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# Phenotype Standardization for Statin-Induced Myotoxicity

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**Statins are widely used lipid-lowering drugs that are effective in reducing cardiovascular disease risk. Although they are generally well tolerated, they can cause muscle toxicity, which can lead to severe rhabdomyolysis. Research in this area has been hampered to some extent by the lack of standardized nomenclature and phenotypic definitions. We have used numerical and descriptive classifications and developed an algorithm to define statin-related myotoxicity phenotypes, including myalgia, myopathy, rhabdomyolysis, and necrotizing autoimmune myopathy.**

Statins, 3-hydroxy-3-methylglutaryl coenzyme A reductase inhibitors, are effective in lowering blood cholesterol as well as in primary and secondary prevention of cardiovascular events. Approximately 25 million people worldwide are taking statins.<sup>1–3</sup> Statins are safe and effective in the majority of patients, but they are associated with muscle toxicity, which, although rare, can be serious and potentially life threatening.<sup>4,5</sup> The clinical spectrum of statin-induced myotoxicity varies greatly from asymptomatic elevations of creatine kinase (CK) without muscle pain, to muscle pain or weakness with raised CK levels, myositis with biopsy-proven muscle inflammation, and, finally, rhabdomyolysis with muscle symptoms, high CK, and potential for acute kidney injury (Table 1).<sup>6,7</sup> Whether statins cause muscle pain in the absence of CK changes remains controversial. Although milder forms of myotoxicity tend to be self-limiting and disappear after cessation of therapy, they can lead to poor quality of life, poor drug compliance, and, consequently, the failure to prevent cardiovascular events.

Because statin-induced myotoxicity is uncommon, it is necessary to pool and analyze data from various sources, including multicenter clinical trials and observational studies, to study genetic etiology. In addition, electronic medical records linked

to biological repositories or to individual patients have been useful in identifying and recruiting patients with drug-induced toxicities.<sup>6,8–12</sup> The Phenotype Standardization Project was started a few years ago to facilitate such multicenter research collaborations on various types of serious adverse drug reactions (ADRs). Phenotype consensus papers on drug-induced liver injury, skin injury, and torsade de pointes have been published by multidisciplinary groups of international experts.<sup>13–15</sup> The aim of this article is to provide consensus definitions for statin-induced myotoxicity phenotypes, to facilitate cross-study comparisons from the existing cohorts, to aid in the recruitment of retrospective and newly diagnosed patients with statin-induced muscle damage, and to define the phenotype for genomic data analyses.

We convened an international expert workshop on statin-induced myotoxicity in December 2013 in Liverpool, UK, to agree on definitions and a minimum set of criteria to help in identification and recruitment. We report on the deliberations of this workshop: first, we summarize evidence of the clinical and biochemical phenotypes that have been reported, and second, we report on our suggested standardization of the terminology and phenotypes of statin-induced muscle toxicity.

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**Table 1 Statin-related myotoxicity phenotype classification**

SRM classification	Phenotype	Incidence	Definition	Reference
SRM 0	CK elevation <4× ULN	1.5–26%	No muscle symptoms	Refs. 1,20,34,35,67
SRM 1	Myalgia, tolerable	190/100,000 Patient-years; 0.3–33%	Muscle symptoms without CK elevation	Refs. 1,19,21,50,68
SRM 2	Myalgia, intolerable	0.2–2/1,000	Muscle symptoms, CK <4× ULN, complete resolution on dechallenge	Ref. 20
SRM 3	Myopathy	5/100,000 Patient-years	CK elevation >4× ULN <10× ULN ± muscle symptoms, complete resolution on dechallenge	Ref. 1
SRM 4	Severe myopathy	0.11%	CK elevation >10× ULN <50× ULN, muscle symptoms, complete resolution on dechallenge	Refs. 20,69
SRM 5	Rhabdomyolysis	0.1–8.4/100,000 Patient-years	CK elevation >10× ULN with evidence of renal impairment + muscle symptoms or CK >50× ULN	Refs. 4,6,25,44,45
SRM 6	Autoimmune-mediated necrotizing myositis	~2/million per year	HMGCR antibodies, HMGCR expression in muscle biopsy, incomplete resolution on dechallenge	Refs. 51,70

Numeric classification was developed by an expert group; descriptive nomenclature was adapted from the recommendation of the American College of Cardiology/American Heart Association/National Heart, Lung, and Blood Institute Clinical Advisory Board.<sup>3,7</sup>

CK, creatine kinase; HMGCR, 3-hydroxy-3-methylglutaryl-coenzyme A reductase; SRM, statin-related myotoxicity; ULN, upper limit of normal.

### CONSENSUS PROCESS

The PREDICTION-ADR consortium, funded by the EU Seventh Framework Programme, organized a joint meeting on 10 December 2013 in Liverpool with a multidisciplinary team of experts on statin-related myopathy and angiotensin converting enzyme-inhibitor angioedema. An international team with known expertise in the area was assembled by invitation. The group comprised clinical and basic pharmacologists, internists, rheumatology and myopathy experts, immunology and clinical chemistry scientists, allergists, regulatory agency representatives, and managers of electronic medical record databases.

