

University of Dundee

Synthesis, molecular modeling and NAD(P)H

Ghorab, Mostafa M.; Alsaïd, Mansour S.; Higgins, Maureen; Dinkova-Kostova, Albena T.; Shahat, Abdelaaty A.; Elghazawy, Nehal H.

Published in:
Journal of Enzyme Inhibition and Medicinal Chemistry

DOI:
[10.3109/14756366.2016.1158714](https://doi.org/10.3109/14756366.2016.1158714)

Publication date:
2016

Document Version
Peer reviewed version

[Link to publication in Discovery Research Portal](#)

Citation for published version (APA):

Ghorab, M. M., Alsaïd, M. S., Higgins, M., Dinkova-Kostova, A. T., Shahat, A. A., Elghazawy, N. H., & Arafa, R. K. (2016). Synthesis, molecular modeling and NAD(P)H: quinone oxidoreductase 1 inducer activity of novel 2-phenylquinazolin-4-amine derivatives. *Journal of Enzyme Inhibition and Medicinal Chemistry*, 31(6), 1612-1618. <https://doi.org/10.3109/14756366.2016.1158714>

General rights

Copyright and moral rights for the publications made accessible in Discovery Research Portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from Discovery Research Portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain.
- You may freely distribute the URL identifying the publication in the public portal.

Take down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Synthesis, molecular modeling and NAD(P)H:quinone oxidoreductase 1 inducer activity of novel 2-phenylquinazolin-4-amine derivatives

Mostafa M. Ghorab^{1,2*}, Mansour S. Alsaïd¹, Maureen Higgins³, Albena T. Dinkova-Kostova^{3,4}, Abdelaaty A. Shahat^{1,5}, Nehal H. Elghazawy⁶, Reem K. Arafa^{6,7}

¹Department of Pharmacognosy, College of Pharmacy, King Saud University, P.O. Box 2457, Riyadh 11451, Kingdom of Saudi Arabia; ²Department of Drug Radiation Research, National Center for Radiation Research & Technology, Atomic Energy Authority, P.O.Box 29 Nacer City, Cairo, Egypt; ³Jacqui Wood Cancer Centre, Division of Cancer Research, Medical Research Institute, University of Dundee, Dundee DD1 9SY, United Kingdom; ⁴Departments of Medicine and Pharmacology and Molecular Sciences, Johns Hopkins University School of Medicine, Baltimore, MD 21205, USA; ⁵Phytochemistry Department National Research Center; ⁶Zewail City of Science and Technology, 12588, Cairo, Egypt; ⁷Department of Pharmaceutical Chemistry, Faculty of Pharmacy, Cairo University, 11562, Cairo, Egypt.

Abstract

Reactive oxygen species (ROS) play an integral role in the pathogenesis of most diseases. This work presents the design and synthesis of novel 2-phenylquinazolin-4-amine derivatives (**2-12**) and evaluation of their NAD(P)H:quinone oxidoreducace 1 (NQO1) inducer activity in murine cells. Also, molecular docking of all the new compounds was performed to assess their ability to inhibit Keap1-Nrf2 protein-protein interaction through occupying the Keap1 Nrf2-binding domain which biologically leads to a consequent Nrf2 accumulation and enhanced gene expression of NQO1. Docking results showed that all compounds have the ability to interact with Keap1; however compound **7**, the most active compound in this study, showed more interactions with key amino acids.

Keywords: 2-phenylquinazolin-4-amine derivatives, molecular modeling, Keap1/Nrf2, cytoprotection, NQO1 induction.

Introduction

Excessive unbalanced production of reactive oxygen species (ROS), namely superoxide anion radical ($O_2^{\cdot-}$), singlet oxygen (1O_2), hydroxyl radical ($\cdot OH$) and hydrogen peroxide (H_2O_2), has been well established to be associated with many pathological conditions including atherosclerosis, stroke, diabetes, Alzheimer's disease, cancer and inflammation¹. In a counter action and to maintain homeostasis, living cells combat the deleterious effects of ROS by an arsenal of biochemical antioxidants.^{2,3} Under oxidative stress conditions; where the balance between ROS and biochemical antioxidants is lost due to over production of the former or under availability of the latter; ROS can attack and damage cellular membrane lipids, DNA nucleotides⁴, proteins' sulphhydryl groups⁵ and ribonucleoproteins⁶. Among the most important antioxidant mechanisms activated under conditions of oxidative stress is the Keap1-Nrf2-AREs pathway⁷. This pathway is associated with the expression of more than 100 cytoprotective genes and their respective enzymes, one of which is NAD(P)H:quinone oxidoreductase 1 (NQO1, EC 1.6.99.2). Under homeostatic conditions, Nrf2 is continuously targeted for ubiquitination and proteasomal degradation by Keap1. However, upon increased oxidative load, reactive cysteine residues within Keap1 serve as sensors for oxidants, leading to loss of its ability to target Nrf2 for degradation, subsequent accumulation of the transcription factor, activation of the pathway and increased transcription of cytoprotective antioxidant proteins⁸⁻¹⁰.

Medicinal chemists view the Keap1-Nrf2 protein-protein interaction as a critical point of the pathway that can be targeted for intervention and for potential management of a variety of oxidative stress-related ailments like cancer, Parkinson's, Alzheimer's and diabetes¹¹⁻¹⁴. Design of non-covalent small molecule modulators of the Keap-Nrf2 interaction has been widely explored^{15,16}. Quinazoline-based nitrogenous heteroaryls

are an excellent reservoir of bioactive substances where a number of biological activities have been associated with antioxidant activity¹⁷⁻¹⁹.

In continuation of our quest for new cytoprotective agents^{20,21}, we herein report the synthesis, NQO1 inducer activity and Keap1 inhibitory in silico screening results for a novel class of N-aryl substituted 4-amino-2-phenylquinazolines.

Materials and Methods

Chemistry

Melting points (uncorrected) were determined in open capillary on a Gallen Kamp melting point apparatus (Sanyo Gallen Kamp, UK). Precoated silica gel plates (Kieselgel 0.25 mm, 60 F254, Merck, Germany) were used for thin layer chromatography. A developing solvent system of chloroform/methanol (8:2) was used and the spots were detected by ultraviolet light. IR spectra (KBr disc) were recorded using an FT-IR spectrophotometer (Perkin Elmer, USA). NMR spectra were scanned on a NMR spectrophotometer (Bruker AXS Inc., Switzerland), operating at 500 MHz for ¹H and 125.76 MHz for ¹³C. Chemical shifts are expressed in δ -values (ppm) relative to TMS as an internal standard, using DMSO-*d*₆ as a solvent. Elemental analyses were done on a model 2400 CHNSO analyzer (Perkin Elmer, USA). All the values were within \pm 0.4 % of the theoretical values. All reagents used were of AR grades. The starting material 4-chloro-2-phenylquinazoline (**1**) was purchased from sigma (USA) and was directly used for the preparation of target compounds.

