</td>
</tr>
<tr>
<td>5</td>
<td>30.0 ± 0.06</td>
<td>0.28 ± 0.030</td>
</tr>
<tr>
<td>6</td>
<td>9.8 ± 0.01</td>
<td>0.67 ± 0.280</td>
</tr>
</tbody>
</table>

† [3H]-Epibatidine competition assay, Kd = 0.02 nM (Ondachi et al., 2016). † Trypto-
phan fluorescence-quenching assay (Hansen et al., 2005). § Isothermal titration
calorimetry (Rucktooa et al., 2012). ¶ [3H]-Nicotine competition assay (Coe et al.,
2005).
complementary side and then Glu170, Glu210, Lys42 and Ser167 from the principal side. This is further detailed below. Phosphate was a component of the crystallization mixture, and the acidic pH used explains why aspartate and glutamate side chains coordinate the oxyanion. In eight binding sites in AcAChBP–1 and seven in AcAChBP–3 the amide forms direct hydrogen bonds to Asp94 on the (–) side, perhaps helped by the ordering influence of the oxyanion. Due to the difference in the ring position of the carboxamide substituent in compound 2, in AcAChBP–2 the carbonyl group of the ligand is tilted towards loop E on the complementary side, in particular Met133, but ring B is positioned further away from the methionine side chain. An adjustment of the Tyr212 side chain is observed, serving to maintain the hydrogen bond to the ring B N atom. The result is to position the carboxamide in roughly the same position in all three complexes. The pyridine ring B of compound 2 in comparison is posed closer to Tyr212, the position of the amide group is adjusted and our analysis indicated that ethane-1,2-diol, the cryoprotectant, and not phosphate interacts with the amide and engages with the same residues that form the phosphate-binding pocket. A link from the amide of compound 2 to Asp94 is retained but via bridging water molecules (not shown). The position of ring B may provide a disruptive block to phosphate binding, and the reduction in van der Waals interactions with Met133 may contribute to compound 2 displaying the lowest affinity for AcAChBP.

Loop C, on the (+) side, contributes significantly to the creation of the orthosteric site and interacts directly with residues on the (–) side. This loop has to present an open conformation to allow ligands to enter the site and then close as the complex forms (Jadey & Auerbach, 2012). When ligands 1–3 occupy the site, loop C closes over them (Fig. 3). The solvent-accessible surface areas of unbound compounds 1–3 are approximately 500 Å², and between 91% and 94% of this surface is buried in the complexes with AcAChBP, with primarily the carboxamide moiety directed towards solvent. Epibatidine is smaller (Fig. 1), with a solvent-accessible area of 360 Å², and is almost entirely buried (>99%) deep in the binding site, with loop C in the agonist-bound state (Fig. 3; Spang et al., 2000).

Compounds 2, 5 and 6 differ only in the position of the N atom on pyridine ring B. The AcAChBP–2 complex indicates a displacement of ring B compared with the other structures and we speculate that this might influence the interactions in and around the observed oxygen-binding site. The placement of the ring B N atom in compound 6 would position a hydrogen-bond acceptor in an ideal position to participate in formation or organization of the anion-binding site. Using an overlay of complexes with compounds 1 and 2 as a template, the N atom in ring B would be about 2.4 Å from a phosphate in compound 6 and around 2.9 Å in compound 5.

Comparisons of the AcAChBP complexes with the cryo-EM structure of nAChR in complex with nicotine (PDB entry 6cnj; Walsh et al., 2018) and the sequences of human α4, α7 and β2 subtypes were carried out (Fig. 4). We considered the orthosteric sites of human
heteromeric α4(+)/β2(−) nAChR and homomeric α7 nAChR and first asked how similar the mode of binding of compounds 1–3, and by inference compounds 4–6, might be. The conservation of sequence and structure, in particular considering the aromatic cage of the binding site, suggests that the orientation of the epibatidine analogues is representative of how this compound series would bind to any nAChR. This conclusion is supported by site-directed mutagenesis studies with α7 nAChR, where changes to six residues abolished epibatidine binding (Thompson et al., 2017). These six residues, which are highly conserved as a group, correspond to Tyr72, Tyr110, Tyr205, Tyr212, Ile123 and Trp164 in AcAChBP (Figs. 2 and 4).