A draft manuscript circulated before the meeting contained a brief literature review on statin-related myopathy and a description of main phenotypes. The expert group was asked to consider the minimum set of criteria for patient recruitment. An introductory talk was given to start the discussions. In addition, an algorithm was designed to aid standardization of nomenclature and phenotypes, and recommendations were made about the types of data to be collected from each patient (Figure 1). After the meeting, the revised manuscript comprising the criteria (Table 1) and algorithm was approved by all contributors.

We did not discuss causality assessment formally, as the challenges of this task are common for many ADRs.<sup>16</sup> They require establishing a temporal relationship, along with dechallenge and rechallenge information, and, particularly important for statin-related myotoxicity, vigilant information on muscle injury from falls, trauma, or vigorous exercise. We also recommend independent assessment of causality by an adjudication panel, using methodology that has been successfully applied to retrospective and prospective recruitment of patients with rare ADRs, including severe drug-induced skin injury.<sup>15</sup>

### INCIDENCE

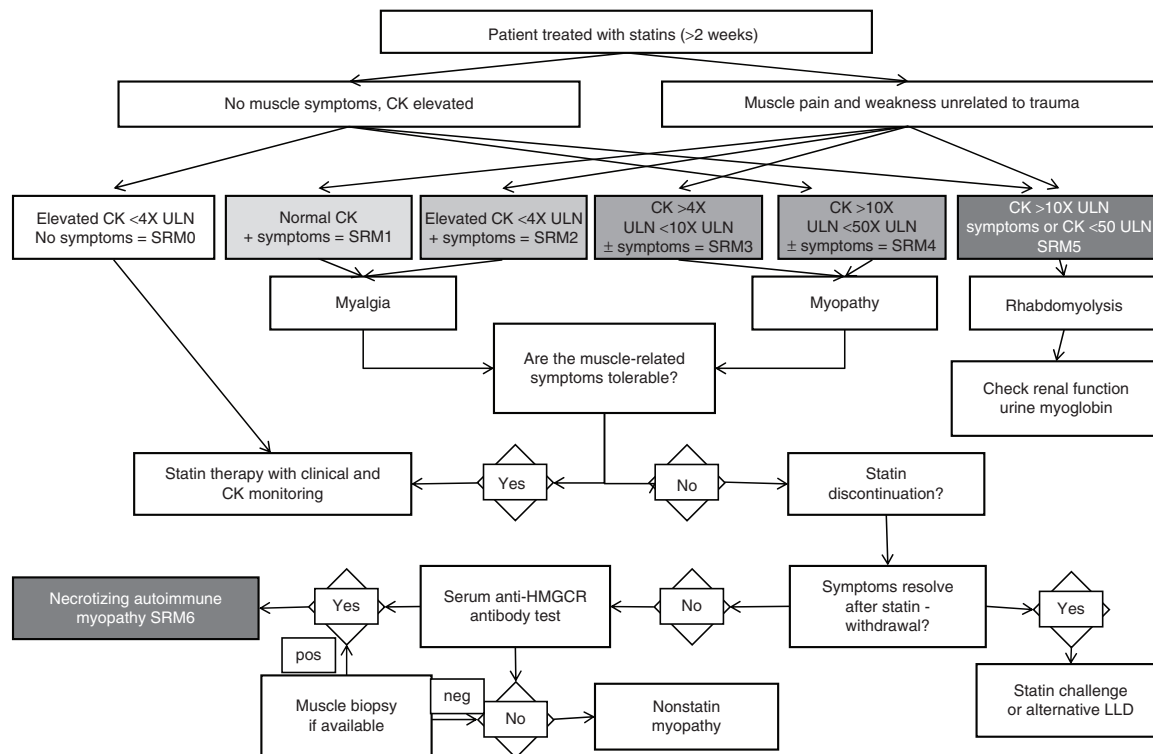
The rate of statin-induced myotoxicity has been estimated from adverse event data in case series and randomized controlled trials.<sup>17–24</sup> However, estimating adverse events from randomized controlled trials may be challenging because (i) they are not

usually powered to capture low-frequency ADRs; (ii) patients are exposed to drugs and monitored only for a short period of time when myotoxicity may not yet have developed; (iii) the diagnostic criteria for reaction events may vary across different trials; and (iv) they may have strict inclusion and exclusion criteria that may exclude some populations who are at higher risk of ADRs.

The risk of rhabdomyolysis among hospitalized patients receiving lipid-lowering drugs has recently been estimated in a real-world clinical setting. Claims data from 9 million members of five US health plans were used to confirm 42 cases in >470,000 patients exposed to lipid-lowering drugs. The risk of rhabdomyolysis for individuals on different statin preparations, including combination therapy, and the risk of comorbidities were estimated to be between 0.3 and 8.4 in 10,000 patient-years.<sup>25</sup> The authors estimated that the risk in comparison with atorvastatin as a reference was significantly higher when statins were used in combination with cytochrome P450 (CYP)3A4 inhibitors (odds ratio: 7.1, 95% confidence interval: 1.6–31.6) and for cerivastatin monotherapy (odds ratio: 4.7, 95% confidence interval: 1.1–21.1).<sup>25</sup>