Synthesis of 2-Phenyl-4-substituted quinazoline derivatives 2-12.

General procedure

A mixture of 2-phenyl 4-chloroquinazoline **1** (2.40 gm, 0.01 mol) and different amines (0.01 mol) in dimethylformamide (10 mL) containing a catalytic amount of trimethylamine (3 drops) was refluxed for 24 h. then left to cool. The solid product formed was collected by filtration and recrystallized from acetic acid to give the desired quinazoline derivatives **2-12**, respectively.

N-(2,3-Dihydro-1H-inden-5-yl)-2-phenylquinazolin-4-amine (**2**).

White solid; yield, 76%; m.p. 118-9 °C. IR (KBr, cm⁻¹): 3401 (NH), 3080 (CH arom.), 2976, 2778 (CH aliph.), 1631 (C=N). ¹H-NMR (DMSO-*d*₆): ppm 1.9-2.8 [m, 6H, 3CH₂, Cyclo], 7.1-9.0 [m, 12H, Ar-H], 11.8 [s, 1H, NH, exchangeable with D₂O]. ¹³C-NMR (DMSO-*d*₆): ppm 25.6, 32.9 (2), 113.1, 121.6, 123.0, 124.4, 125.4, 128.3 (2), 129.3, 129.4, 129.7, 129.8, 133.7 (2), 135.3, 136.1, 142.3, 144.5 (2), 157.3, 159.1. MS m/z (%): 337 (M⁺) (16.73), 219 (100). Anal. Calcd. For C₂₃H₁₉N₃ (337): C, 81.87; H, 5.68; N, 12.45. Found: C, 81.53; H, 5.94; N, 12.14.

N-(Benzo[*d*][1,3]dioxol-5-yl)-2-phenylquinazolin-4-amine (**3**).

Pale yellow solid; yield, 89%; m.p. 107-8 °C. IR (KBr, cm⁻¹): 3441 (NH), 3081 (CH arom.), 2984, 2874 (CH aliph.), 1617 (C=N). ¹H-NMR (DMSO-*d*₆): ppm 6.0 [s, 2H, O-CH₂-O], 7.0-8.5 [m, 12H, Ar-H], 9.7 [s, 1H, NH, exchangeable with D₂O]. ¹³C-NMR (DMSO-*d*₆): ppm 101.4, 104.8, 108.6, 114.4, 115.8, 123.3, 126.2 (2), 128.3, 128.5, 128.8 (2), 130.6, 133.5, 133.9, 138.8 (2), 147.4, 150.8, 159.6, 162.7. MS m/z (%): 341 (M⁺) (3.65), 294 (100). Anal. Calcd. For C₂₁H₁₅N₃O₂ (341): C, 73.89; H, 4.43; N, 12.31. Found: C, 74.11; H, 4.75; N, 12.65.

N-(Benzo[*d*][1,3]dioxol-5-ylmethyl)-2-phenylquinazolin-4-amine (**4**).

Tan white solid; yield, 80%; m.p. 180-1 °C. IR (KBr, cm⁻¹): 3238 (NH), 3069 (CH arom.), 2944, 2791 (CH aliph.), 1618 (C=N). ¹H-NMR (DMSO-*d*₆): ppm 4.8 [s, 2H, CH₂], 5.9 [s, 2H, O-CH₂-O], 6.8-8.6 [m, 12H, Ar-H], 10.2 [s, 1H, NH, exchangeable with D₂O]. ¹³C-NMR (DMSO-*d*₆): ppm 44.5, 101.3, 108.4, 108.5, 113.4, 120.9, 124.2, 127.2 (2), 129.1 (2), 132.4 (2), 132.9, 133.3, 134.8 (2), 146.5, 146.7, 147.7, 158.4, 163.4. MS m/z (%): 355 (M⁺) (22.15), 233 (100). Anal. Calcd. For C₂₂H₁₇N₃O₂ (355): C, 74.35; H, 4.82; N, 11.82. Found: C, 74.08; H, 4.59; N, 12.10.

2-Phenyl-N-(5,6,7,8-tetrahydronaphthalen-1-yl)quinazolin-4-amine (5).

White solid; yield, 77%; m.p. 192-3 °C. IR (KBr, cm^{-1}): 3412 (NH), 3068 (CH arom.), 2922, 2782 (CH aliph.), 1609 (C=N). $^1\text{H-NMR}$ ($\text{DMSO-}d_6$): ppm 1.6-2.8 [m, 8H, 4 CH_2 Cyclo], 7.0-9.0 [m, 12H, Ar-H], 11.4 [s, 1H, NH, exchangeable with D_2O]. $^{13}\text{C-NMR}$ ($\text{DMSO-}d_6$): ppm 22.6 (2), 25.0, 29.6, 113.0 (2), 125.1, 125.3, 125.8, 128.0, 128.7 (2), 129.1 (2), 129.3 (2), 133.1, 134.1, 135.7, 135.8, 138.4, 157.7, 160.1, 163.4. MS m/z (%): 351 (M^+) (7.13), 294 (100). Anal. Calcd. For $\text{C}_{24}\text{H}_{21}\text{N}_3$ (351): C, 82.02; H, 6.02; N, 11.96. Found: C, 82.34; H, 5.81; N, 12.21.

N-(Naphthalen-1-yl)-phenylquinazolin-4-amine (6).

White solid; yield, 79%; m.p. 143-5 °C. IR (KBr, cm^{-1}): 3401 (NH), 3100 (CH arom.), 2981, 2780 (CH aliph.), 1629 (C=N). $^1\text{H-NMR}$ ($\text{DMSO-}d_6$): ppm 7.1-8.9 [m, 16H, Ar-H], 11.2 [s, 1H, NH, exchangeable with D_2O]. $^{13}\text{C-NMR}$ ($\text{DMSO-}d_6$): ppm 113.7, 119.4, 122.4, 122.8, 124.1, 124.7, 125.2, 126.0, 126.2, 126.5 (2), 127.3, 127.4, 128.2, 128.7, 129.0, 130.0, 131.8, 133.1, 134.1, 135.0, 158.6, 160.8, 163.4. MS m/z (%): 347 (M^+) (11.23), 269 (100). Anal. Calcd. For $\text{C}_{24}\text{H}_{17}\text{N}_3$ (347): C, 82.97; H, 4.93; N, 12.10. Found: C, 82.64; H, 5.21; N, 11.86.