The similarity of the amino acids implicated in interactions with the compound set 1–6 extends beyond these six residues. On the principal side, Val165 of AcAChBP, which participates in van der Waals interactions with the ligands, aligns with Thr183 and Ser172 in the α4 and α7 subtypes, respectively. In terms of size, these represent conservative substitutions. On loop C, Ser206 is changed to a glutamate in nAChR but the side chain is directed away from the binding site. On the complementary side, Met133 and Ile135 of AcAChBP are

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**Figure 3**

Loop C of AcAChBP adopts different conformations. Ribbon representation of loop C segments with the bound agonist epibatidine (cyan, PDB entry 2byq; Hansen et al., 2005), antagonist strychnine (grey, PDB entry 5o8t; Dawson et al., 2019) and the AcAChBP–1 and AcAChBP–3 complexes (red and black, respectively). Compound 3 is shown and coloured with the following scheme: C, black; O, red; N, blue; F, dark green. The overlay was calculated using all Cα atoms of a single subunit.

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**Figure 4**

(a) Alignment of selected sequence segments that form the orthosteric binding sites in AcAChBP and α4, α7 and β2 nAChRs. Loops are labelled and are split into principal (+) and complementary (−) sides. Residues coloured blue are key and are discussed in the text. Val125 and Thr127 of AcAChBP are in the N-terminal section of loop E but are left out for clarity. (b) A schematic of the orthosteric binding site listing the key residues of AcAChBP and the corresponding residues in the α4(+)/β2(−) nAChR heteromeric site. Loop F is out of range of the ligands discussed in this work and has been omitted. Arg96 is included given its role in binding the anion (see text). Boxes are coloured yellow to highlight strict conservation, grey for the (+) side and cyan for the (−) side. (c) Schematic with the same format for the α7 nAChR homomeric site.
Phe144 and Leu146 in nAChR β2 and Gln139 and Leu141 in α7, respectively. At the periphery of the binding site the combination of Val125 and Thr127 in AcAChBP is Val136 and Ser138 in β2 and Asn133 and Leu131 in α7. The noteworthy difference is the Met133Gln substitution, which in α7 nAChR will place a polar side chain near the ligands. It is possible that the latter would provide a steric restriction on the placement of ring B in 1 and 3, and also that the glutamine could form a hydrogen bond to the carboxamide substituent when binding 2.

Different ideas have been proposed to explain the activity of agonists and antagonists of pLGICs based on the crystal structures of AChBP–ligand complexes (Taly et al., 2009; Lemoine et al., 2012; Bertrand et al., 2015). The use of AcAChBP provides binding and structural data that inform on affinity and aspects of molecular recognition, but it is important to recognize the limitations of this surrogate (for example it has no transmembrane domains) and exercise caution when considering aspects of channel opening and closing. At least three factors appear to contribute to the distinctive responses of ligands that bind to the orthosteric site of pLGICs. Firstly, residues on the complementary side are the primary determinants of ligand affinity; in particular, by combining the presence of cation–π and van der Waals interactions with the aromatic cage to stabilize the complex. This area of the orthosteric site is well targeted by compounds 1–6; hence, they are high-affinity ligands and their potent antagonist effects on nicotine-induced antinociception at nAChRs (Ondachi et al., 2016) is likely to be a consequence of being bioavailable and able to outcompete nicotine (Table 2).

Secondly, the conformation of loop C appears to be relevant. In general, this loop displays three states. There is a fully contracted, closed and clamped conformation when binding small agonists (for example nicotine or epibatidine), a fully extended and open form in the presence of larger antagonist ligands (for example strychnine) and an intermediate state that is stabilized in the presence of partial agonists (for example varenicline). However, the conformation of loop C does not always correspond in such a simple fashion to the pharmacological profile of a ligand. The noteworthy exception is the high-potency antagonist dihydro-β-erythroidine, which induces a loop C conformation similar to that of agonists (Shahsavari et al., 2012).