### DIAGNOSTIC CRITERIA

The diagnosis of statin-induced myotoxicity is based on medical history, clinical examination, and laboratory tests and can be confirmed by muscle biopsy (Supplementary Table S1 online). Muscle biopsies can reveal muscle fiber necrosis, type II fiber atrophy, and increased lipid stores in muscle fibers or inflammation.<sup>26–29</sup> In clinical practice, a pragmatic approach is adopted: discontinuation of the culprit drug and avoidance of its future use.<sup>30</sup> Monitoring of statin therapy for muscle toxicity includes CK measurements, although routine laboratory testing is recommended only for symptomatic patients.<sup>31</sup> The potential harm of introducing routine CK monitoring in all patients who take statins may outweigh the benefits from several perspectives, including false-positive results with potentially ensuing invasive investigations, a psychological effect on patients that may result in reduction of statin use, and an increased cost to the health-care system. A recent study on patients' perceptions



**Figure 1** Algorithm for defining the type of statin-related myotoxicity. New nomenclature was introduced to reflect the phenotype classification and severity (SRM 0 to SRM 6) of statin-related toxicity. CK, creatine kinase; HMGCR, 3-hydroxy-3-methylglutaryl-coenzyme A reductase; LLD, lipid-lowering drug; SRM, statin-related myotoxicity; ULN, upper limit of normal.

of statin therapy has demonstrated reduced adherence to statin therapy in those concerned about ADRs.<sup>32,33</sup>

**CLINICAL PRESENTATIONS OF STATIN-INDUCED MUSCLE TOXICITY**

Statin-induced muscle toxicity can present in many different ways, but the recognized phenotypes and degrees of severity are classified into seven different categories, as represented in **Table 1** (statin-related myotoxicity (SRM) 0–SRM 6).

**Muscle symptoms**

Statin-induced muscle toxicity may present with a wide variety of symptoms, including fatigue, muscle pain, muscle weakness, muscle tenderness, and cramps. These symptoms are usually proximal and symmetrical but may be generalized. They can be aggravated by vigorous exercise and sometimes by addition of a new medication.<sup>2</sup> Pain tolerance varies greatly in different individuals, and many studies have relied on self-reporting of muscle symptoms. If patients can tolerate mild muscle pain, statins are not usually discontinued (SRM 1, **Table 1**).

**Plasma CK elevation**

Asymptomatic serum CK elevations (SRM 0) and muscle pain without an increase in CK (SRM 1) have been the two most commonly (occurring in up to 33% of patients) described features of statin-induced toxicity.<sup>20,34,35</sup> With the increasing use of electronic medical records, it may be possible to use CK elevations to identify patients with statin myotoxicity. However,

because the use of CK as a screening test for muscle injury is not recommended, the identification of an isolated CK elevation in an electronic (or paper-based) record may indicate suspicion of the clinician of muscle toxicity, even if symptoms are variably recorded. CK levels are often used as a crude estimate of severity, but the correlation between muscle symptoms and CK levels is imperfect, and the clinical interpretation of CK levels is complex.<sup>36</sup> There is considerable variability in the inclusion criteria and CK levels in the literature, particularly in genetic susceptibility studies. Some authors investigate patients with self-reported myalgia and CK levels from 1 to 3× the upper limit of normal (ULN) (SRM 2), whereas others apply more stringent criteria with CK elevations >4× (ref. 1) (SRM 3) or >10× the ULN (SRM 4) for myopathy and ≥50× the ULN for rhabdomyolysis (SRM 5),<sup>9,35,37–42</sup> which are the criteria used in some recent industry-funded studies. To prevent inclusion of patients with CK elevations from causes other than statin myotoxicity, we have adopted the >4× the ULN for myopathy (SRM 3) and >10× the ULN for severe myopathy (SRM 4). Although these cutoff points have been commonly used in clinical trials and in several guidelines, they are somewhat arbitrary. However, future analyses based on these cutoff points (<4× the ULN, 4–10× the ULN, and >10× the ULN) will help in defining the sensitivity and specificity of different genetic markers, and provide a first step in future refinement of cutoff levels. Some investigators have also used alanine aminotransferase elevation to identify muscle injury;<sup>35</sup> this may be of use when measured in combination with CK. Isolated elevation of alanine aminotransferase, in the

**Table 2** Reported risk factors for developing statin-induced myopathy

Risk factor	Description	Reference
Patient factors		
Advanced age	>80 years)	Ref. 7
Female gender		Ref. 42
Low body mass index		
Comorbidities	Untreated hypothyroidism	Ref. 50, Supplementary refs. 71 and 72
	Low vitamin D level	Supplementary ref. 73 online
	Chronic renal insufficiency (GFR 60 ml/min), especially when associated with diabetes	Ref. 7,25
	Infection, liver impairment, hypertension	Ref. 25
	Alcohol abuse	Ref. 7
Physical exercise		Supplementary refs. 74–76
Surgery	The American Heart Association recommends temporary cessation of statins before major surgery	Ref. 7
History of statin myopathy	Personal or family history of statin myopathy	Ref. 50
Drug factors		
Higher statin dose	Increased frequency of myopathy	Supplementary refs. 77 and 78
Interacting drugs <sup>a</sup>	CYP3A4 enzyme inhibitors (particularly important for CYP3A4 substrates simvastatin, lovastatin, and atorvastatin): diltiazem, verapamil, clarithromycin, telithromycin, erythromycin, itraconazole, cyclosporine, protease inhibitors (ritonavir, indinavir, and saquinavir), amiodarone, and fusidic acid	Refs. 59–62, Supplementary refs. 79–81
	CYP2C9 enzyme inhibitors (with effects on fluvastatin, a CYP2C9 substrate): omeprazole and fluconazole	Supplementary refs. 82–84
	OATP1B1 inhibition (with effects on simvastatin, pravastatin, lovastatin, and rosuvastatin): gemfibrozil <sup>b</sup>	Refs. 53–57 Supplementary refs. 85 and 86
Diet-related interactions	Grapefruit juice (>200 ml daily) increases levels of simvastatin, atorvastatin, and lovastatin	

Supplementary refs. 71–87 are provided with Supplementary Table S1 online.