2-Phenyl-N-(quinolin-2-yl)quinazolin-4-amine (7).

White solid; yield, 69%; m.p. 191-2 °C. IR (KBr, cm^{-1}): 3411 (NH), 3024 (CH arom.), 2977, 2839, 2780 (CH aliph.), 1635 (C=N). $^1\text{H-NMR}$ ($\text{DMSO-}d_6$): ppm 7.1-8.9 [m, 15H, Ar-H], 9.1 [s, 1H, NH, exchangeable with D_2O]. $^{13}\text{C-NMR}$ ($\text{DMSO-}d_6$): ppm 112.6, 116.7, 117.2, 123.1, 126.6 (2), 127.7, 127.8 (2), 128.0 (2), 129.2 (2), 129.3, 129.5, 129.8, 133.1, 135.4, 138.4 (2), 157.1, 161.7, 164.8. MS m/z (%): 348 (M^+) (24.07), 209 (100). Anal. Calcd. For $\text{C}_{23}\text{H}_{16}\text{N}_4$ (348): C, 79.29; H, 4.63; N, 16.08. Found: C, 79.55; H, 4.38; N, 16.36.

2-Phenyl-N-(quinolin-3-yl)quinazolin-4-amine (8).

White solid; yield, 91%; m.p. 281-2 °C. IR (KBr, cm^{-1}): 3321 (NH), 3059 (CH arom.), 1624 (C=N). $^1\text{H-NMR}$ (DMSO- d_6): ppm 7.4-9.3 [m, 15H, Ar-H], 10.3 [s, 1H, NH exchangeable with D_2O]. $^{13}\text{C-NMR}$ (DMSO- d_6): ppm 114.4, 121.4, 124.8, 125.6, 126.3, 126.5, 127.0 (2), 127.9, 128.1, 128.4, 128.9 (2), 129.1, 130.2, 131.8, 132.2, 133.9, 138.2, 144.7, 149.2, 162.7, 163.2. MS m/z (%): 348 (M^+) (39.41), 270 (100). Anal. Calcd. For $\text{C}_{23}\text{H}_{16}\text{N}_4$ (348): C, 79.29; H, 4.63; N, 16.08. Found: C, 79.56; H, 4.32; N, 16.29.

N-(2-Methylquinolin-4-yl)-2-phenylquinazolin-4-amine (**9**).

White solid; yield, 88%; m.p. 236-8 °C. IR (KBr, cm^{-1}): 3433 (NH), 3023 (CH arom.), 2976, 2839, 2779 (CH aliph.), 1609 (C=N). $^1\text{H-NMR}$ (DMSO- d_6): ppm 2.5 [s, 3H, CH_3], 7.5-8.0 [m, 14H, Ar-H], 12.5 [s, 1H, NH exchangeable with D_2O]. $^{13}\text{C-NMR}$ (DMSO- d_6): ppm 19.8, 102.0, 115.5, 119.7, 121.4, 124.2, 126.1, 126.4 (2), 127.0, 127.9, 128.2, 128.9 (2), 129.7, 131.8, 133.1, 133.9, 149.1, 152.8, 153.2, 158.1, 162.7 (2). MS m/z (%): 362(M^+) (24.09), 347 (100). Anal. Calcd. For $\text{C}_{24}\text{H}_{18}\text{N}_4$ (362): C, 79.54; H, 5.01; N, 15.46. Found: C, 79.22; H, 4.87; N, 15.18.

4-Methyl-7-(2-phenylquinazolin-4-ylamino)-2H-chromen-2-one (**10**).

Orange solid; yield, 78%; m.p. 279-80 °C. IR (KBr, cm^{-1}): 3356 (NH), 3062 (CH arom.), 2946, 2869, 2765 (CH aliph.), 1699 (C=O), 1618 (C=N). $^1\text{H-NMR}$ (DMSO- d_6): ppm 2.4 [s, 3H, CH_3], 6.2 [s, 1H, CH Chromene], 7.5-8.6 [m, 12H, Ar-H], 10.1 [s, 1H, NH exchangeable with D_2O]. $^{13}\text{C-NMR}$ (DMSO- d_6): ppm 18.5, 108.1 (2), 112.7, 114.5, 115.3, 123.5, 125.9, 126.7 (2), 128.3, 128.7, 129.0 (2), 130.9, 134.0, 138.6, 143.3, 151.0, 153.6, 154.0, 158.1, 159.3, 160.6. MS m/z (%): 379 (M^+) (9.64), 294 (100). Anal. Calcd. For $\text{C}_{24}\text{H}_{17}\text{N}_3\text{O}_2$ (379): C, 75.97; H, 4.52; N, 11.08. Found: C, 75.61; H, 4.87; N, 10.78.

7-(2-Phenylquinazolin-4-ylamino)-4-(trifluoromethyl)-2H-chromen-2-one (**11**).

Yellow solid; yield, 72%; m.p. 255-7 °C. IR (KBr, cm^{-1}): 3355 (NH), 3058 (CH arom.), 1697 (C=O), 1617 (C=N). $^1\text{H-NMR}$ (DMSO- d_6): ppm 6.2 [s, 1H, CH Chromene], 7.2-8.6 [m, 12H, Ar-H], 10.1 [s, 1H, NH exchangeable with D_2O]. $^{13}\text{C-NMR}$ (DMSO- d_6): ppm 106.3, 108.1, 112.6, 112.9, 114.5, 121.4, 123.5, 124.7, 125.9 (2), 126.5, 127.0, 128.3 (2), 129.0, 130.9,

133.1, 143.5, 149.2, 154.0, 158.1, 160.6, 160.7, 162.6. MS m/z (%): 433 (M⁺) (1.98), 355 (100). Anal. Calcd. For C₂₄H₁₄F₃N₃O₂ (433): C, 66.51; H, 3.26; N, 9.70. Found: C, 66.21; H, 3.54; N, 9.98.