In a study of the drug varenicline, Billen et al. (2012) drew attention to a third factor in ligand response. Partial agonists can desensitize pLGICs with the ability to induce opening with higher affinity but with lower efficacy than a full agonist. By targeted site-directed mutagenesis and electrophysiology on α4β2 nAChR they showed that residues on loops D and E, the complementary side of the orthosteric site, contribute to desensitization and channel opening. In particular, interactions of β2 nAChR residues Trp82 and Leu146 are important for channel opening. In AcAChBP these correspond to Tyr72 on loop D and Ile135 on loop E (Fig. 4). These residues are conserved and interact with compounds 1–3, and by implication also with compounds 4–6. Compounds 1 and 3 represent an anomaly, being agonists of α7 nAChR (Ondachi et al., 2016). The loop C conformations we observe are similar and in an intermediate state. The AcAChBP complex structures with compounds 1 and 3 suggest van der Waals interactions with Val125 and Met133, which are on loop E. These two residues differ in the α7 and β2 nAChR forms (Fig. 4). In β2 Val125 and Met133 correspond to a valine and a phenylalanine, respectively. In the α7 form they are a leucine and glutamine, respectively, and we speculate that different interactions between the ligands and this part of the orthosteric site may be linked to the agonist property of compounds 1 and 3.

Allosteric control is an important facet of pLGIC function and has been studied extensively in nAChRs (Taly et al., 2014; Chatzidaki & Millar, 2015). Of note is the role of Ca2+, which increases the affinity for agonists and potentiates their activity on nAChRs, producing an increase in current amplitudes. A number of residues at distinct sites on nAChR structures are implicated in Ca2+ binding, and this is indicative of multiple sites of regulation (Galzi et al., 1996). In this context, the identification of oxyanion binding in two of the structures reported here, involving charged and conserved residues at the subunit interface near the orthosteric site and which bring the key loop C into play, is intriguing (Fig. 5). These residues have not been previously discussed or investigated in the context of Ca2+ binding or allosteric regulation to the best of our knowledge. It may seem counterintuitive to invoke cation binding to a site where an oxyanion is found. However, our crystals were grown in the presence of phosphate and at acidic pH. The bound phosphate may be an artefact of these conditions, but nevertheless draws attention to a group of charged residues that are able to interact with each other or...
with an ion, which is exactly the type of feature that is likely to be involved in allosteric regulation.

Asp94 and Glu170 in AcAChBP, which as mentioned are likely to be protonated, are replaced by residues which are well established as phosphate-interacting amino acids in nAChR sequences. In human \(\beta_2\), \(\alpha_4\) and \(\alpha_7\) nAChR, Asp94 corresponds to Lys104, Ser110 and Thr99, respectively, whilst Glu170 corresponds to Glu182, Lys188 and Ser177, respectively. These are adjacent to the strictly conserved Arg96 (Arg110 and Arg101 in \(\beta_2\) and \(\alpha_7\), respectively) on the complementary side. From the principal side AcAChBP Glu210, on loop C, aligns with Ser219 in the \(\beta_2\) form, Glu228 in \(\alpha_4\) and Glu215 in \(\alpha_7\) (Fig. 3). Note, however, that loop C in the \(\beta_2\) form is potentially very different from those of \(\alpha_4\) and \(\alpha_7\) (Fig. 4). Adjacent to Glu210 is Ser167, which corresponds to Asp185 (\(\alpha_4\)), Gly174 (\(\alpha_7\)) and Asp179 (\(\beta_2\)). It would be too speculative to align Lys42 in AcAChBP with a specific residue in \(\alpha_4\) or \(\alpha_7\) since it is in a region where, as shown by an overlay with the cyro-EM structure of an nAChR (not shown), conservation is lacking. Nevertheless, the residue types in this location would be compatible with an anion-binding site and such an event may influence the activity of compounds 1-6. This site could represent a point of allosteric control to gating.

4. Conclusions

Using AcAChBP as a surrogate of nAChRs, we characterized the binding affinity of a series of epibatidine analogues and were able to determine three crystal structures to inform on protein–ligand interactions. These data may inform the design of new nAChR targeting ligands with defined pharmacological properties. Differing interactions with Tyr212 and Asp94 of AcAChBP appeared to mediate ligand affinities, with interactions at Asp94 potentially also explaining their subtype-selective nAChR pharmacological profiles due to the differing equivalent residues. It may be possible to modify the epibatidine framework, or indeed some other scaffold, to build interactions at a putative ion-binding site to investigate further. Data derived from compounds 1 and 3 suggest that distinction between \(\alpha_4/\beta_2\) and \(\alpha_7\) nAChR forms might involve two residues on loop E, the complementary side of the orthosteric binding site, and it could be instructive to test this hypothesis. Future work will seek to address these issues.

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References