CYP, cytochrome P450; GFR, glomerular filtration rate; OATP, organic anion-transporting polypeptide.

<sup>a</sup>Only a selection of drug interactions with statins has been presented. Readers should refer to more extensive reviews for a fuller list of interacting drugs (Kellick *et al.*, 2014 (Supplementary ref. 87), <http://www.mhra.gov.uk/Safetyinformation/DrugSafetyUpdate/DrugSafetyUpdatesearchresults/index.htm>, and <http://pharmacistsletter.therapeuticresearch.com/pl/ArticleDD.aspx?nidchk=1&cs=&s=PL&pt=2&fpt=31&dd=280606&pb=PL&cat=4803&segment=4421&AspxAutoDetectCookieSupport=1>).<sup>b</sup>The main mechanism of the interaction is mentioned in the table, but it is important to note that in some cases there may be multiple mechanisms. For example, with gemfibrozil, in addition to OATP1B1 inhibition, CYP2C8 and UGT1 inhibition may also contribute. UGT, uridine diphosphate glucuronyltransferase.

absence of an increase in CK levels, should raise the suspicion of liver injury, which can also rarely occur with statins.<sup>43</sup>

### Rhabdomyolysis

Rhabdomyolysis (SRM 5), the most serious ADR associated with statins,<sup>4,6,25,44,45</sup> is characterized by muscle necrosis, release of myoglobin into the bloodstream, and sometimes acute renal failure.<sup>4,6,26,45,46</sup> Muscle symptoms are accompanied by marked CK elevation, typically greater than 50× the ULN. Pigment nephropathy with brown urine is typically evident due to myoglobinuria and is consistent with serum creatinine elevation.<sup>7</sup>

### HMGR autoantibodies

A number of studies (e.g., ref.46) have reported the occurrence of an autoimmune-mediated necrotizing myopathy (SRM 6) after statin exposure, with symptoms that

continued after drug withdrawal. Serum autoantibodies against 3-hydroxy-3-methylglutaryl-coenzyme A reductase (HMGCR), the pharmacological target of statins, have been reported in patients with statin-induced autoimmune myopathy.<sup>47</sup> Muscle biopsies can also be diagnostically useful for the detection of HMGCR expression on the cell surface of regenerating muscle fibers.<sup>47</sup> In 51 patients with self-limited statin intolerance, anti-HMGCR antibodies were not observed in a single individual.<sup>48</sup> This suggests that self-limited statin myopathy has a distinct biological mechanism from statin-induced autoimmune myopathy. In addition, atorvastatin and simvastatin promote a proinflammatory or Th1 response in activated peripheral blood mononuclear cells by increasing the number of interferon- $\gamma$ -secreting T cells.<sup>49</sup> It should be noted, however, that many autoimmune myopathy cases are likely to be excluded from studies because withdrawal of the statins does not alleviate symptoms and

thus statins may be discounted as the causal agents for myopathy pathogenesis.

### TIME TO ONSET

In the PRIMO (Prediction of Muscular Risk in Observational Conditions) study, the median time to onset in the 832 of 7,924 patients who developed muscular symptoms was one month after statin initiation.<sup>50</sup> A study of 45 patients reported a mean duration of statin therapy before myopathic symptoms of 6.3 months, with a maximum of 9.8 months.<sup>19</sup>

A large study, using a case-crossover design to determine statin myotoxicity in two primary-care databases comprising 93,831 patients, suggested that most cases occur within the first 12 weeks of statin exposure.<sup>21</sup> The authors recommended a 26-week cutoff to enable fewer misclassifications of exposed cases.<sup>21</sup> A 26-week cutoff seems sensible for those patients who have been on a stable dose of statin monotherapy; however, this may need to be extended. It is important to note that some patients can develop statin myotoxicity either after a statin dose increase or after the concomitant administration of an interacting drug, which may occur anytime during the life cycle of statin use. Furthermore, autoimmune myopathy may take longer to develop, with a time to onset as long as 3 years.<sup>51</sup>

### RISK FACTORS

#### Concomitant medications

**Fibrates.** A significant body of evidence suggests an increased risk of statin myopathy, particularly rhabdomyolysis, in patients taking statins in combination with fibrates.<sup>4</sup> Analysis of the US Food and Drug Administration Adverse Event Reporting System between 1998 and 2002 was used to determine reporting rates for rhabdomyolysis in patients taking fenofibrate and gemfibrozil in combination with statins.<sup>52</sup> Gemfibrozil, a fibric acid derivative, increases systemic exposure to active simvastatin acid by inhibiting both glucuronidation and organic anion-transporting polypeptide (OATP)1B1 transporter-mediated uptake into the liver.<sup>53,54</sup>

Overall rates of rhabdomyolysis for any statin medication users coprescribed fenofibrate or gemfibrozil were 4.5 and 8.7 per million prescriptions, respectively. When stratified to those patients receiving cerivastatin only, the rates increased to 140 and 4,600 per million prescriptions with fenofibrate/cerivastatin and gemfibrozil/cerivastatin combination therapy, respectively.<sup>52</sup> Although the effect of fibrate coadministration on statin bioavailability exists for the majority of statins,<sup>53,55–57</sup> the increase in myopathy risk is especially pronounced with cerivastatin.<sup>5,58</sup> Although cerivastatin was withdrawn in 2001, the identification of predisposing pharmacogenomics factors may still be of use for elucidating predisposing factors of muscle toxicity with the statins more commonly used nowadays.