N-(4-Morpholinophenyl)-2-phenylquinazolin-4-amine (**12**).

White solid; yield, 86%; m.p. 228-9 °C. IR (KBr, cm⁻¹): 3334 (NH), 3059 (CH arom.), 2977, 2891, 2860 (CH aliph.), 1617 (C=N). ¹H-NMR (DMSO-*d*₆): ppm 2.8-3.7 [m, 8H, 4CH₂ Cyclo], 7.0-8.5 [m, 13H, Ar-H], 9.9 [s, 1H, NH exchangeable with D₂O]. ¹³C-NMR (DMSO-*d*₆): ppm 49.2 (2), 66.6 (2), 114.5 (2), 115.4, 123.4 (2), 126.1, 128.3 (2), 128.5 (2), 128.8 (2), 130.6, 131.6, 133.4 (2), 138.9, 148.0, 159.6, 162.7. MS m/z (%): 382 (M⁺) (12.23), 295 (100). Anal. Calcd. For C₂₄H₂₂N₄O (382): C, 75.37; H, 5.80; N, 14.65. Found: C, 75.04; H, 5.44; N, 14.31.

Biological evaluation

Hepa1c1c7 murine hepatoma cells were maintained in a humidified atmosphere at 37 °C, 5% CO₂ in α-MEM supplemented with 10% (v/v) fetal bovine serum. Cells (10⁴ per well) were grown for 24 h in 96-well plates. For evaluation of the potential NQO1 inducer activity of the new 2-phenylquinazolin-4-amines **2-12**, the medium was replaced with inducer-containing fresh medium, and the cells were further grown for 48 h. There were eight replicates of each treatment of eight different concentrations of inducers. All compounds were dissolved in DMSO, and diluted in the cell culture medium at a ratio of 1:1000 prior to screening experiments. The final solvent concentration in the medium was 0.1% (v/v) in all wells. At the end of the treatment period, cell lysates were prepared in digitonin and the specific activity of NQO1 was determined using menadione as a substrate as previously described²². The Concentration which Doubles the specific activity of NQO1 (CD value) was used as a measure of inducer potency.

Mean values for the eight replicate wells are shown for each data point. The standard deviation for each data point was within 5% of the value.

Molecular modeling study

The 3D molecular models of the newly synthesized phenylquinazolinamine derivatives were built according to the default parameters, with the MOE software suite version 10.2009. Geometry optimization as well as a systematic conformational search was carried out to an RMS gradient of 0.01 Å employing the ConfSearch module implemented in MOE. All computations were performed with the Merck Force Field (MMFF94s). To evaluate our compounds' ability to access and block the Nrf2-binding site of Keap1, a molecular docking study was performed using the crystallographic structure of the Kelch domain of Keap1 obtained from the Protein Data Bank (PDB ID: 4IQK). The protein target was prepared for docking by addition of the missing hydrogens and calculating the partial charges. Internal validation was performed to the native ligand followed by docking of the compounds where the target protein was kept rigid, while ligands were allowed to rotate to accommodate freely inside the protein cavity. Following multiple separate docking simulations using default parameters, the best conformations were chosen based on the combination of S score data, E conformation and appropriate fitting with the relevant amino acids in the binding pocket.

Results and discussion

Chemistry

The aim of this work was to design and synthesize a novel series of quinazolines bearing biologically active heterocyclic moieties to evaluate and explore their NQO1 inducer activity. Therefore, 4-aminoquinazoline derivatives **2-12** were synthesized starting from 4-chloro-2-phenylquinazoline (**1**) as depicted in Scheme 1. Compounds **2-12** were obtained in good yield via reaction of **1** with different amines, namely 5-aminoindane, 3,4-(methylenedioxy)aniline, pipronylamine, 5,6,7,8-tetrahydro-1-naphthylamine, 1-naphthylamine, 2-aminoquinoline, 3-aminoquinoline, 4-aminoquinaldine, 7-amino-4-

methyl-coumarin, 7-amino-4-(trifluoromethyl) coumarin, and 4-morpholinoaniline in dry dimethyl formamide (Scheme 1). All the synthesized compounds were established on basis of elemental analyses, IR, $^1\text{H-NMR}$, $^{13}\text{C-NMR}$ and mass spectral data. IR spectra of compounds **2-12** revealed the presence of the characteristic bands for the NH group ranging from 3441 to 3238 cm^{-1} , the aromatic CH around 3100 to 3023 cm^{-1} , the C=N group from 1635 to 1609 cm^{-1} . $^1\text{H-NMR}$ spectra of compounds **2-12** exhibited signals at δ 12.5-9.1 ppm attributed to the NH group. $^1\text{H-NMR}$ spectrum of compound **2** revealed a multiplet at δ 1.9-2.8 ppm assigned to the 3 CH_2 groups. $^{13}\text{C-NMR}$ spectrum of compound **2** exhibited signals at δ 25.6, 32.9, 32.9 ppm for 3 CH_2 groups. Also, $^1\text{H-NMR}$ spectrum of compound **3** showed a signal at δ 6.2 ppm for the O- CH_2 -O group while its $^{13}\text{C-NMR}$ spectrum revealed a singlet at δ 101.4 ppm for the same group. In addition, $^1\text{H-NMR}$ spectrum of compound **5** gave a multiplet at δ 1.6-2.8 ppm assigned to the 4 CH_2 groups. $^{13}\text{C-NMR}$ spectrum of compound **5** revealed signals at 22.6, 22.6, 25.0 and 29.6 ppm attributed to the aforementioned groups. On the other hand, $^1\text{H-NMR}$ spectrum of compound **10** revealed the presence of a singlet at 2.4 ppm for the CH_3 group and 6.2 ppm for the CH of chromene. $^{13}\text{C-NMR}$ spectrum of compound **10** showed a singlet at 18.5 ppm attributed to the CH_3 group, 112.7 ppm due to the chromene CH and 159.3 ppm for the C=O group. IR spectrum of **11** showed the presence of a characteristic band at 1697 cm^{-1} for the C=O. $^1\text{H-NMR}$ spectrum of compound **11** revealed the presence of a singlet at 6.2 ppm for the chromene CH group. $^{13}\text{C-NMR}$ spectrum of compound **11** showed signals at 112.6, 123.5 and 160.7 ppm for CH chromene, CF_3 and C=O groups, respectively. $^1\text{H-NMR}$ spectrum of compound **12** revealed the presence of a multiplet at 2.8-3.7 ppm for the morpholino 4 CH_2 groups. $^{13}\text{C-NMR}$ spectrum of compound **12** showed signals at 49.2, 66.6 ppm corresponding to the $\text{CH}_2\text{-N-CH}_2$ and the $\text{CH}_2\text{-O-CH}_2$ morpholino 4 CH_2 groups, respectively.