**CYP3A4 inhibition.** CYP3A4 inhibitors such as azole antifungals, protease inhibitors, amiodarone, cyclosporine, calcium channel blockers, and macrolide antibiotics, to name a few, increase risk of myopathy for statins that undergo CYP3A4

metabolism (simvastatin, atorvastatin, and lovastatin).<sup>59–62</sup> In addition, some foods such as grapefruit juice, which contains furanocoumarins, irreversibly inhibit CYP3A4 in the gut. The effect is reduced gut wall metabolism of statins (particularly simvastatin) and increased systemic exposure, which can lead to adverse effects.<sup>60,63</sup>

#### Comorbidities

A list of comorbidities that are associated with an increased risk of developing statin-related myotoxicity is shown in **Table 2**. They include hypothyroidism, chronic renal insufficiency, infection, impaired liver function, hypertension, physical exertion, and diabetes.

#### Exercise

It is commonly believed that vigorous exercise increases the risk of statin-induced myopathy. Indeed, it is thought that myopathic symptoms may occur in 25% of statin users who exercise, as compared with a population incidence estimated at 1–5% for those who exercise but do not take statins.<sup>64</sup> In professional athletes, it is estimated that as many as 75% of those taking statins may develop muscular symptoms;<sup>65</sup> however, this may be overestimated.

### CONCLUSIONS

Given the high prevalence of statin use worldwide and their significance in the prevention of cardiovascular and cerebrovascular disease, extensive research on the prediction and prevention of serious adverse effects such as statin-related myotoxicity is justified. Improved patient tolerability and adherence to statins is crucial because it reduces the incidence and the cost to any health-care system of treating cardiovascular disease. Large prospective studies of patient cohorts treated with statins are required to identify new genetic susceptibility biomarkers for statin-related myotoxicity that could be implemented into clinical practice. To date, one of the problems in comparing the results of observational studies on statin myotoxicity has been the lack of phenotype classification and standardization of nomenclature. In addition, given the rarity of the most severe phenotypes, a small sample size of several studies has hampered genetic biomarker discovery.

We have adapted a previously described consensus approach<sup>15</sup> to define phenotypic criteria that can be used in an effort to standardize statin-related myotoxicity phenotypes using the numerical and descriptive nomenclature given in **Table 1**. Our standardization was based on expert opinion from a multidisciplinary group and the literature. The following are key criteria to be used in the deep phenotyping of patients with suspected statin-induced muscle injury:

- A CK level that is  $>4\times$  the ULN in the presence or absence of clinical symptoms for the definition of myopathy. We have adopted the  $>4\times$  the ULN level pragmatically, as we feel, based on the literature, that this cutoff provides the right balance in preventing inclusion of patients with CK elevation due to normal variation or other causes, while

simultaneously ensuring that we do not unnecessarily exclude valuable patients in studies investigating genetic factors predisposing to statin myotoxicity. A CK >10× the ULN should be categorized as rhabdomyolysis if accompanied by renal impairment. The adoption of the classification shown in **Table 1** may help in more clearly defining the various phenotypes that are recruited in different studies. Papers reporting genetic factors predisposing to statin-induced muscle injury should show the effect size of the genetic polymorphism based on the degree of CK elevation, as has been done in recent studies.<sup>9,66</sup>

- Clinical symptoms need to be carefully recorded in the patients; these should include not only the muscle symptoms but also whether there is any involvement of the kidneys, which might indicate rhabdomyolysis. Causality assessment as to whether the statin was responsible should include details of the temporal relationship between onset of statin use and the occurrence of myotoxicity, the effect of dechallenge, and the effect of any rechallenge. Dechallenge may not always be successful, for example, in patients with autoimmune myopathy, and should not necessarily be used to exclude statins as etiological agents.
- Apart from CK, measurement of alanine aminotransferase, urine myoglobin levels (when clinically indicated), and renal function may be useful. In patients with a suspected autoimmune myopathy, the measurement of anti-HMGCR antibodies and muscle biopsy should be considered.
- Predisposing factors, including interacting drugs, comorbidities, and exercise, should be evaluated in all patients. Factors such as trauma that lead to muscle injury irrespective of statin use need to be excluded.
- The time to onset can be variable and can be delayed as long as 3 years in patients with autoimmune myopathy. However, in general, most cases of statin-induced myotoxicity occur within 6 months to 1 year of statin onset, an increase in the dose of the statin, or the concomitant administration of an interacting drug.

We have also developed an algorithm that will help assign phenotypes to individual patients based on clinical and biochemical parameters (**Figure 1**).

We hope the criteria described in this article will help clinicians and researchers to categorize phenotypes in patients with statin-related myotoxicity in order to facilitate research in this area.

**SUPPLEMENTARY MATERIAL** is linked to the online version of the paper at <http://www.nature.com/cpt>

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#### CONFLICT OF INTEREST

The authors declared no conflict of interest.

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1. Law, M. & Rudnicka, A.R. Statin safety: a systematic review. *Am. J. Cardiol.* **97**, 52C–60C (2006).
2. Sathasivam, S. Statin induced myotoxicity. *Eur. J. Intern. Med.* **23**, 317–324 (2012).
3. Stone, N.J. *et al.* 2013 ACC/AHA Guideline on the Treatment of Blood Cholesterol to Reduce Atherosclerotic Cardiovascular Risk in Adults: A Report of the American College of Cardiology/American Heart Association Task Force on Practice Guidelines. *Circulation* 2013; e-pub ahead of print 12 November 2013.
4. Graham, D.J. *et al.* Incidence of hospitalized rhabdomyolysis in patients treated with lipid-lowering drugs. *JAMA* **292**, 2585–2590 (2004).
5. Staffa, J.A., Chang, J. & Green, L. Cerivastatin and reports of fatal rhabdomyolysis. *N. Engl. J. Med.* **346**, 539–540 (2002).
6. Floyd, J.S., Heckbert, S.R., Weiss, N.S., Carrell, D.S. & Psaty, B.M. Use of administrative data to estimate the incidence of statin-related rhabdomyolysis. *JAMA* **307**, 1580–1582 (2012).
7. Pasternak, R.C., Smith, S.C. Jr, Bairey-Merz, C.N., Grundy, S.M., Cleeman, J.I. & Lenfant, C.; American College of Cardiology; American Heart Association; National Heart, Lung and Blood Institute. ACC/AHA/NHLBI Clinical Advisory on the Use and Safety of Statins. *Circulation* **106**, 1024–1028 (2002).
8. Aithal, G.P. & Daly, A.K. Preempting and preventing drug-induced liver injury. *Nat. Genet.* **42**, 650–651 (2010).
9. Carr, D.F. *et al.* SLCO1B1 genetic variant associated with statin-induced myopathy: a proof-of-concept study using the clinical practice research datalink. *Clin. Pharmacol. Ther.* **94**, 695–701 (2013).
10. Chang, C.H., Kusama, M., Ono, S., Sugiyama, Y., Orii, T. & Akazawa, M. Assessment of statin-associated muscle toxicity in Japan: a cohort study conducted using claims database and laboratory information. *BMJ Open* **3**, (2013).
11. Pirmohamed, M., Aithal, G.P., Behr, E., Daly, A. & Roden, D. The phenotype standardization project: improving pharmacogenetic studies of serious adverse drug reactions. *Clin. Pharmacol. Ther.* **89**, 784–785 (2011).
12. Sai, K. *et al.* Development of a detection algorithm for statin-induced myopathy using electronic medical records. *J. Clin. Pharm. Ther.* **38**, 230–235 (2013).
13. Aithal, G.P. *et al.* Case definition and phenotype standardization in drug-induced liver injury. *Clin. Pharmacol. Ther.* **89**, 806–815 (2011).
14. Behr, E.R. *et al.* The International Serious Adverse Events Consortium (ISAEC) phenotype standardization project for drug-induced torsades de pointes. *Eur. Heart J.* **34**, 1958–1963 (2013).
15. Pirmohamed, M. *et al.* Phenotype standardization for immune-mediated drug-induced skin injury. *Clin. Pharmacol. Ther.* **89**, 896–901 (2011).
16. Agbabiaka, T.B., Savović, J. & Ernst, E. Methods for causality assessment of adverse drug reactions: a systematic review. *Drug Saf.* **31**, 21–37 (2008).
17. Cham, S., Evans, M.A., Denenberg, J.O. & Golomb, B.A. Statin-associated muscle-related adverse effects: a case series of 354 patients. *Pharmacotherapy* **30**, 541–553 (2010).
18. de Lemos, J.A. *et al.*; Investigators. Early intensive vs a delayed conservative simvastatin strategy in patients with acute coronary syndromes: phase Z of the A to Z trial. *JAMA* **292**, 1307–1316 (2004).
19. Hansen, K.E., Hildebrand, J.P., Ferguson, E.E. & Stein, J.H. Outcomes in 45 patients with statin-associated myopathy. *Arch. Intern. Med.* **165**, 2671–2676 (2005).
20. Kashani, A. *et al.* Risks associated with statin therapy: a systematic overview of randomized clinical trials. *Circulation* **114**, 2788–2797 (2006).
21. Molokhia, M., McKeigue, P., Curcin, V. & Majeed, A. Statin induced myopathy and myalgia: time trend analysis and comparison of risk associated with statin class from 1991–2006. *PLoS One* **3**, e2522 (2008).
22. Oshima, Y. Characteristics of drug-associated rhabdomyolysis: analysis of 8,610 cases reported to the U.S. Food and Drug Administration. *Intern. Med.* **50**, 845–853 (2011).
23. Pfeffer, M.A. *et al.* Safety and tolerability of pravastatin in long-term clinical trials: prospective Pravastatin Pooling (PPP) Project. *Circulation* **105**, 2341–2346 (2002).
24. Ridker, P.M. *et al.*; JUPITER Study Group. Rosuvastatin to prevent vascular events in men and women with elevated C-reactive protein. *N. Engl. J. Med.* **359**, 2195–2207 (2008).
25. Cziraky, M.J. *et al.* Risk of hospitalized rhabdomyolysis associated with lipid-lowering drugs in a real-world clinical setting. *J. Clin. Lipidol.* **7**, 102–108 (2013).