Biological evaluation

NQO1 inducer activity assessment represented in Figure 1 and Table 1 revealed that while the dihydroindenyl derivative **2** was devoid of NQO1 inducer activity, its dioxo

bioisostere compound **3** displayed relatively high potency (extrapolated CD = 0.5 μ M), however, this was of low magnitude and lacked dose-dependency. In addition, the N-benzo[d][1,3]dioxol-5-yl derivative **3** was cytostatic/cytotoxic at concentrations greater than 12.5 μ M as evidenced by the more than 50% lower levels of protein content in the lysates of treated cells. On the other hand, introduction of methylene spacer between the 4-amino group of the quinazoline and the benzo[d][1,3]dioxole ring gave a weakly active derivative **4** where a CD value wasn't reached. Regarding the tetrahydronaphthyl derivative **5** and the morpholinophenyl compound **12**, both displayed weak activity (a CD value was not reached), with no clear dose dependence. In contrast, the naphth-1-yl derivative **6** compound showed a moderate dose-dependent inducer activity (CD = 1 μ M). Furthermore, the 2-methyl-quinolin-4-yl analog **9** displayed a moderate inducer potency (CD = 4 μ M) which was dose-dependent, indicating that these modifications were not much in favor of activity as compared to the naphthyl bioisostere **6**. The bioisosteres **7** and **8** demonstrated a big difference in activity. Whereas, the quinolin-2-yl derivative **7** had the highest potency in this study (CD = 0.4 μ M), the 3 positional isomer **8** was more than twice less potent (CD = 0.9 μ M). Finally, with regards to the bioisosteric pair **10** and **11** (CD = 3 and 1.6 μ M, respectively) it was evident that replacement of the methyl group with a trifluoromethyl almost doubled the potency. Although almost similar in potency, compounds **6** (CD = 1 μ M) and **8** (CD = 0.9 μ M) were very different in terms of dose responsiveness. Thus, whereas induction was dose-dependent for compound **6**, it was not for compound **8**. Compound **7** was the most potent inducer (CD = 0.4 μ M), and importantly, had the most favorable inducer properties within this series.

Concentration (μM)	Compound #										
	2	3	4	5	6	7	8	9	10	11	12
0.09765	NR*	NR	NR	NR	NR	1.34	NR	NR	NR	NR	NR
0.1953	NR	NR	NR	NR	NR	1.49	NR	NR	NR	NR	NR
0.3906	NR	NR	1.27	NR	NR	1.81	NR	NR	NR	NR	NR
0.781	1.16	2.31	1.41	1.19	1.82	2.43	1.93	1.34	1.50	1.83	1.64

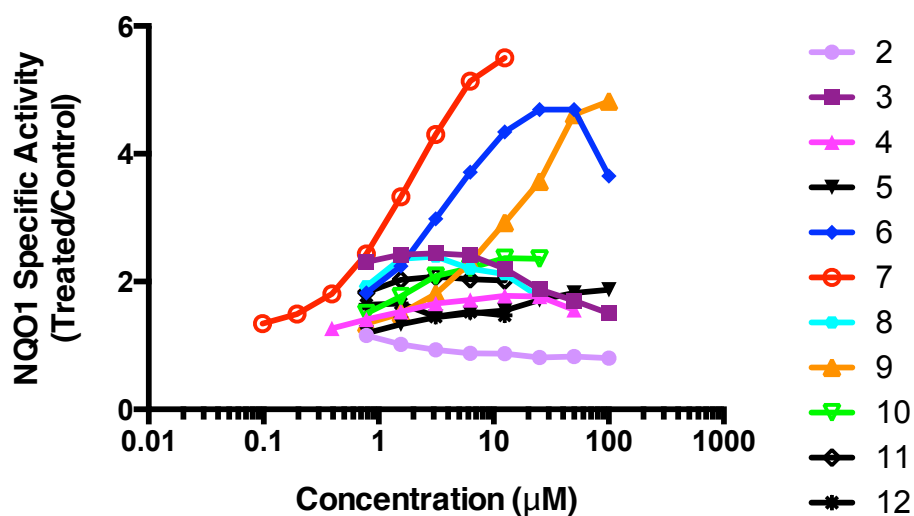


Figure 1: Concentration dependence of NQO1 inducer activity of quinazoline derivatives.