26. Gilad, R. & Lampl, Y. Rhabdomyolysis induced by simvastatin and ketoconazole treatment. *Clin. Neuropharmacol.* **22**, 295–297 (1999).
27. Meriglioli, M.N., Barboi, A.C., Rowin, J. & Cochran, E.J. HMG-CoA Reductase Inhibitor Myopathy: Clinical, Electrophysiological, and Pathologic Data in Five Patients. *J. Clin. Neuromuscul. Dis.* **2**, 129–134 (2001).
28. Phillips, P.S. *et al.*; Scripps Mercy Clinical Research Center. Statin-associated myopathy with normal creatine kinase levels. *Ann. Intern. Med.* **137**, 581–585 (2002).
29. Pierce, L.R., Wysowski, D.K. & Gross, T.P. Myopathy and rhabdomyolysis associated with lovastatin-gemfibrozil combination therapy. *JAMA* **264**, 71–75 (1990).
30. Hohenegger, M. Drug induced rhabdomyolysis. *Curr. Opin. Pharmacol.* **12**, 335–339 (2012).
31. Elhayany, A., Mishaal, R.A. & Vinker, S. Is there clinical benefit to routine enzyme testing of patients on statins? *Expert Opin. Drug Saf.* **11**, 185–190 (2012).
32. Fung, E.C. & Crook, M.A. Statin myopathy: a lipid clinic experience on the tolerability of statin rechallenge. *Cardiovasc. Ther.* **30**, e212–e218 (2012).
33. Lemstra, M., Blackburn, D., Crawley, A. & Fung, R. Proportion and risk indicators of nonadherence to statin therapy: a meta-analysis. *Can. J. Cardiol.* **28**, 574–580 (2012).
34. Blaier, O., Lishner, M. & Elis, A. Managing statin-induced muscle toxicity in a lipid clinic. *J. Clin. Pharm. Ther.* **36**, 336–341 (2011).
35. Link, E. *et al.*; SEARCH Collaborative Group. SLCO1B1 variants and statin-induced myopathy—a genomewide study. *N. Engl. J. Med.* **359**, 789–799 (2008).
36. Wilke, R.A. *et al.*; Clinical Pharmacogenomics Implementation Consortium (CPIC). The clinical pharmacogenomics implementation consortium: CPIC guideline for SLCO1B1 and simvastatin-induced myopathy. *Clin. Pharmacol. Ther.* **92**, 112–117 (2012).
37. Brunham, L.R. *et al.* Differential effect of the rs4149056 variant in SLCO1B1 on myopathy associated with simvastatin and atorvastatin. *Pharmacogenomics J.* **12**, 233–237 (2012).
38. Donnelly, L.A. *et al.* Common nonsynonymous substitutions in SLCO1B1 predispose to statin intolerance in routinely treated individuals with type 2 diabetes: a go-DARTS study. *Clin. Pharmacol. Ther.* **89**, 210–216 (2011).
39. Donnelly, L.A. *et al.* Robust association of the LPA locus with low-density lipoprotein cholesterol lowering response to statin treatment in a meta-analysis of 30 467 individuals from both randomized control trials and observational studies and association with coronary artery disease outcome during statin treatment. *Pharmacogenet. Genomics* **23**, 518–525 (2013).
40. Linde, R., Peng, L., Desai, M. & Feldman, D. The role of vitamin D and SLCO1B1\*5 gene polymorphism in statin-associated myalgias. *Dermatoendocrinol.* **2**, 77–84 (2010).
41. Marcianti, K.D. *et al.* Cerivastatin, genetic variants, and the risk of rhabdomyolysis. *Pharmacogenet. Genomics* **21**, 280–288 (2011).
42. Voora, D. *et al.* The SLCO1B1\*5 genetic variant is associated with statin-induced side effects. *J. Am. Coll. Cardiol.* **54**, 1609–1616 (2009).
43. Newman, C., Tsai, J., Szarek, M., Luo, D. & Gibson, E. Comparative safety of atorvastatin 80 mg versus 10 mg derived from analysis of 49 completed trials in 14,236 patients. *Am. J. Cardiol.* **97**, 61–67 (2006).
44. Antons, K.A., Williams, C.D., Baker, S.K. & Phillips, P.S. Clinical perspectives of statin-induced rhabdomyolysis. *Am. J. Med.* **119**, 400–409 (2006).
45. Black, C. & Jick, H. Etiology and frequency of rhabdomyolysis. *Pharmacotherapy* **22**, 1524–1526 (2002).
46. Needham, M., Fabian, V., Knezevic, W., Panegyres, P., Zilko, P. & Mastaglia, F.L. Progressive myopathy with up-regulation of MHC-I associated with statin therapy. *Neuromuscul. Disord.* **17**, 194–200 (2007).
47. Mammen, A.L. *et al.* Autoantibodies against 3-hydroxy-3-methylglutaryl-coenzyme A reductase in patients with statin-associated autoimmune myopathy. *Arthritis Rheum.* **63**, 713–721 (2011).
48. Mammen, A.L. *et al.* Rarity of anti-3-hydroxy-3-methylglutaryl-coenzyme A reductase antibodies in statin users, including those with self-limited musculoskeletal side effects. *Arthritis Care Res. (Hoboken)*. **64**, 269–272 (2012).