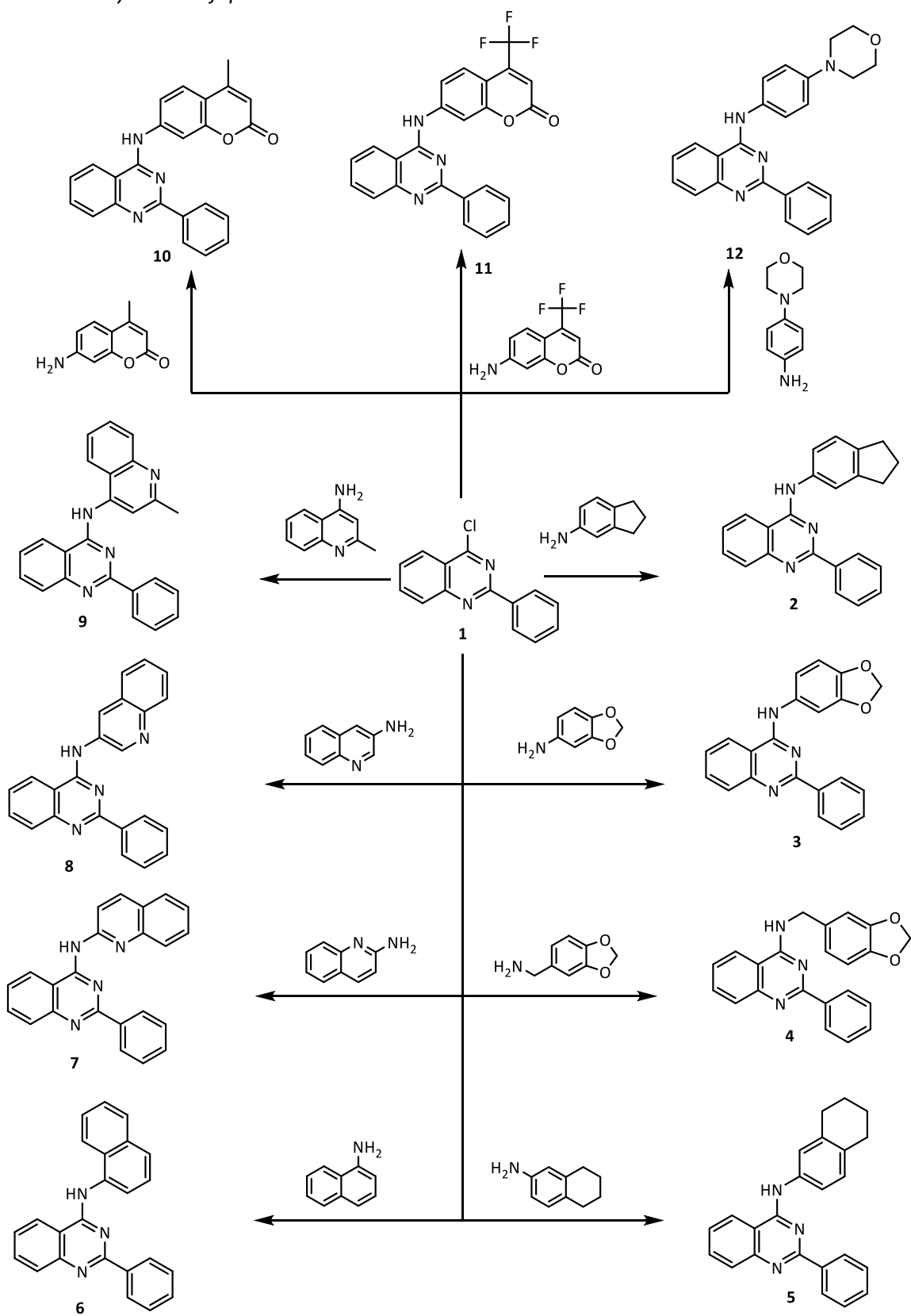
Table 1: NQO1 inducer activity and CD values of the test compounds.

1.563	1.02	2.42	1.53	1.33	2.24	3.33	2.36	1.5	1.77	2.03	1.66
3.125	0.93	2.44	1.66	1.44	2.98	4.3	2.39	1.81	2.09	2.08	1.45
6.25	0.88	2.41	1.71	1.5	3.71	5.14	2.2	2.3	2.21	2.04	1.53
12.5	0.87	2.2	1.78	1.55	4.34	5.5	2.12	2.92	2.37	2.02	1.47
25	0.81	1.88	1.77	1.71	4.69	NR	1.76	3.57	2.36	NR	NR
50	0.83	1.7	1.56	1.82	4.69	NR	NR	4.61	NR	NR	NR
100	0.8	1.5	NR	1.87	3.65	NR	NR	4.82	NR	NR	NR
CD**	NR	0.5	NR	NR	1	0.4	0.9	4	3	1.6	NR

*NR = not recorded

** CD data presented are the averages of 3 independent experiments, each with 8 replicates wells of cells, and SD (standard deviation) for each data point was within 5% of the value.

Scheme 1: Synthesis of quinazoline derivatives 2-12.



Molecular modeling study

Keap1 is known to bind to Nrf2 promoting its degradation, resulting in low levels of cytoprotective gene products. Various small molecule compounds were reported to bind to the Keap1 Kelch domain and antagonize its activity^{23,24,25}. For assessing the newly synthesized compounds' ability to access and block the Kelch domain of Keap1, a molecular docking study was performed using the Keap1 crystal coordinates obtained from the Protein Data Bank (PDB ID: 4IQK). The main interactions observed between the validated native ligand and the protein target were found to be arene-cation interaction with Arg415, arene-arene interaction with Tyr525 and 3 hydrogen bonds with Ser602, Ser508 and Ser555 with $S = -13.306$ Kcal/mol and rmsd 0.6635 Kcal/mol/Å (Figure 2). Upon docking of the synthesized compounds, they all showed an arene-cation binding interaction with Arg415 via their aromatic rings. Compound **11** ($S = -10.913$ Kcal/mol) has adopted a conformation allowing the presence of 3 arene-cation interactions with Arg415 and multiple rings of the ligand (Figures 3 and 4).

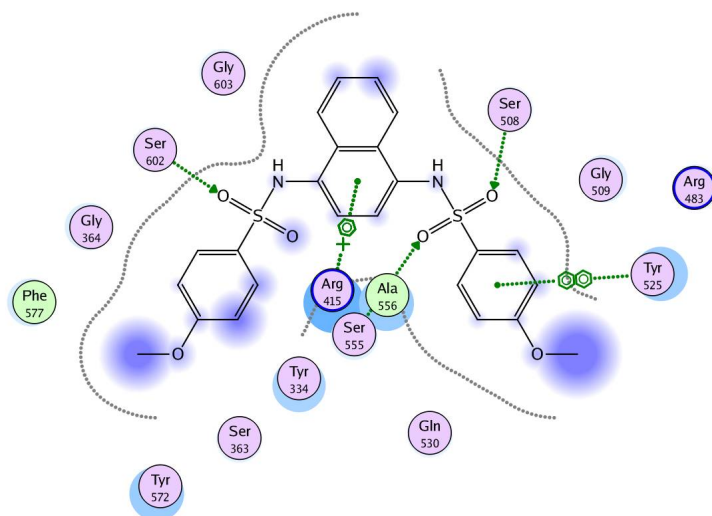


Figure 2: Interactions of the native ligand with the Kelch domain of Keap1 (PDB ID: 4IQK).