49. Coward, W.R., Marei, A., Yang, A., Vasa-Nicotera, M.M. & Chow, S.C. Statin-induced proinflammatory response in mitogen-activated peripheral blood mononuclear cells through the activation of caspase-1 and IL-18 secretion in monocytes. *J. Immunol.* **176**, 5284–5292 (2006).
50. Bruckert, E., Hayem, G., Dejager, S., Yau, C. & Bégaud, B. Mild to moderate muscular symptoms with high-dosage statin therapy in hyperlipidemic patients—the PRIMO study. *Cardiovasc. Drugs Ther.* **19**, 403–414 (2005).
51. Grable-Espósito, P., Katzberg, H.D., Greenberg, S.A., Srinivasan, J., Katz, J. & Amato, A.A. Immune-mediated necrotizing myopathy associated with statins. *Muscle Nerve* **41**, 185–190 (2010).
52. Jones, P.H. & Davidson, M.H. Reporting rate of rhabdomyolysis with fenofibrate + statin versus gemfibrozil + any statin. *Am. J. Cardiol.* **95**, 120–122 (2005).
53. Backman, J.T., Kyrklund, C., Kivistö, K.T., Wang, J.S. & Neuvonen, P.J. Plasma concentrations of active simvastatin acid are increased by gemfibrozil. *Clin. Pharmacol. Ther.* **68**, 122–129 (2000).
54. Shitara, Y. & Sugiyama, Y. Pharmacokinetic and pharmacodynamic alterations of 3-hydroxy-3-methylglutaryl coenzyme A (HMG-CoA) reductase inhibitors: drug-drug interactions and interindividual differences in transporter and metabolic enzyme functions. *Pharmacol. Ther.* **112**, 71–105 (2006).
55. Kyrklund, C., Backman, J.T., Kivistö, K.T., Neuvonen, M., Laitila, J. & Neuvonen, P.J. Plasma concentrations of active lovastatin acid are markedly increased by gemfibrozil but not by bezafibrate. *Clin. Pharmacol. Ther.* **69**, 340–345 (2001).
56. Kyrklund, C., Backman, J.T., Neuvonen, M. & Neuvonen, P.J. Gemfibrozil increases plasma pravastatin concentrations and reduces pravastatin renal clearance. *Clin. Pharmacol. Ther.* **73**, 538–544 (2003).
57. Schneck, D.W. *et al.* The effect of gemfibrozil on the pharmacokinetics of rosuvastatin. *Clin. Pharmacol. Ther.* **75**, 455–463 (2004).
58. Ozdemir, O., Boran, M., Gökçe, V., Uzun, Y., Koçak, B. & Korkmaz, S. A case with severe rhabdomyolysis and renal failure associated with cerivastatin-gemfibrozil combination therapy—a case report. *Angiology* **51**, 695–697 (2000).
59. Ray, G.M. Antiretroviral and statin drug-drug interactions. *Cardiol. Rev.* **17**, 44–47 (2009).
60. Sorokin, A.V., Duncan, B., Panetta, R. & Thompson, P.D. Rhabdomyolysis associated with pomegranate juice consumption. *Am. J. Cardiol.* **98**, 705–706 (2006).
61. Zhou, S., Chan, E., Li, X. & Huang, M. Clinical outcomes and management of mechanism-based inhibition of cytochrome P450 3A4. *Ther. Clin. Risk Manag.* **1**, 3–13 (2005).
62. Zhou, S. *et al.* Mechanism-based inhibition of cytochrome P450 3A4 by therapeutic drugs. *Clin. Pharmacokinet.* **44**, 279–304 (2005).
63. Mazokopakis, E.E. Unusual causes of rhabdomyolysis. *Intern. Med. J.* **38**, 364–367 (2008).
64. Dirks, A.J. & Jones, K.M. Statin-induced apoptosis and skeletal myopathy. *Am. J. Physiol. Cell Physiol.* **291**, C1208–C1212 (2006).
65. Sinzinger, H. & O'Grady, J. Professional athletes suffering from familial hypercholesterolaemia rarely tolerate statin treatment because of muscular problems. *Br. J. Clin. Pharmacol.* **57**, 525–528 (2004).
66. O'Meara, H. *et al.* Electronic health records for biological sample collection: feasibility study of statin-induced myopathy using the Clinical Practice Research Datalink. *Br. J. Clin. Pharmacol.* **77**, 831–838 (2014).
67. Bays, H. Statin safety: an overview and assessment of the data—2005. *Am. J. Cardiol.* **97**, 6C–26C (2006).
68. Moghadasian, M.H., Mancini, G.B. & Frohlich, J.J. Pharmacotherapy of hypercholesterolaemia: statins in clinical practice. *Expert Opin. Pharmacother.* **1**, 683–695 (2000).
69. Heart Protection Study Collaborative Group. MRC/BHF Heart Protection Study of cholesterol lowering with simvastatin in 20,536 high-risk individuals: a randomised placebo-controlled trial. *Lancet* **360**, 7–22 (2002).
70. Mohassel, P. & Mammen, A.L. Statin-associated autoimmune myopathy and anti-HMGCR autoantibodies. *Muscle Nerve* **48**, 477–483 (2013).



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