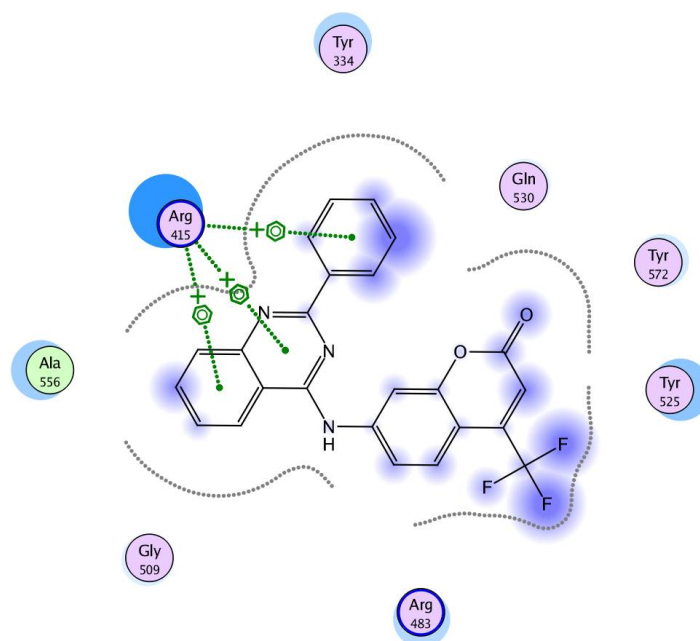


Figure 3: Interactions of compound **11** with the Kelch domain of Keap1 (PDB ID: 41QK).

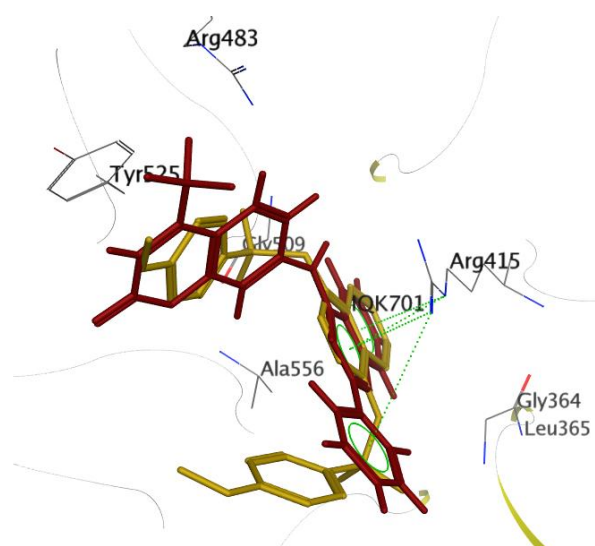


Figure 4: Overlap of Compound **11** (maroon) over native ligand (yellow) (PDB ID: 41QK)

Finally, compound **7** that showed the most potent NQO1 inducer activity (CD = 0.4 μ M) was the only compound that showed 3 arene-cation interactions with Arg415 and 2 arene-arene interactions with Tyr525 indicating a possible correlation between those multiple interactions and the noted high potency ($S = -10.7652$ Kcal/mol) (Figures 5 and 6).

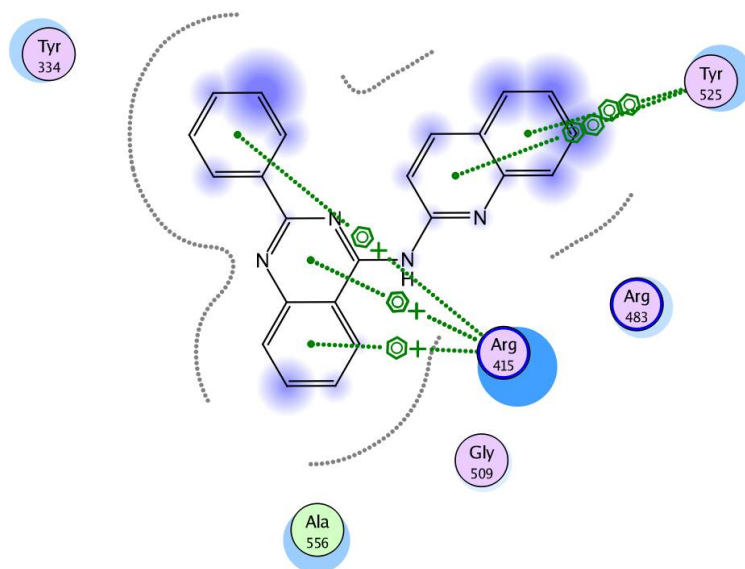


Figure 5: Interaction of compound **7** with the Kelch domain of Keap1 (PDB ID: 41QK)

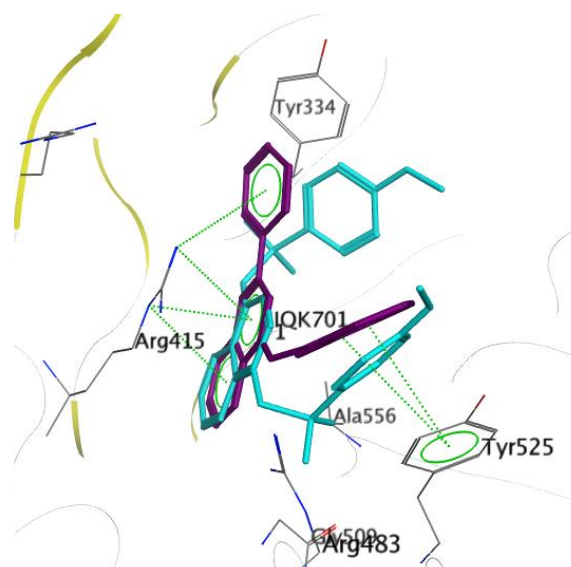


Figure 6: Overlap of Compound **7** (magenta) over native ligand (cyan) (PDB ID: 41QK)

One limitation of our study is the lack of a well-established biological assay for assessing the ability of small molecules to disrupt the Keap1-Nrf2 protein-protein interactions. Therefore, although the molecular docking study suggests that these compounds can potentially bind to Keap1 and disrupt its interaction with Nrf2, further experimental

evidence is required to confirm this possibility. Regardless of the precise mode of action in activating Nrf2, the inducer potency of compound 7 in upregulating NQO1 strongly suggests that this compound could be used as a lead for further optimization of the quinazoline scaffold.

Conclusion

In conclusion, this study presents the synthesis of *N*-substituted 2-phenylquinazolin-4-amine based derivatives **2-12** as cytoprotective NQO1 inducers. The most potent inducer in this study was the analogue **7** which caused a dose-dependent induction of NQO1 with a CD value of 0.4 μ M. In silico evaluation of these derivatives as Keap1-Nrf2 protein-protein interaction inhibitors was performed using molecular docking techniques employing MOE software version 10.2009. Derivatives were found to bind to key amino acids in the binding site with a network or arene-arene and arene-cationic interactions. These molecules represent promising quinazoline scaffold-based leads for further optimization as cytoprotective agents and for future investigations into other plausible modes of action.

Declaration of interest

The authors would like to extend their sincere appreciation to the Deanship of Scientific Research at King Saud University for funding of this research through the Research Group Project no. **RGP-VPP-302**. Maureen Higgins and Albena T. Dinkova-Kostova are also grateful to Cancer Research UK (C20953/A10270 and C20953/A18644) for financial support.

References

1. Waris G, Ahsan H. Reactive oxygen species: role in the development of cancer and various chronic condition. *J Carcinog* 2006;5:14.
2. Scandalios JG. The rise of ROS. *Trends Biochem Sci* 2002;27:483–486.

3. Klein JA, Ackerman SL. Oxidative stress, cell cycle, and neurodegeneration. *J Clin Invest* 2003;111:785-93.
4. Ahsan H, Ali A, Ali R. Oxygen free radicals and systemic autoimmunity. *Clin Exp Immunol* 2003;131:398–404.
5. Knight JA. Diseases related to oxygen-derived free radicals. *Ann Clin Lab Sci* 1995;25:111–121.
6. Waris G, Alam K. Attenuated antigenicity of ribonucleoproteins modified by reactive oxygen species. *Biochem Mol Biol Int* 1998;45:33–45.
7. Kensler TW, Wakabayashi N, Biswal S. Cell survival responses to environmental stresses via the Keap1-Nrf2-ARE pathway. *Annu Rev Pharmacol Toxicol* 2007;47:89–116.
8. Kobayashi M, Li L, Iwamoto N, et al. The antioxidant defense system Keap1-Nrf2 comprises a multiple sensing mechanism for responding to a wide range of chemical compounds. *Mol Cell Biol* 2009;29:493–502.
9. McMahon M, Lamont DJ, Beattie KA, Hayes JD. Keap1 perceives stress via three sensors for the endogenous signaling molecules nitric oxide, zinc, and alkenals. *Proc Natl Acad Sci USA* 2010;107:18838–18843.
10. Dinkova-Kostova AT, Holtzclaw WD, Cole RN, Itoh K, Wakabayashi N, Katoh Y, Yamamoto M, Talalay P. Direct evidence that sulfhydryl groups of Keap1 are the sensors regulating induction of phase 2 enzymes that protect against carcinogens and oxidants. *Proc Natl Acad Sci U S A*. 2002;99:11908-11913.
11. Li J, Ichikawa T, Janicki JS, Cui T. Targeting the Nrf2 pathway against cardiovascular disease. *Expert Opin Ther Targets* 2009;13:785–794.
12. Zhao J, Redell JB, Moore AN, Dash PK. A novel strategy to activate cytoprotective genes in the injured brain. *Biochem Biophys Res Commun* 2011;407:501–506.
13. Steel R, Cowan J, Payerne E, O'Connell MA, Searcey M. Anti-inflammatory effect of a cell-penetrating peptide targeting the Nrf2/Keap1 Interaction. *ACS Med Chem Lett* 2012;3:407-410.

14. Williamson TP, Amirahmadi S, Joshi G, Kaludov NK, Martinov MN, Johnson DA, Johnson JA. Discovery of potent, novel Nrf2 inducers via quantum modeling, virtual screening, and in vitro experimental validation *Chem Biol Drug Des* 2012;80:810-820.
15. Hu L, Magesh S, Chen L, Wang L, Lewis TA, Chen Y, Khodier C, Inoyama D, Beamer L J, Emge TJ, Shen J, Kerrigan JE, Kong AN, Dandapani S, Palmer M, Schreiber SL, Munoz B. Discovery of a small-molecule inhibitor and cellular probe of Keap1-Nrf2 protein-protein interaction. *Bioorg Med Chem Lett* 2013;23: 3039-3043.
16. Zhuang C, Miao Z, Sheng C, Zhang W. Updated research and applications of small molecule inhibitors of Keap1-Nrf2 protein-protein interaction: a review. *Curr Med Chem* 2014;21:1861-1870.
17. Nesterova NA, Kovalenko SI, Belenichev IF, Karpenkos OV, Sidorova IV. Formation of combinational library of quinazoline-4-yl-hydrazones with antioxidant activity. *Ukraine Med. Khim* 2004;6:14-21.
18. Nesterova NA, Kovalenko SI, Karpenkos OV, Belenichev IF. Synthesis and antioxidant activity of 4-arylidenehydrazinoquinazolines. *Ukr Farmatsevtichnii Zhurnal (Kiev)* 2004;22:5-10.
19. Al-Omar MA, Al-Rashood ST, El-Subbagh HI, Abdel Hamide SG. Interaction of 2-thio-4-oxo-quinazoline derivatives with guinea pig liver molybdenum hydroxylases, xanthine oxidase and aldehyde oxidase. *J Biological Sci* 2005;5:370-378.
20. Ghorab MM, Higgins M, Alsaied MS, Arafa RK, Shahat AA, Dinkova-Kostova AT. Synthesis, molecular modeling and NAD(P)H:quinone oxidoreductase 1 inducer activity of novel cyanoenone and enone benzenesulfonamides. *J Enzyme Inhib Med Chem* 2014; 29:840-845.
21. Ghorab MM, Higgins M, Dinkova-Kostova AT, Alsaied MS, Shahat AA. Novel thioureido derivatives carrying thione and sulfonamide moieties induce the cytoprotective enzyme NAD(P)H:quinone oxidoreductase 1. *Asian J Chem* 2014;26:8501-8504.

22. Prochaska HJ, Santamaria AB. Direct measurement of NAD(P)H:quinone reductase from cells cultured in microtiter wells: a screening assay for anticarcinogenic enzyme inducers. *Anal Biochem.* 1988;169:328-336.
23. Marcotte D, Zeng W, Hus JC, et al. Small molecules inhibit the interaction of Nrf2 and the Keap1 Kelch domain through a noncovalent mechanism. *Bioorg Med Chem* 2013;21:4011–4019.
24. Magesh S, Chen Y, Hu L. Small molecule modulators of Keap1- Nrf2-ARE pathway as potential preventive and therapeutic agents. *Med Res Rev* 2012;32:687–726.
25. Bertrand HC, Schaap M, Baird L, Georgakopoulos ND, Fowkes A, Thiollier C, Kachi H, Dinkova-Kostova AT, Wells G. Design, synthesis, and evaluation of triazole derivatives that induce Nrf2 dependent gene products and inhibit the Keap1-Nrf2 protein-protein Interaction. *J Med Chem.* 2015;58:7186-7194.